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Genomic analysis of a multidrug-resistant clinical *Providencia rettgeri* (PR002) strain with the novel integron *In1483* and an A/C plasmid replicon

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Whole-genome sequence analysis was performed on a multidrug-resistant *Providencia rettgeri* PR002 clinical strain isolated from the urine of a hospitalized patient in Pretoria, South Africa, in 2013. The resistome, mobilome, pathogenicity island(s), as well as virulence and heavy-metal resistance genes of the isolate, were characterized using whole-genome sequencing and bioinformatic analysis. PR002 had a genome assembly size of 4,832,624 bp with a GC content of 40.7%, an A/C₂ plasmid replicase gene, four integrons/gene cassettes, 17 resistance genes, and several virulence and heavy metal resistance genes, confirming PR002 as a human pathogen. A novel integron, *In1483*, harboring the gene *bla*_{OXA-2}, was identified, with other uncharacterized class 1 integrons harboring *aacA4cr* and *dfxA1*. *Aac(3')-IIa* and *bla*_{SCO-1}, as well as *bla*_{PER-7}, *sul2*, and *tet(B)*, were found bracketed by composite Tn3 transposons, and IS91, IS91, and IS4 family insertion sequences, respectively. PR002 was resistant to all antibiotics tested except amikacin, carbapenems, cefotaxime-clavulanate, ceftazidime-clavulanate, cefoxitin, and fosfomicin. PR002 was closely related to PR1 (USA), PRET_2032 (SPAIN), DSM_1131, and NCTC7477 clinical *P. rettgeri* strains, but not close enough to suggest it was imported into South Africa from other countries. Multidrug resistance in *P. rettgeri* is rare, particularly in clinical settings, making this case an important incident requiring urgent attention. This is also the first report of an A/C plasmid in *P. rettgeri*. The array, multiplicity, and diversity of resistance and virulence genes in this strain are concerning, necessitating stringent infection control, antibiotic stewardship, and periodic resistance surveillance/monitoring policies to preempt further horizontal and vertical spread of these resistance genes.

Keywords: *Providencia rettgeri*; IncA/C₂ plasmid; resistome; mobilome; virulence; South Africa

Introduction

Providencia rettgeri is a Gram-negative opportunistic human pathogenic bacterium that belongs to Proteae bacteria, which comprises the Morganella and Proteus genera. It is found in hospital settings and is mainly associated with urinary tract infections (UTI).¹ *P. rettgeri* has also been reported to cause other infections, such as bacteremia, eye infections, meningitis, endocarditis, pneumonia, UTI, and diarrhea.^{2,3} *P. rettgeri* is intrinsically

resistant to colistin and tigecycline,⁴ which can be transferred horizontally to *Escherichia coli*, and generally resistant to gentamycin, tobramycin, aminopenicillins, and first-generation cephalosporins.³ It is known to harbor various virulence and resistance genes, which could easily be mobilized owing to their locations on mobile genetic elements (MGEs). These MGEs, specifically plasmids, integrons, insertion sequences (ISs), and transposons, contribute significantly to its pathogenicity and resistance.^{3,5}

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Supporting information

Additional supporting information may be found in the online version of this article.

Figure S1. Prophages of *P. rettgeri* PR002 were identified using the PHAge Search Tool (PHAST). PR002 contained four intact prophages (red color), one incomplete prophage (gray color), and one questionable prophage (green color).

Table S1. Metadata (biosample data) of all sequenced *P. rettgeri* isolates from NCBI GenBank database

Table S2. Genes associated with general COG functional categories in the genome of *P. rettgeri* PR002.

Table S3. Pathogenicity islands (PAIs) predicted for PR002 from VFDB (http://www.paidb.re.kr/about_paidb.php).

Table S5. Diversity of metal resistance gene determinants found in PR002 genome.

File S1. Metadata of isolates used in phylogenomics.

Competing interests

The authors declare no competing interests.

