MITOCHONDRIAL TOXICITIES, BODY-FAT ABNORMALITIES AND THE POSSIBLE ASSOCIATED CHANGE IN CARDIOVASCULAR RISK OF HIGHLY ACTIVE ANTI-RETROVIRAL THERAPY IN HIV-INFECTED INDIVIDUALS - A SOUTH AFRICAN PERSPECTIVE.

COLIN NIGEL MENEZES

A thesis submitted to the Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa, for the degree of Doctor of Philosophy.
DECLARATION

I declare that this thesis is my own work. It is being submitted for the degree of Doctor of Philosophy to the University of the Witwatersrand, Johannesburg.

This thesis is submitted in the optional format, approved by the faculty, of published work with a supporting introduction, as a literature review, and discussion.

This work has not been submitted before for any degree or examination at any other University.

........................................

Colin Nigel Menezes
DEDICATION

This work is dedicated to the memory of my father, Jose Francisco da Piedade Menezes and to my mother, Maria Hazel de Quadros Menezes for her constant support and encouragement.
ABSTRACT

Despite the improved survival of human immunodeficiency virus (HIV) infected individuals with the introduction of highly active anti-retroviral therapy (HAART) in the South African public sector in 2004, new challenges have been brought to the fore. These include drug-related toxicities, particular those of stavudine, which remains in common use within developing countries.

A prospective analysis of 9040 HIV-1-infected adults initiated on HAART from 2004 to 2007 at the Themba Lethu Clinic, Helen Joseph Hospital in Johannesburg, confirmed the ability to roll out a successful HAART programme in a resource limited environment with a high retention rate of 70%. Nearly 30% of patients switched to non-stavudine based regimens due to side effects - predominantly peripheral neuropathy, symptomatic hyperlactataemia and lipoatrophy.

In an attempt to look for safer options, a prospective randomized controlled trial comparing standard and low dose stavudine with tenofovir was undertaken in 2009. Sixty patients were randomized 1:1:1 to either standard (30-40 mg), low (20-30 mg) dose stavudine or tenofovir (300 mg) each combined with lamivudine and efavirenz. Adipocyte mitochondrial DNA (mtDNA) levels, gene expression, anthropometry, markers of inflammation, lipid and glucose metabolism were assessed at various time intervals.

Results demonstrate early mitochondrial depletion among black South African patients receiving low and standard doses of stavudine, with preservation of gene expression levels, except for NRF1 and MTCYB, when compared to patients on tenofovir. Mitochondrial toxicities occurred in both the stavudine arms. Immunological and virological outcomes were similar for all three arms. Both drugs caused lipid changes, but tenofovir had a more
favourable effect on anthropometry and adipokines. Both stavudine regimens increased fasting insulin and C-peptide levels, with the higher stavudine dose also causing increased fasting glucose and HOMA levels.

This study demonstrates an early association between mitochondrial depletion and stavudine therapy in the black South African population and shows that tenofovir has a minimal effect on mitochondrial numbers. Only two of eight adipocyte genes were significantly affected by stavudine therapy when compared with tenofovir, but this was only seen with the standard dose. This study highlights the occurrence of significant metabolic abnormalities with both drugs. Therefore, awareness of the potential increased cardiovascular risk should be of concern with tenofovir and stavudine, although toxicity is lower in the low dose compared to the standard dose stavudine regimen with no attenuation of anti-retroviral effectivity.
ACKNOWLEDGEMENTS

I would like to thank my supervisors, Professors Nigel Crowther, Frederick Raal and Ian Sanne for guiding me through the process of this work.

I would also like to thank Dr Alastair Moodley, Prof Sarala Naicker, Dr Raquel Duarte, Dr Caroline Dickens, Ms Therese Dix-Peek, Mrs Desiree Van Amsterdam, Dr Mhairi Maskew, Dr Denise Evans, Dr Melanie-Anne John, Dr Pru Ive, Mrs Mellissa Hero, Mrs Marlene Naidoo, Professor Patrick MacPhail, Dr Gajendra Chita, Prof Ken Huddle, Dr Alison Bentley and Ms Jean Johnstone who made suggestions, comments or helped me with various issues during this project.

I would to thank Emma Hammond from the Centre for Clinical Immunology and Biomedical Statistics, Royal Perth Hospital and Murdoch University, Australia, for providing us with the methodology, primer sequences and standard information for the mitochondrial DNA copy number determination.

I am grateful to the patients who participated in this study and to the staff of the Themba Lethu Clinic, Clinical HIV Research Unit and the Departments of Internal Medicine and Radiology at Helen Joseph Hospital.

I would like to acknowledge CIPLA SA for their donation of some the drugs for the duration of this study.
I would also like to acknowledge the Clinical HIV Research Unit, the Faculty of Health Sciences, University of the Witwatersrand, and the National Health Laboratory Service for their funding for this project.

Lastly, I would like to thank my family and friends for all their support and never doubting my ability to finish this PhD project.
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LIST OF OUTCOMES OF THIS WORK

Publications:

Paper 1:
A longitudinal study of stavudine-associated toxicities in a large cohort of South African HIV infected subjects. BMC Infect Dis. 2011.11:244

Paper 2:

Paper 3:

Poster presentation:

Wits Research Day, September 2012, Johannesburg, South Africa.

**Conference presentation:**


Metabolic complications of HIV and HAART: The hyperlactataemia syndromes
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<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>$2^{\Delta\text{Cq}}$</td>
<td>Relative quantities.</td>
</tr>
<tr>
<td>ABC</td>
<td>Abacavir.</td>
</tr>
<tr>
<td>AE</td>
<td>Adverse event.</td>
</tr>
<tr>
<td>AIDS</td>
<td>Acquired immunodeficiency syndrome.</td>
</tr>
<tr>
<td>ALT</td>
<td>Alanine aminotransferase.</td>
</tr>
<tr>
<td>ANOVA</td>
<td>Analysis of variance.</td>
</tr>
<tr>
<td>AST</td>
<td>Aspartate aminotransferase.</td>
</tr>
<tr>
<td>ATV</td>
<td>Atazanavir.</td>
</tr>
<tr>
<td>AZT</td>
<td>Zidovudine.</td>
</tr>
<tr>
<td>B2M</td>
<td>$\beta$2 microglobulin.</td>
</tr>
<tr>
<td>BMI</td>
<td>Body mass index.</td>
</tr>
<tr>
<td>bp</td>
<td>Base pair.</td>
</tr>
<tr>
<td>CCR5</td>
<td>Chemokine (C-C motif) receptor 5.</td>
</tr>
<tr>
<td>CD4 count</td>
<td>Cluster of differentiation 4 count.</td>
</tr>
<tr>
<td>CI</td>
<td>Confidence interval.</td>
</tr>
<tr>
<td>COX 3</td>
<td>Cytochrome c oxidase subunit III.</td>
</tr>
<tr>
<td>COX 4</td>
<td>Cytochrome c oxidase subunit IV.</td>
</tr>
<tr>
<td>CPK</td>
<td>Creatinine kinase.</td>
</tr>
<tr>
<td>$\Delta\text{Cq}$</td>
<td>Quantification cycle.</td>
</tr>
<tr>
<td>CRABP1</td>
<td>Cellular retinoic acid binding protein 1.</td>
</tr>
<tr>
<td>CT imaging</td>
<td>Computerised tomography imaging.</td>
</tr>
<tr>
<td>CVD</td>
<td>Cardiovascular disease.</td>
</tr>
<tr>
<td>CXCR4</td>
<td>Chemokine (C-X-C motif) receptor 4.</td>
</tr>
</tbody>
</table>
DAD Study  | Data collection on Adverse Effects of Anti-HIV Drugs Study.
DDI       | Didanosine.
DEXA      | Dual-energy radiographic absorptiometry.
DM        | Diabetes mellitus.
dNTP      | Deoxynucleoside triphosphate.
DRV       | Darunavir.
DSMB      | Data safety and monitoring board.
d4T       | Stavudine.
ECG       | Electrocardiogram.
EFV       | Efavirenz.
ELISA     | Enzyme-linked immunosorbent assay.
ETV       | Etravirine.
FTC       | Emtricitabine.
GFR       | Cockcroft-Gault formula.
GGT       | Gamma-glutamyltransferase.
GLUT 4    | Glucose transporter type 4.
HAART     | Highly active anti-retroviral therapy.
HDL cholesterol | High-density lipoprotein cholesterol.
HGH       | Human growth hormone
HIV       | Human immunodeficiency virus.
HOMA      | Homeostasis model assessment.
HPRT      | Hypoxanthine-guanine phosphoribosyltransferase.
hs-CRP    | High sensitivity C-reactive protein.
ICAM-1    | Intercellular Adhesion Molecule -1.
<table>
<thead>
<tr>
<th>Abbreviation (IFG, IGT, IL, IQR, LA, LD, LDL cholesterol, LEP, LPL, LPV, LRP1, MCC, MRI, MTCYB, mtDNA, MVC, ND, NNRTI, NRF1, NRTI, Nts, NVP, PGC-1α, PI)</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>IFG</td>
<td>Impaired fasting glucose.</td>
</tr>
<tr>
<td>IGT</td>
<td>Impaired glucose tolerance.</td>
</tr>
<tr>
<td>IL</td>
<td>Interleukin.</td>
</tr>
<tr>
<td>IQR</td>
<td>Interquartile range.</td>
</tr>
<tr>
<td>LA</td>
<td>Lactic acidosis.</td>
</tr>
<tr>
<td>LD</td>
<td>Lipodystrophy.</td>
</tr>
<tr>
<td>LDL cholesterol</td>
<td>Low-density lipoprotein cholesterol.</td>
</tr>
<tr>
<td>LEP</td>
<td>Leptin.</td>
</tr>
<tr>
<td>LPL</td>
<td>Lipoprotein lipase.</td>
</tr>
<tr>
<td>LPV</td>
<td>Lopinavir.</td>
</tr>
<tr>
<td>LRP1</td>
<td>Low density lipoprotein receptor related protein 1.</td>
</tr>
<tr>
<td>MCC</td>
<td>Medicines Control Council.</td>
</tr>
<tr>
<td>MRI</td>
<td>Magnetic resonance imaging.</td>
</tr>
<tr>
<td>MTCYB</td>
<td>Mitochondrial cytochrome B.</td>
</tr>
<tr>
<td>mtDNA</td>
<td>Mitochondrial DNA.</td>
</tr>
<tr>
<td>MVC</td>
<td>Maraviroc.</td>
</tr>
<tr>
<td>ND</td>
<td>Not done.</td>
</tr>
<tr>
<td>NNRTI</td>
<td>Non-nucleoside reverse transcriptase inhibitor.</td>
</tr>
<tr>
<td>NRF1</td>
<td>Nuclear respiratory factor-1.</td>
</tr>
<tr>
<td>NRTI</td>
<td>Nucleoside reverse transcriptase inhibitor.</td>
</tr>
<tr>
<td>Nts</td>
<td>Nucleotides.</td>
</tr>
<tr>
<td>NVP</td>
<td>Nevirapine.</td>
</tr>
<tr>
<td>PGC-1α</td>
<td>PPAR-γ coactivator 1α.</td>
</tr>
<tr>
<td>PI</td>
<td>Protease inhibitor.</td>
</tr>
</tbody>
</table>
PN  Peripheral neuropathy.
PPARγ  Peroxisome-proliferator activated receptor γ.
qPCR  Quantitative polymerase chain reaction.
RAL  Raltegravir.
RNA RPL 13A  60S ribosomal protein L 13A.
RTV  Ritonavir.
RXR-PPARγ  Retinoid X receptor-peroxisome proliferator-activated receptor γ.
SH  Symptomatic hyperlactatemia.
SMART  Strategies for management of antiretroviral therapy.
SQV  Saquinavir.
SREBP-1  Sterol regulatory element binding protein-1.
TDF  Tenofovir.
TFAM  Mitochondrial transcription factor-A.
TNF  Tumour necrosis factor.
T20  Enfuvirtide.
3TC  Lamivudine.
VCAM-1  Vascular cell adhesion molecule-1.
vWF  von Willebrand factor.
WHO  World Health Organization.
PREFACE

I formulated the study concept and design with Professor Ian Sanne, which was overseen by my supervisors, Professors Nigel Crowther, Frederick Raal and Ian Sanne. Assistance was received with the data collection and management from Mrs Desiree Van Amsterdam, Dr Mhairi Maskew, Ms Doreen Schulze and Mrs Avril Swarts Prinsloo. Dr Raquel Duarte, Dr Caroline Dickens and Ms Therese Dix-Peek assisted with the laboratory work. Professor Nigel Crowther, Dr Mhairi Maskew and Dr Denise Evans assisted with the data analysis. Mrs Marlene Naidoo assisted with the regulatory work. Dr Melanie-Anne John, Dr Francesca Condradie, Dr Mohammed Rassol, Dr Sharlaa Badal – Faesen, Dr Faizel Laher and Dr Pru Ive assisted with patient management.

This thesis is divided into five chapters:

**Chapter 1** provides a background on human immunodeficiency virus (HIV)-1 infection and the issues around the management of HIV-infected individuals, especially the complications associated with highly active anti-retroviral therapy (HAART), and introduces the aims of the thesis. **Chapter 2** contains the results of a published prospective analysis of a large cohort of HIV-1-infected adults initiated on HAART. It discusses the issues related to use of stavudine as first line therapy and forms the basis for the clinical trial using tenofovir as an option. The published version of this paper is included in Appendix 1. **Chapter 3** presents a published paper of the clinical trial and discusses the early effects of low and standard stavudine therapy when compared to tenofovir, on adipocyte gene expression, mitochondrial DNA copy number and metabolic parameters in South African HIV-infected patients after four weeks of therapy. The published version of this paper is included in Appendix 2.
Chapter 4 contains the follow up paper (published after submission) on the longer term metabolic parameters after 48 weeks of therapy in this same cohort of patients. The published version of this paper is included in Appendix 3. Chapter 5 summarises the results of the project and discusses new directions for research in this field. Each chapter has a list of its own references.
CHAPTER 1

1. INTRODUCTION AND LITERATURE REVIEW

1.1. The human immunodeficiency virus (HIV)-1 infection and its natural history

HIV-1 infection is known to be transmitted through unprotected sexual intercourse, contaminated needles, blood products or through mother-to-child transmission. The HIV-1 binds to CD4+ T cells which include T-helper cells, monocytes/macrophages, eosinophils, microglial cells and dendritic cells (Fanales-Belasio et al, 2010).