References

1. Marquez-Ortiz, R.A. et al. 2017. Genomic epidemiology of NDM-1-encoding plasmids in Latin American clinical isolates reveals insights into the evolution of multidrug resistance. *Genome Biol. Evol.* **9**: 1725–1741.
2. Choi, H.K., Y.K. Kim, H.Y. Kim, et al. 2015. Clinical and microbiological features of *Providencia* bacteremia: experience at a tertiary care hospital. *Korean J. Intern. Med.* **30**: 219–225.
3. Abdallah, M. & A. Balshi. 2018. First literature review of carbapenem-resistant *Providencia*. *New Microbes New Infect.* **25**: 16–23.
4. Osei Sekyere, J., U. Govinden, L.A. Bester & S.Y. Essack. 2016. Colistin and tigecycline resistance in carbapenemase-producing gram negative bacteria: emerging resistance mechanisms and detection methods. *J. Appl. Microbiol.* **121**: 601–617.
5. Olaitan, A.O., S.M. Diene, M.V. Assous & J.-M. Rolain. 2016. Genomic plasticity of multidrug-resistant NDM-1 positive clinical isolate of *Providencia rettgeri*. *Genome Biol. Evol.* **8**: 723–728.
6. Vogan, A.A. & P.G. Higgs. 2011. The advantages and disadvantages of horizontal gene transfer and the emergence of the first species. *Biol. Direct* **6**: 1.
7. Pedersen, T. et al. 2018. Spread of plasmid-encoded NDM-1 and GES-5 carbapenemases among extensively drug-resistant and pandrug-resistant clinical Enterobacteriaceae in Durban, South Africa. *Antimicrob. Agents Chemother.* **62**. <https://doi.org/10.1128/AAC.02178-17>.
8. Kopotsa, K., J. Osei Sekyere & N.M. Mbelle. 2019. Plasmid evolution in carbapenemase-producing *Enterobacteriaceae*: a review. *Ann. N.Y. Acad. Sci.* **1457**: 61–91.
9. Stokes, H.W. & M.R. Gillings. 2011. Gene flow, mobile genetic elements and the recruitment of antibiotic resistance genes into Gram-negative pathogens. *FEMS Microbiol. Rev.* **35**: 790–819.
10. Lynch, T., A. Petkau, N. Knox, et al. 2016. A primer on infectious disease bacterial genomics. *Clin. Rev. Microbiol.* **29**: 881–913.
11. Aminov, R.I. 2011. Horizontal gene exchange in environmental microbiota. *Front. Microbiol.* **2**: 1–19.
12. Engering, A., L. Hogerwerf & J. Slingenberg. 2013. Pathogen—host—environment interplay and disease emergence. *Emerg. Microbes Infect.* **2**: 1–7.
13. Osei Sekyere, J. 2019. Mcr colistin resistance gene: a systematic review of current diagnostics and detection methods. *Microbiologyopen* **8**: e00682.
14. Clinical and Laboratory Standards Institute (CLSI). 2017. Performance standards for antimicrobial susceptibility testing. In *27th Informational Supplement M100-S27*, CLSI, Wayne, PA.
15. European Committee on Antimicrobial Susceptibility Testing. 2016. Breakpoint tables for interpretation of MICs and zone diameters. Version 6.0. 1–78. Last accessed: August 1, 2019. <http://www.eucast.org>.
16. Bolger, A.M., M. Lohse & B. Usadel. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **30**: 2114–2120.
17. Bankevich, A. et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J. Comput. Biol.* **19**: 455–477.
18. Tatusova, T. et al. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res.* **44**: 6614–6624.
19. Aziz, R.K. et al. 2008. The RAST Server: rapid annotations using subsystems technology. *BMC Genomics* **9**: 75.
20. Benson, G. 1999. Tandem repeats finder: a program to analyze DNA sequences. *Nucleic Acids Res.* **27**: 573–580.
21. Grissa, I., G. Vergnaud & C. Pourcel. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res.* **35**: 52–57.
22. Larsen, M.V. et al. 2014. Benchmarking of methods for genomic taxonomy. *J. Clin. Microbiol.* **52**: 1529–1539.
23. Thomsen, M.C.F. et al. 2016. A bacterial analysis platform: an integrated system for analysing bacterial whole genome