The virus enters the cell by attaching to the CD4 receptor and its co-receptor, either chemokine (C-C motif) receptor 5 (CCR5) or chemokine (C-X-C motif) receptor 4 (CXCR4) on the surface of the cell, releasing its RNA and its various enzymes, which include the reverse transcriptase, the integrase enzyme, and the protease enzyme into the cell. Using the reverse transcriptase enzyme, a single-strand RNA genome is then transcribed into a double-strand DNA, and is integrated into a host chromosome. This viral DNA is then transported into the cell nucleus, where its integration into the cell's genome is carried out by the integrase enzyme. Using the host’s nuclear transcription processes, viral proteins are produced which are cleaved into active forms by the protease enzyme and are packaged into new virions ready to infect more cells (Fanales-Belasio et al, 2010).

The natural history of HIV disease, which depends on the underlying host immune response and viral virulence factors, determines the outcome of the infection (Fanales-Belasio et al, 2010). If not treated, most patients will develop the acquired immunodeficiency syndrome (AIDS) which may result in an opportunistic infection or a malignancy.
1.2. The classes of antiretroviral drugs available

There are five classes of antiretroviral drugs available. These include the nucleoside reverse transcriptase inhibitors (NRTIs), which are analogues that mimic normal building blocks of DNA, preventing transcription of viral RNA to DNA; non-nucleoside viral reverse transcriptase inhibitor (NNRTIs), which fit into the genomic binding site of reverse transcriptase and directly inhibit its action; protease inhibitors (PI) which inhibit the viral protease enzyme, therefore inhibiting the final maturation stages of HIV replication resulting in non-infective viral particles; entry inhibitors that bind to the viral gp41, the host cell CD4 or chemokine (CCR5) receptors and block the entry of the HIV into the host cell; and integrase inhibitors that block the viral DNA integration into the host cell genome (Figure 1).

Only some of these drug classes (Table 1) are currently available for use in the South African public sector, these include the NRTIs, NNRTIs, PIs and more recently the integrase inhibitors (Southern African HIV Clinicians Society Guidelines for Antiretroviral Therapy in Adults, 2012).

1.3. HIV variability and response to therapy

There are two forms of HIV, HIV-1 and HIV-2. In addition, the HIV-1 is further divided into three groups, M (Major), O (Outlier) and N (non-M/non-O). The group M includes nine subtypes or clades, designed A to K. There are at least 6 sub-subtypes within clades A and F (A1, A2, A3 and A4, and F1 and F2) (Fanales-Belasio et al, 2010).

It is possible that viruses of different subtypes can infect the same cell, sharing their genetic material, resulting in hybrids of new viruses. As a result, viruses within the same HIV-1 subtype may differ by up to 20%. In sub-Saharan Africa, there is a wide HIV-1 heterogeneity but subtypes A, C and D are the most prevalent, with subtype B viruses being rare but

2
prevalent in North and South America, Central and Western Europe, and Australia. HIV-2 is endemic in West Africa, but has spread to Europe and to India (Fanales-Belasio et al., 2010). Therefore, this might affect pathogenesis, infectivity and response to antiretroviral therapy with different HIV subtypes having different susceptibilities to antiretroviral drugs (Kaleebu et al., 2001. Velazquez-Campoy et al., 2001. Fanales-Belasio et al., 2010). Importantly, however, other reasons need to be considered, such as differences in resistance patterns to antiretroviral therapy at the time of initiation of therapy, differences in pharmacokinetics and adherence issues (van den Berg et al., 2005). A study in non-B HIV-1 subtypes noted poorer
### Table 1: List of classes of antiretroviral drugs available in Southern Africa (Adapted from the Southern African HIV Clinicians Society Guidelines for Antiretroviral Therapy in Adults, 2012).

<table>
<thead>
<tr>
<th>Name of the class of antiretroviral drugs</th>
<th>Name of the available drugs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nucleoside reverse transcriptase inhibitors (NRTIs)</td>
<td>Stavudine (d4T)</td>
</tr>
<tr>
<td></td>
<td>Lamivudine (3TC)</td>
</tr>
<tr>
<td></td>
<td>Zidovudine (AZT)</td>
</tr>
<tr>
<td></td>
<td>Didanosine (DDI)</td>
</tr>
<tr>
<td></td>
<td>Tenofovir disoproxil fumarate (TDF)</td>
</tr>
<tr>
<td></td>
<td>Emtricitabine (FTC)*</td>
</tr>
<tr>
<td>Non-nucleoside viral reverse transcriptase inhibitor (NNRTIs)</td>
<td>Efavirenz (EFV)</td>
</tr>
<tr>
<td></td>
<td>Nevirapine (NVP)</td>
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<tr>
<td></td>
<td>Etravirine (ETV)</td>
</tr>
<tr>
<td>Protease inhibitor (PI)</td>
<td>Ritonavir (RTV)</td>
</tr>
<tr>
<td></td>
<td>Lopinavir (LPV)**</td>
</tr>
<tr>
<td></td>
<td>Atazanavir (ATV)</td>
</tr>
<tr>
<td></td>
<td>Saquinavir (SQV)</td>
</tr>
<tr>
<td></td>
<td>Darunavir (DRV)</td>
</tr>
<tr>
<td>Integrase inhibitors</td>
<td>Raltegravir (RAL)</td>
</tr>
</tbody>
</table>

NB: Entry inhibitors - enfuvirtide (T20) and maraviroc (MVC) are not available.
* available only as a combination drug with tenofovir.
** available only as a combination drug with ritonavir.

Virological outcome on PIs (Perno et al, 2001). Conversely, other studies have suggested that there are no differences in outcomes to tenofovir, adeovir and zidovudine for subtypes A–G (Palmer et al, 2001), and between subtypes F and A (Caride et al, 2000), or between B and C (Shafer et al, 1997) for NRTIs and NNRTIs. In two studies where African were compared to European HIV -1 infected patients, on various regimens of antiretroviral drugs, the one study (van den Berg et al, 2005) found an early poor virological response whilst the other study (Frater et al, 2002) found poorer longer term response in the African group but this was thought to be because of lack of adherence to therapy.
1.4. The burden of HIV-1 infection

According to the World Health Organization (WHO), by 2011, more than 60 million people have been infected with HIV since the beginning of the HIV-1 epidemic and nearly 30 million people have died of AIDS. In 2010, there were nearly 34 million people living with the HIV-1 infection, with an estimated 2.7 million new infections, and 1.8 million AIDS-related deaths. Sub-Saharan Africa continues to bear the global burden of HIV-1 with an estimated 68% of the HIV-1 infections worldwide. This region also accounted for 69% of the world’s AIDS related deaths by that time (WHO, 2011).

More importantly, despite having the highest rates of this epidemic, there has been a noticeable drop in new HIV-1 infections in South Africa, this by at least 41%, between 2001 and 2011 (UNAIDS, 2012). This may be because of the preventative programs, and the rapid scale up of highly active antiretroviral therapy (HAART) by the South African Government.

1.5. Access to antiretroviral therapy

A record 2.3 million people have been added to the anti-HIV treatment programs in the last two years in sub-Saharan Africa and this was an increase of 59%. South Africa has scaled up its treatment services to reach 1.7 million people which was an increase of nearly 75% in the preceding two years (UNAIDS, 2012).

The South African Government rolled out its HAART programme in the public health sector in 2004 (National Antiretroviral Treatment Guidelines, 2004), offering two regimens in the public sector. These contain a triple drug course, which consisted of 2 NRTIs, with either a NNRTI or PI. At diagnosis, unless contraindicated, patients were put on a first line regimen which consisted of stavudine, lamivudine and either efavirenz or nevirapine, and switched to second line therapy if they had virological failure to first line therapy. The second line
therapy consisted of zidovudine, didanosine and ritonavir/lopinavir. These drugs were cheap and easy to administer in the short term but were associated with more side effects in the long term. At that time, HAART was recommended when the CD4 count was 200 cells/µl or less, or when the patient presented at stage 4 of the WHO clinical staging system (National Antiretroviral Treatment Guidelines, 2004).

1.6. **The issue of side effects of stavudine therapy: mitochondrial toxicities, body fat abnormalities and cardiovascular risk**

The success of controlling the HIV-1 infection with HAART has been affected by significant acute and long term complications. Acute complications are noted to occur frequently because of the use of stavudine. The hyperlactatemia syndromes (symptomatic hyperlactatemia and lactic acidosis) are the most serious side effects of stavudine use and are related to mitochondrial toxicity with high mortality rates (Bolhaar et al, 2007. Hernandez Perez et al, 2010).

Chronic complications include lipodystrophy, dyslipidaemias, insulin resistance, and the development of new onset diabetes mellitus (De Wit et al, 2008). These metabolic abnormalities, but not necessarily the lipodystrophy, result in an increased risk for cardiovascular disease (Fourie et al, 2011).

In 2010, the WHO changed its guidelines, advocating an earlier introduction of HAART at CD4 counts of 350 cells/µl or less and the use of less toxic drugs such as tenofovir and zidovudine as first line therapy, rather than stavudine (WHO Antiretroviral therapy guidelines, 2010 revision). In April that year, South Africa followed suit, and changed its guidelines to the use of tenofovir as first line therapy instead of stavudine (National Department of Health South Africa HAART guidelines, 2010). Tenofovir is a nucleotide
analogue, and was approved for use in 2001 in the United States. It is commonly used in initial therapy and has been shown to have a favourable lipid when compared to stavudine, with no difference in virological responses (Gallant et al, 2004). Despite the WHO advising countries to change their HAART guidelines, it is known that up to 56% of HIV-1-positive patients in other low and middle income countries are still receiving stavudine as first line therapy (WHO 2010) mainly because of its low cost and its availability in fixed drug combinations.

1.6.1. The hyperlactatemia syndromes (symptomatic hyperlactatemia and lactic acidosis)

1.6.1.1. Epidemiology

The hyperlactataemia syndromes are associated with the use of NRTIs. Asymptomatic hyperlactataemia is common but does not predict for the symptomatic form of the disease. The symptomatic form of hyperlactataemia may have a good prognosis if recognised early and there is no associated liver dysfunction. Lactic acidosis is present if there is an associated metabolic acidosis and leads to multiple organ dysfunction. The first case reports were published in the early 1990s (Freiman et al, 1993. Chattha et al, 1993) and described a rare but serious disease evidenced by a lactic acidosis, hepatic steatosis and occasionally liver failure (Chariot et al, 1999. Bissuel et al, 1994. Johri et al, 2000). Incidence rates of lactic acidosis from cohorts in Africa were much higher (Table 2) compared to studies in the West which have moved away from using stavudine and didanosine. Mortality rates were as high as 30% in one South African cohort (Bolhaar et al, 2007). In another South African study, substitutions of stavudine due to symptomatic hyperlactatemia occurred between six months and 18 months on treatment with a cumulative
incidence of 4.7% after three years on HAART, with women seven times more affected than men, after being on more than six months of therapy (Boulle et al., 2007).

1.6.1.2. Pathogenesis

Mitochondria are found in all cells of the body except for erythrocytes (Maagaard et al., 2009) and because the mitochondria lack enzymes for DNA repair, they are particularly susceptible to mutagenic agents (Graziewicz et al., 2006). Mitochondrial DNA are responsible for the production of 13 mitochondrial polypeptides that are involved in the respiratory chain and essential for oxidative phosphorylation (Kakuda et al., 2000).

The mechanism by which NRTIs cause mitochondrial dysfunction is by inhibiting mtDNA polymerase γ with subsequent mtDNA depletion. In addition, NRTIs are also known to inhibit other specific enzymes involved in the tricarboxylic acid cycle and electron transport chain as well as inhibition of mitochondrial adenylate kinase and the ADP/ATP translocator (Moyle et al., 2000. Barile et al., 1994. Barile et al., 1997, 1998). Early effects of mitochondrial toxicity include decreased energy production and increased production of lactate, by affecting oxidative phosphorylation (Kakuda et al., 2000. Montaner et al., 2004). This would result in an increase in electron leakage from the electron-transport chain, which in turn increases the production of reactive oxygen species leading to a cascade of further oxidative damage and lipid peroxidation (Kakuda et al., 2000. Montaner et al., 2004).

In the presence of mitochondrial dysfunction, lactic acidosis (with or without an elevated anion gap and altered bicarbonate) may occur as aerobic respiration (oxidative phosphorylation) shifts to anaerobic respiration (glycolysis) (Kakuda et al., 2000 Montaner et al., 2004).
1.6.1.3. Management

Early recognition and intervention is vital because lactic acidosis can progress to liver failure and death. It is a diagnosis of exclusion; other conditions need to be excluded such as diabetic ketoacidosis, renal failure, sepsis, heart failure, pancreatitis, severe anaemia, liver failure, bowel ischaemia and other drugs (e.g. metformin, INH overdose) (Southern African HIV Clinicians Society, 2012). The mainstay of management includes withdrawal of all NRTIs even before the diagnosis is confirmed if there is a high index of suspicion.

The Southern African HIV clinician society guidelines are based on other published guidelines (Schambelan et al, 2002. British HIV Association (BHIVA) guidelines, 2005.). Patients with mild asymptomatic hyperlactataemia (lactates of less than 5·0 mmol/L) would include cautious continuation of NRTIs which are less “mitochondrial toxic” while monitoring the patient closely for the development of symptoms and/or further increases in lactate (Southern African HIV Clinicians Society, 2012).

Patients who have serum lactate concentrations of /or above 5·0 mmol/L whether asymptomatic or symptomatic warrant discontinuation of therapy. Administration of intravenous fluids to pre-empt cardiovascular collapse and assist hepatic and renal clearance of lactate is important. The use of intravenous sodium bicarbonate is controversial as it may trigger respiratory acidosis. Ventilation may be required if respiratory fatigue occurs. Dialysis, inotropes and other supportive measures may be required as necessary. Other therapies such as thiamine, carnitine, riboflavin, co-enzyme Q and vitamin C which are either cofactors in oxidative phosphorylation or antioxidants may also be used (Brinkman et al, 2001. John et al, 2002. Falco et al, 2002. Southern African HIV Clinicians Society, 2012).