- sequencing data for clinical diagnostics and surveillance. *PLoS One* **11**: e0157718.
24. Zankari, E. *et al.* 2012. Identification of acquired antimicrobial resistance genes. *J. Antimicrob. Chemother.* **67**: 2640–2644.
 25. Siguier, P., J. Perochon, L. Lestrade, *et al.* 2006. ISfinder: the reference centre for bacterial insertion sequences. *Nucleic Acids Res.* **34**: D32–D36.
 26. Carattoli, A. *et al.* 2014. In silico detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob. Agents Chemother.* **58**: 3895–3903.
 27. Osei Sekyere, J. & D.G. Amoako. 2017. Genomic and phenotypic characterisation of fluoroquinolone resistance mechanisms in Enterobacteriaceae in Durban, South Africa. *PLoS One* **12**: e0178888.
 28. Moura, A. *et al.* 2009. INTEGRALL: a database and search engine for integrons, integrases and gene cassettes. *Bioinformatics* **25**: 1096–1098.
 29. Zhou, Y., Y. Liang, K.H. Lynch, *et al.* 2011. PHAST: a Fast Phage Search Tool. *Nucleic Acids Res.* **39**: 347–352.
 30. Cosentino, S., M. Voldby Larsen, F. Møller Aarestrup & O. Lund. 2013. PathogenFinder—distinguishing friend from foe using bacterial whole genome sequence data. *PLoS One* **8**: e77302.
 31. Chen, L. *et al.* 2005. VFDB: a reference database for bacterial virulence factors. *Nucleic Acids Res.* **33**: 325–328.
 32. Treangen, T.J., B.D. Ondov, S. Koren & A.M. Phillippy. 2014. The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. *Genome Biol.* **15**: 524.
 33. Jacoby, G.A. 2009. AmpC beta-lactamases. *Clin. Microbiol. Rev.* **22**: 161–182, Table of Contents.
 34. Osei Sekyere, J. & D.G. Amoako. 2017. Carbonyl cyanide m-chlorophenylhydrazine (CCCP) reverses resistance to colistin, but not to carbapenems and tigecycline in multidrug-resistant Enterobacteriaceae. *Front. Microbiol.* **8**: 228.
 35. Carattoli, A., L. Villa, L. Poirel, *et al.* 2012. Evolution of IncA/C blaCMY₂-carrying plasmids by acquisition of the blaNDM₁ carbapenemase gene. *Antimicrob. Agents Chemother.* **56**: 783–786.
 36. Manageiro, V. *et al.* 2015. Draft genome sequence of the first NDM-1-producing *Providencia stuartii* strain isolated in Portugal. *Genome Announc.* **3**. <http://doi.org/10.1128/genomeA.01077-15>.
 37. Wailan, A.M., D.L. Paterson, M. Caffery, *et al.* 2015. Draft genome sequence of NDM-5-producing *Escherichia coli* sequence type 648 and genetic context of blaNDM-5 in Australia. *Genome Announc.* **3**. <http://doi.org/10.1128/genomeA.00194-15>.
 38. Arpin, C. *et al.* 2012. Evolution of an incompatibility group IncA/C plasmid harboring blaCMY-16 and qnrA6 genes and its transfer through three clones of *Providencia stuartii* during a two-year outbreak in a Tunisian burn unit. *Antimicrob. Agents Chemother.* **56**: 1342–1349.
 39. Fernández-Alarcón, C., R.S. Singer & T.J. Johnson. 2011. Comparative genomics of multidrug resistance-encoding IncA/C plasmids from commensal and pathogenic *Escherichia coli* from multiple animal sources. *PLoS One* **6**: e23415.
 40. Martínez-Freije, P. *et al.* 1998. Class I integrons in Gram-negative isolates from different European hospitals and association with decreased susceptibility to multiple antibiotic compounds. *J. Antimicrob. Chemother.* **42**: 689–696.
 41. Penadés, J.R., J. Chen, N. Quiles-Puchalt, *et al.* 2015. Bacteriophage-mediated spread of bacterial virulence genes. *Curr. Opin. Microbiol.* **23**: 171–178.
 42. Haiko, J. & B. Westerlund-Wikström. 2013. The role of the bacterial flagellum in adhesion and virulence. *Biology (Basel)* **2**: 1242–1267.
 43. Coburn, B., I. Sekirov & B.B. Finlay. 2007. Type III secretion systems and disease. *Clin. Microbiol. Rev.* **20**: 535–549.
 44. Bouchet, V. *et al.* 2003. Host-derived sialic acid is incorporated into *Haemophilus influenzae* lipopolysaccharide and is a major virulence factor in experimental otitis media. *Proc. Natl. Acad. Sci. USA* **100**: 8898–8903.
 45. Garduño, R.A., E. Garduño & P.S. Hoffman. 1998. Surface-associated Hsp60 chaperonin of *Legionella pneumophila* mediates invasion in a HeLa cell model. *Infect. Immun.* **66**: 4602–4610.
 46. Birben, E. *et al.* 2012. Oxidative stress and antioxidant defense. *World Allergy Organ. J.* **5**: 9–19.
 47. Li, Y.H. & X.L. Tian. 2016. Quorum sensing and bacterial social interactions in biofilms: bacterial cooperation and competition. *Stress Environ. Regul. Gene Expr. Adapt. Bact.* **2**: 1197–1205.
 48. Beceiro, A. *et al.* 2014. Biological cost of different mechanisms of colistin resistance and their impact on virulence in *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* **58**: 518–526.
 49. Kale, S.P., D. Salaskar, S. Ghosh & S. Sounderajan. 2015. Isolation and identification of arsenic resistant *Providencia rettgeri* (KDM3) from industrial effluent contaminated soil and studies on its arsenic resistance mechanism. *J. Microb. Biochem. Technol.* **7**: 1–8.
 50. Yamina, B., B. Tahar, M. Lila, *et al.* 2014. Study on cadmium resistant-bacteria isolated from hospital wastewaters. *Adv. Biosci. Biotechnol.* **5**: 718–726.
 51. Di Cesare, A. *et al.* 2016. Co-occurrence of integrase 1, antibiotic and heavy metal resistance genes in municipal wastewater treatment plants. *Water Res.* **94**: 208–214.
 52. Summers, A.O. *et al.* 1993. Mercury released from dental silver fillings provokes an increase in mercury- and antibiotic-resistant bacteria in oral and intestinal floras of primates. *Antimicrob. Agents Chemother.* **37**: 825–834.
 53. Baker-Austin, C., M.S. Wright, R. Stepanauskas & J.V. McArthur. 2006. Co-selection of antibiotic and metal resistance. *Trends Microbiol.* **14**: 176–182.
 54. Dobrindt, U. & J. Hacker. 2001. Whole genome plasticity in pathogenic bacteria. *Curr. Opin. Microbiol.* **4**: 550–557.
 55. Partridge, S.R., S.M. Kwong, N. Firth & S.O. Jensen. 2018. Mobile genetic elements associated with antimicrobial resistance. *Clin. Microbiol. Rev.* **31**: 1–61.