The use of NRTIs in future regimens should be avoided. It can take several weeks to months for lactate concentrations to normalise. If a patient is on NNRTI regimen, a boosted PI
<table>
<thead>
<tr>
<th>Toxicity</th>
<th>Reference</th>
<th>Year</th>
<th>Country</th>
<th>Study type</th>
<th>Sample size</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peripheral neuropathy</td>
<td>Idoko et al</td>
<td>2002</td>
<td>Nigeria</td>
<td>Clinical trial</td>
<td>1 patient</td>
<td>On zidovudine, zalcitabine, or a PI. Severe PN</td>
</tr>
<tr>
<td></td>
<td>Wester et al</td>
<td>2005</td>
<td>Botswana</td>
<td>Longitudinal</td>
<td>153 patients</td>
<td>4.3 fold increase in moderate to severe PN if placed on a stavudine/didanosine regimen.</td>
</tr>
<tr>
<td></td>
<td>Boulle et al</td>
<td>2007</td>
<td>South Africa</td>
<td>Cohort</td>
<td>2679 patients; PN related stavudine substitutions in 6.2% patients (95% CI 4.3-8.5)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Hawkins et al</td>
<td>2007</td>
<td>Kenya</td>
<td>Longitudinal, observational</td>
<td>1286 patients</td>
<td>Predominantly stavudine related regimens. 20.7% PN</td>
</tr>
<tr>
<td></td>
<td>Forna et al</td>
<td>2007</td>
<td>Uganda</td>
<td>Cohort</td>
<td>1029 patients</td>
<td>96% on stavudine containing regimens. 36% PN, 9% were severe. 17.2% required drug change.</td>
</tr>
<tr>
<td></td>
<td>Canestri et al</td>
<td>2007</td>
<td>Senegal</td>
<td>Cohort</td>
<td>40 patients</td>
<td>37.5% PN on a stavudine/didanosine regimen.</td>
</tr>
<tr>
<td></td>
<td>Jamisse et al</td>
<td>2007</td>
<td>Mozambique</td>
<td>Prospective</td>
<td>146 patients</td>
<td>18% PN on either a stavudine or zidovudine based regimen.</td>
</tr>
<tr>
<td></td>
<td>Sanne et al</td>
<td>2009</td>
<td>South Africa</td>
<td>Cohort</td>
<td>7583 patients</td>
<td>Mainly stavudine. Incidence rates of 8.7/100 person years (95% CI 8.1-9.2)</td>
</tr>
<tr>
<td></td>
<td>Minzi et al</td>
<td>2009</td>
<td>Tanzania</td>
<td>Retrospective review</td>
<td>23 clinics</td>
<td>Increase 0.9% to 1% in the Dar-es-Salaam; 1.3% to 3.8% in the Mbeya in PN over a year if on stavudine.</td>
</tr>
<tr>
<td></td>
<td>Sacktor et al</td>
<td>2009</td>
<td>Uganda</td>
<td>Prospective</td>
<td>102 patients</td>
<td>38% PN on stavudine based therapy.</td>
</tr>
<tr>
<td></td>
<td>Beadles et al</td>
<td>2009</td>
<td>Malawi</td>
<td>Retrospective review</td>
<td>4591 patients</td>
<td>On stavudine, 35% symptoms experienced but only 13% given diagnosis of PN.</td>
</tr>
<tr>
<td></td>
<td>Shurie et al</td>
<td>2010</td>
<td>Ethiopia</td>
<td>Cross sectional</td>
<td>203 patients</td>
<td>43% on 30mg stavudine vs. 74% on 40mg stavudine; 20.7% on zidovudine</td>
</tr>
<tr>
<td></td>
<td>van Griensven et al</td>
<td>2010</td>
<td>Rwanda</td>
<td>Cohort</td>
<td>2190 patients</td>
<td>On stavudine based therapy. 8.0% PN. Incidence rates of 52/1000 patient years</td>
</tr>
<tr>
<td>Toxicity</td>
<td>Reference</td>
<td>Year</td>
<td>Country</td>
<td>Study type</td>
<td>Sample size</td>
<td>Result</td>
</tr>
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<td>---------------------------------------------</td>
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</tr>
<tr>
<td>Lactic acidosis/symptomatic hyperlactatemia</td>
<td>Geddes et al</td>
<td>2006</td>
<td>South Africa</td>
<td>Observational case series</td>
<td>891 patients</td>
<td>Stavudine related LA: Incidence rate of 19/1000 person years (95% CI 9-29).</td>
</tr>
<tr>
<td></td>
<td>Wester et al</td>
<td>2007</td>
<td>Botswana</td>
<td>Randomized control trial</td>
<td>650 patients</td>
<td>On stavudine or didanosine based therapy. 2% moderate to severe SH; 1% LA</td>
</tr>
<tr>
<td></td>
<td>Boulle et al</td>
<td>2007</td>
<td>South Africa</td>
<td>Cohort</td>
<td>2679 patients</td>
<td>LA/SH related stavudine substitutions in 4.7% (95% CI 3.0-6.8)</td>
</tr>
<tr>
<td></td>
<td>Bolhaar et al</td>
<td>2007</td>
<td>South Africa</td>
<td>Retrospective cohort analysis</td>
<td>1735 patients</td>
<td>On predominantly stavudine based therapy. Overall incidence rate 10.6/1000 patient years; 16.1/1000 patient years (females); 1.2/1000 patient years (males); LA: 30.4% died. SH: None died.</td>
</tr>
<tr>
<td></td>
<td>Sanne et al</td>
<td>2009</td>
<td>South Africa</td>
<td>Cohort</td>
<td>7583 patients</td>
<td>Mainly stavudine. LA/SH: Incidence rates of 5.1 per 100 person years (95% CI 4.7-5.5)</td>
</tr>
<tr>
<td></td>
<td>van Griensven et al</td>
<td>2009</td>
<td>Rwanda</td>
<td>Cohort</td>
<td>2190 patients</td>
<td>On stavudine based therapy. LA/SH 3.1%; incidence rate 20/1000 patient years.</td>
</tr>
<tr>
<td></td>
<td>Hernandez et al</td>
<td>2010</td>
<td>South Africa</td>
<td>Retrospective</td>
<td>1719 patients</td>
<td>On stavudine based therapy. LA: Incidence rate 13.5 cases/1000 patient years (95% CI 9-29), 22.2% mortality. SH: Incidence rate 31.79 cases/1000 patient years (95% CI 14-40).</td>
</tr>
</tbody>
</table>

*PN: Peripheral neuropathy; LA: Lactic acidosis; SH: Symptomatic hyperlactatemia; 95% CI = 95% confidence interval. Definitions for LA and SHL in different studies vary.
should be added. If the patient has already failed on NNRTI regimen and is on a boosted PI, other alternatives such as raltegravir or etravirine should be added if available. Otherwise the patients should be continued on the boosted PI only (Southern African HIV Clinicians Society, 2012).

1.6.2. Peripheral neuropathy

1.6.2.1. Epidemiology

A distal sensory peripheral neuropathy, characterized by the presence of symmetrical distal anesthesia and/or painful dysesthesia is another common side effect of antiretroviral therapy especially with the use of stavudine and didanosine, and is clinically indistinguishable from peripheral neuropathy associated with the HIV infection per se (Dalakas et al, 2001. Cornblath et al, 2006). Prevalence rates vary from country to country, with rates in the West between 10 and 35% (Keswani et al, 2002. McArthur et al, 2005. Cornblath et al, 2006). Data from neuropathy studies performed in Africa are shown in Table 2. One study from South Africa showed incidence rates of 8.7 per 100 person years (Sanne et al, 2009), with lower rates of 5.2 per100 person-years in Rwanda (van Griensven et al, 2010) and 2.8 per 100 person-years in another site in South Africa (Boullé et al, 2007). Prevalence rates were much higher in African studies compared to those in studies performed in Europe or the USA: 35% and above noted in African countries (Beadles et al, 2009. Canestri et al, 2007. Forna et al, 2007. Sacktor et al, 2009). Risk factors in most studies included male gender, a higher baseline BMIs, age and advanced clinical HIV disease (Boullé et al, 2007. van Griensven et al, 2009. Sanne et al, 2009). Most studies showed that toxicity was cumulative and resulted in a switch from stavudine to other alternative therapies, rather than being due to
treatment failure or contraindications and remained the most important trigger for treatment change (Boulle et al, 2007. van Griensven et al, 2009).

1.6.2.2. Pathogenesis
Peripheral neuropathy is pathologically characterized by distal axonal degeneration of small myelinated and unmyelinated nerve fibers (Zhou et al, 2007), with evidence that abnormal mitochondria and mtDNA depletion are seen with dideoxycytidine (ddC)-associated peripheral neuropathy (Dalakas et al, 2001). Interestingly, there are also genetic predictors for the development of peripheral neuropathy as seen in white patients with a common European mitochondrial haplogroup T (Canter et al, 2008). This was also observed in black patients who belonged to mtDNA subhaplogroup L1c who were found to be at increased risk of developing peripheral neuropathy on antiretroviral therapy (Canter et al, 2010).

1.6.2.3. Management
History and physical examination are important in the diagnosis of peripheral neuropathy. Patients who present with symptoms of numbness or dysesthesia after initiation of antiretroviral therapy especially with the use of stavudine and didanosine should be considered as having a peripheral neuropathy due to the antiretroviral therapy, once other causes are excluded. There are however no grading protocols for the severity of peripheral neuropathies. Whilst the ACTG grading system is available for the ACTG clinical trials, it is subjective and it is not a well known grading system to HIV clinicians especially those who are working in a resource limited environment and those who do not work in a research setting. This would explain variability in the rates observed in most studies.
Diagnostic tests would include nerve conduction studies which would confirm diminished or absent sensory action potential amplitudes with normal or slightly slower motor conduction velocities (Harrison et al, 1995). Other tests would include electromyography which could demonstrate denervation of distal muscles (Simpson et al, 1995), and biopsy of the sural or cutaneous nerve which would show axonal degeneration of myelinated and unmyelinated fibers (Simpson et al, 1995).

The goal of treatment is to control the pain and help maintain physical function for activities of daily living and quality of life. Some patients may need to switch antiretroviral therapy. Pharmacological treatment can be used to manage the patient’s pain. For milder forms of pain, nonsteroidal anti-inflammatory drugs (NSAIDs) may be prescribed. Severe pain can be treated with stronger opioids, such as morphine and methadone. Antidepressants are commonly used in resource limited settings although a controlled study that compared amitriptyline and mexiletine showed equivalent pain relief to that of a placebo (Kieburtz et al, 1998). Anticonvulsants such as gabapentin were found to be effective in reducing pain (Hahn et al, 2004), and carbamazepine, and phenytoin are also commonly used in resource limited settings but there are little data to support their efficacy.

1.6.3. Lipodystrophy

1.6.3.1. Epidemiology

When HAART was introduced in the 1990s, there was a dramatic improvement in patient morbidity and mortality (Brinkman et al, 1999). But soon afterwards, systemic metabolic alterations were observed, often in combination with body-fat distribution changes, and this was called the lipodystrophy syndrome (Brinkman et al, 1999). These metabolic alterations, which were characterized by insulin resistance and dyslipidaemia, were similar to that of the
metabolic syndrome that is seen with obesity. The body fat alterations were associated with wasting in subcutaneous adipose tissue in the face, arms, legs and buttocks i.e. lipoatrophy, with central fat accumulation, with some patients even presenting with breast enlargement as well as with fat accumulation in the dorso-cervical area, also known as “buffalo hump pads” (Lo et al, 1998. Carr et al, 1998, 2003). Some patients developed generalized or central fat accumulation i.e. lipohypertrophy, although most patients would present with a combination of both (mixed lipodystrophy) (Carr et al, 1998, 2003).

The prevalence rates of lipodystrophy vary widely ranging from 20% to 80% in HIV-1 patients receiving HAART (Carr et al, 2003, Jacobson et al, 2005) and this is due to an inconsistency in the diagnostic criteria that are used for identifying these patients. (Carr et al, 2003).

With regards to risk factors for the development of lipodystrophy; the recognition of lipodystrophy in patients on antiretroviral treatment coincided with the use of PIs, which caused visceral fat accumulation and the systemic metabolic changes. The use of NRTIs also contributed to fat wasting, but the concomitant use of PIs may have added to this effect. (Brinkman et al, 1999, Cossarizza A et al, 2001, Carr et al, 2003). Studies identified the use of stavudine and zidovudine, both NRTIs, as major risk factors for lipoatrophy (Mallal et al, 2000. John et al, 2001). Subsequent studies (Gallant et al, 2004. Podzamczer et al, 2008. Young et al, 2005) have confirmed the association between the use of stavudine and the risk of lipoatrophy. Other risk factors include increasing age, female gender, CD4 counts and viral loads (Carr et al, 2003).

The frequency of lipodystrophy in resource-limited countries where predominantly NRTI based regimens are being used is high, although it does vary from country to country (see Table 3). Studies from African countries have revealed rates as high as 34% (Mutimura et al,
2007. Sinxadi et al, 2010). Studies demonstrated that lipoatrophy tended to be common after one year of therapy, especially in patients who were on higher doses of stavudine (40mg) and in patients with a higher body weight (van Griensven et al 2010). Females were also more affected than males (Mutimura et al, 2007. van Griensven et al 2010).

1.6.3.2. Pathogenesis

A proposed aetiology is presented in Figure 2. PIs were initially implicated because of their homology to lipid and adipocyte regulatory proteins (Carr et al, 1998). PIs can inhibit lipogenesis by reducing the differentiation of pre-adipocytes to mature adipocytes by reducing the levels of transcription factors involved in adipocyte differentiation - these include the sterol regulatory element binding protein-1 (SREBP-1), and peroxisome-proliferator activated receptor γ (PPARγ) (Zhang et al, 1999, Lenhard et al, 2000. Caron et al, 2001).

Mitochondria may be affected by the HIV infection itself and may contribute to the mitochondrial toxicity seen in patients on NRTIs. Some studies have confirmed significant lower levels of mtDNA in HIV-1 positive patients than in uninfected patients in peripheral blood mononuclear cells (Côté et al, 2002. Miró et al, 2004. Chiappini et al, 2004). In addition, studies have also confirmed decreased mtDNA content in skeletal muscle (Hauggaard et al, 2005) and nerve tissue (Dalakas et al, 2005). Other tissues involved include adipose tissue (Villarroya et al, 2007).

There are several mechanisms by which NRTIs contribute to mitochondrial toxicity resulting in lipoatrophy. NRTIs impair mtDNA polymerase γ activity resulting in depletion of mtDNA. This has been confirmed by mtDNA depletion in subcutaneous adipose tissue seen in patients with lipoatrophy when compared to uninfected individuals or untreated HIV-1-

However, other studies have also shown alterations in mitochondrial gene expression. Expression of mitochondrial and nuclear genes involved in mitochondrial biogenesis, adipogenesis, and metabolic and endocrine function are linked. *SREBP1* and *PPARY*, together with *PPARA* regulate transcription of lipid metabolism genes and are linked to mitochondrial physiology through *PPARY* coactivator 1 (*PGC1*). In turn, *PGC1* regulates transcription of mitochondrial genes through the master transcription factors - nuclear respiratory factor (NRF) 1 and 2, which regulate mitochondrial transcription factor A (*mtTFAM*) which in turn stimulates mitochondrial gene transcription, thus representing a link between mitochondrial biogenesis and adipose cell function (Enriquez et al, 1999. Spiegelman et al, 2000. Puigserver et al, 2003. Rosen et al, 2006). Mitochondria produce cellular energy through the process of oxidative phosphorylation and the mitochondrial electron transport chain; it is thought that depletion of mtDNA may decrease cellular oxidative phosphorylation (Lewis et al, 2003). The mitochondrial electron transport chain which comprises 5 complexes (I—V) are encoded by both nuclear and mitochondrial genes. Mitochondrial DNA-encoded subunits including cytochrome *b* (*Cyt b*) of complex III and cytochrome *c* oxidase subunits I, II, and III (COX1, COX 2, and COX 3) of complex IV, play functional roles whereas nuclear-encoded components (COX4), play a more regulatory than a functional role (Enriquez et al, 1999. Barrientos et al, 2002). Therefore through a process of mitochondrial fine tuning, mitochondria can adapt transcription of mtRNA to meet altered oxidative requirements independent of regulation by nuclear factors or changes in mtDNA level (Martin et al, 1993. Enriquez et al, 1999).
### Table 3: Body fat abnormalities, dysglycaemia and dyslipidaemia in African HIV-1-infected populations

<table>
<thead>
<tr>
<th>Toxicity</th>
<th>Reference</th>
<th>Year</th>
<th>Country</th>
<th>Study type</th>
<th>Sample size</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lipodystrophy</td>
<td>Boulle <em>et al</em></td>
<td>2007</td>
<td>South Africa</td>
<td>Cohort</td>
<td>2679 patients</td>
<td>LD related stavudine substitutions in 9.0% (95% CI 5.2-14.1)</td>
</tr>
<tr>
<td></td>
<td>Mutimura <em>et al</em></td>
<td>2007</td>
<td>Rwanda</td>
<td>Prospective cohort</td>
<td>571 patients</td>
<td>On either a stavudine (predominantly) or zidovudine based regimen. LD observed in 34%. Peripheral LA with abdominal LH noted in 72% of LD patients.</td>
</tr>
<tr>
<td></td>
<td>Sanne <em>et al</em></td>
<td>2009</td>
<td>South Africa</td>
<td>Cohort</td>
<td>7583 patients</td>
<td>Mainly stavudine. LD: Incidence rate 4.9/100 person years (95% CI 4.5-5.3).</td>
</tr>
<tr>
<td></td>
<td>van Griensven <em>et al</em></td>
<td>2009</td>
<td>Rwanda</td>
<td>Cohort</td>
<td>2190 patients</td>
<td>On stavudine based therapy. LD: 7.2%; incidence rate of 47/1000 patient years.</td>
</tr>
<tr>
<td></td>
<td>Mercier <em>et al</em></td>
<td>2009</td>
<td>Senegal</td>
<td>Prospective cohort</td>
<td>180 patients</td>
<td>On either a stavudine or zidovudine or a PI based regimen. Moderate-severe LD: 31.1% (95% CI 24.3-37.9); LA:13.3%; LH:14.5%;mixed forms,3.3%</td>
</tr>
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<td></td>
<td>George <em>et al</em></td>
<td>2009</td>
<td>South Africa</td>
<td>Longitudinal study</td>
<td>42 patients</td>
<td>On stavudine based therapy. LD: 42.9%; Hypertriglyceridemia: 30.8%. Hypercholesterolemia: 35% in LD group. 35%</td>
</tr>
<tr>
<td></td>
<td>Sinxadi <em>et al</em></td>
<td>2010</td>
<td>South Africa</td>
<td>Cross-sectional study</td>
<td>47 patients</td>
<td>On stavudine. LD: 34%.</td>
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<td>Diabetes/insulin resistance</td>
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<td>2007</td>
<td>Rwanda</td>
<td>Prospective cohort</td>
<td>571 patients</td>
<td>On either a stavudine (predominantly) or zidovudine based regimen. IFG: 18% of LD (P=0.006 vs. HIV negative group); 16% in the non-LD group (P=0.01 vs. HIV negative group); 2% for HIV negative group.</td>
</tr>
<tr>
<td></td>
<td>Bakari <em>et al</em></td>
<td>2007</td>
<td>Nigeria</td>
<td>Case report</td>
<td>1 patient</td>
<td>Zidovudine, PIs. Developed diabetes</td>
</tr>
<tr>
<td></td>
<td>Manuthu <em>et al</em></td>
<td>2008</td>
<td>Kenya</td>
<td>Cross sectional</td>
<td>295 patients</td>
<td>On stavudine. Dysglycemia: noted in 20.7%, with 22.9% on HAART and 17.9% HAART- naïve (p=0.248, OR 0.731, 95% CI 0.412-1.298)</td>
</tr>
<tr>
<td></td>
<td>Mercier <em>et al</em></td>
<td>2009</td>
<td>Senegal</td>
<td>Prospective cohort</td>
<td>180 patients</td>
<td>On either a stavudine or zidovudine or a PI based regimen. Increase in fasting glucose in cases vs.</td>
</tr>
<tr>
<td>Toxicity</td>
<td>Reference</td>
<td>Year</td>
<td>Country</td>
<td>Study type</td>
<td>Sample size</td>
<td>Result</td>
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<td>controls (4.86 vs. 3.60, P&lt;0.0001). No difference in the fasting insulin levels.</td>
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<tr>
<td></td>
<td>Sinxadi <em>et al</em></td>
<td>2010</td>
<td>South Africa</td>
<td>Cross-sectional study</td>
<td>47 patients</td>
<td>On stavudine. Dysglycaemia: 23%; IFG: 19%; IGT: 4%</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td>Mutimura <em>et al</em></td>
<td>2007</td>
<td>Rwanda</td>
<td>Prospective cohort</td>
<td>571 patients</td>
<td>On either a stavudine (predominantly) or zidovudine based regimen. Hypertriglyceridemia: 9% in LD group. Hypercholesterolemia: 14% in LD group.</td>
</tr>
<tr>
<td></td>
<td>Manuthu <em>et al</em></td>
<td>2008</td>
<td>Kenya</td>
<td>Cross sectional</td>
<td>295 patients</td>
<td>On stavudine predominantly. Hypercholesterolemia: 39.2% in HAART group vs. HAART naïve group, prevalence of an elevated LDL in the HAART group was significantly higher than the HAART naïve group (40% vs. 11%; p value &lt;0.0001)</td>
</tr>
<tr>
<td></td>
<td>Buchacz <em>et al</em></td>
<td>2008</td>
<td>Uganda</td>
<td>Prospective</td>
<td>374 patients</td>
<td>On a stavudine regimen with a single drug substitution to zidovudine. Increase mean lipid levels from 0 to 24 months in total cholesterol 31 mg/dl (95% CI 26-36), LDL-cholesterol 26 mg/dl (95% CI 22-29), HDL-cholesterol 19 mg/dl (95% CI 17-21)</td>
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<tr>
<td></td>
<td>Mercier <em>et al</em></td>
<td>2009</td>
<td>Senegal</td>
<td>Prospective cohort</td>
<td>180 patients</td>
<td>On either a stavudine or zidovudine or a PI based regimen. Triglyceride (1.03 mmol/L vs. 0.89 mmol/L, p=0.007) and HDL-C levels (1.33 mmol/L vs. 0.37 mmol/L, p=0.008) were increased in patients vs. controls</td>
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<td></td>
<td>Lesi <em>et al</em></td>
<td>2009</td>
<td>Nigeria</td>
<td>Prospective cross sectional</td>
<td>113 patients</td>
<td>On ‘HAART’. 28.3% of patients with fasting cholesterol values &gt;200mg/dl, 24% had an elevated LDL-cholesterol; 35% had elevated triglyceride levels.</td>
</tr>
<tr>
<td></td>
<td>Salami <em>et al</em></td>
<td>2009</td>
<td>Nigeria</td>
<td>Prospective cohort</td>
<td>327 patients</td>
<td>Hypertriglyceridaemia was greater in the PI vs. NNRTI group (79% vs. 54%) with a higher risk RR 2.95 (95% CI 1.89-4.59) P= 0.0003; with hypercholesterolaemia (51% vs. 31%), also with a higher risk, RR 2.44(95% CI11.72-3.47) P =0.0001</td>
</tr>
<tr>
<td>Toxicity</td>
<td>Reference</td>
<td>Year</td>
<td>Country</td>
<td>Study type</td>
<td>Sample size</td>
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<td></td>
<td>Sinxadi et al</td>
<td>2010</td>
<td>South Africa</td>
<td>Cross-sectional study</td>
<td>47 patients</td>
<td>On stavudine. Hypertriglyceridaemia: 23%.</td>
</tr>
</tbody>
</table>

* LD: Lipodystrophy; IFG: Impaired fasting glucose; IGT: Impaired glucose tolerance; DM: Diabetes mellitus; HIV: Human immunodeficiency virus; HAART: Highly active antiretroviral therapy; PI: Protease inhibitor; NNRTI: Non-nucleoside reverse transcriptase inhibitor; 95% CI = 95% confidence interval.
In a study where HIV negative patients were exposed to dual NRTIs for two weeks, decreased transcription of mtRNA was observed with no significant depletion of mtDNA in adipose tissue, and this occurred with the up-regulation of *NRF1* and *TFAM* whereas *PGC1α* was down-regulated. The investigators proposed that NRTIs caused mitochondrial dysfunction by means other than through inhibition of DNA polymerase-γ, and that disruption of expression of lipid metabolism genes played a role in NRTI-induced lipoatrophy (Mallon *et al.*, 2005).

In another study when comparing stavudine versus zidovudine, patients who developed drug induced lipoatrophy after 18 months of therapy, were noted to develop significant mtDNA depletion and significant decrease in the ratio of *COX3* to *COX4* expression. In addition, a
significantly lower expression of SREBP1 and the glucose transporter (GLUT)4 gene were also noted in the lipoatrophic patients. It was also noted that PGC1B was significantly higher in the lipoatrophic group than in those that had no lipoatrophy. In addition, in those patients with lipoatrophy, the stavudine group had a significantly lower expression of POLG1, COX3/COX4 ratio and SREBP1 than the zidovudine group (Sievers et al, 2009).

The role of NNRTIs in lipoatrophy has also been investigated. In one study, efavirenz was associated with a great risk of limb fat loss than PIs when combined with NRTIs (stavudine and zidovudine containing regimens) (Haubrich et al, 2009). In a recent systematic review, this causal relationship between NRTIs and lipoatrophy was confirmed, but it was suggested that the concomitant use of PIs had an ameliorating effect on the body fat changes. It was also suggested that efavirenz caused an additive toxicity. In addition, they noted that central fat gain was as a consequence of treating the HIV infection and not linked to any antiretroviral class (de Waal et al, 2013).

HIV-1 infection itself can also lead to alterations in gene expression for mitochondrial proteins. The expression of several nuclear and mtDNA-encoded genes for mitochondrial proteins has been shown to be reduced in adipose tissue from HIV-1-infected patients who were not on antiretroviral therapy (Giralt et al, 2006). This was consistent with previous studies of mtDNA depletion and mitochondrial respiratory dysfunction in peripheral blood mononuclear cells and subcutaneous fat from HIV-1-infected patients before antiretroviral therapy (Casula et al, 2005. Garrabou et al, 2011).

1.6.3.3. Management

The diagnosis of lipodystrophy is generally clinically obvious; the patient may present with lipoatrophy or lipohypertrophy and this can be confirmed by anthropometric measurements.
over time. Additionally, radiological imaging if available should also be used to confirm the diagnosis. This can be done by using dual energy x-ray absorptiometry (DEXA) scans, bioelectrical impedance analysis, computed tomography (CT), or magnetic resonance imaging (MRI) (Carr et al, 2003).

There are medical and surgical strategies for the management of lipoatrophy and there is a possibility that lipoatrophy changes can recover although it may be slow, or incomplete. This can include antiretroviral drug substitution. An improvement in lipoatrophy may occur when NRTIs are switched, although the dyslipidaemia maybe unaffected. Options include switching stavudine to abacavir or tenofovir (Carr et al, 2002. Moyle et al, 2006) which can result in a significant increase in limb fat mass for up to two years but may not result in complete resolution of clinical lipoatrophy. One could also reduce the stavudine dose (Milinkovic et al, 2007). Other drug interventions that can be used to reverse fat loss include the use of thiazolidinediones which improve adipocyte differentiation and increase limb fat mass. A meta-analysis has shown that pioglitazone and not rosiglitazone was more effective in increasing limb fat (Raboud et al, 2010). Statins and uridine have also been shown to improve lipoatrophy in small trials (Mallon et al, 2006. Sutinen et al, 2007).

Cosmetic surgery is an option for facial lipoatrophy. This includes surgically placed alloplastic, autologous, or synthetic implants, injection of temporary fillers such as poly-L-lactic acid, and injection of permanent fillers such as silicone (Guaraldi et al, 2011).

Liposuction has been shown to be helpful in patients with lipohypertrophy although there was a potential for recurrence (Hultman et al, 2007). Exercise or diet may improve lipoatrophy (Leyes et al, 2008), but one should be careful in patients with low body mass indices (BMIs).

Exercise training has been shown to improve outcomes in body composition and metabolic profiles in patients in resource limited settings (Mutimura et al, 2008).
On the other hand, in patients with lipohypertrophy, low caloric feeding and combined resistance and aerobic exercise has been shown to be beneficial in obese, HIV-infected women although there was no improvement in insulin resistance or lipids (Engelson et al, 2006).

1.6.4. Insulin resistance and diabetes mellitus

1.6.4.1. Epidemiology

Prior to the advent of antiretroviral therapy, disorders of glucose metabolism were relatively uncommon in patients with HIV-1 infection (Samaras et al, 2009). Determination of the prevalence and incidence of glucose disorders in HIV-1-infected patients is complicated by multiple confounders, ranging from inherent differences in study populations to differences in the diagnostic methods used (Paik et al, 2011).

Early cases of diabetes were described with PI use (Dubé et al, 1997. Walli et al, 1998), with other metabolic complications such as dyslipidaemia and lipodystrophy occurring at the same time, making it difficult to determine which factors were contributing to the changes in glucose metabolism (Samaras et al, 2009).

Evidence for increased prevalence of diabetes and insulin resistance was also derived from cohorts of patients on antiretroviral therapy with lipodystrophy where the rates of disorders of glucose metabolism such as diabetes mellitus, impaired fasting glucose, and impaired glucose tolerance were as high as 25% and were considered to be related to the specific PIs that were used (Carr et al, 1998. Carr et al, 1999). NRTIs also played a role, where cumulative exposure to nucleoside analogs was associated with insulin resistance in some studies (Brown et al, 2005. Tien et al, 2008). Rates from Africa vary (Table 3), with rates of dysglycaemia as high as 20% in two cross sectional studies (Manuthu et al, 2008. Sinxadi et al, 2010). Fat
redistribution may also contribute to insulin resistance (Meininger et al, 2002); one study suggested that the severity of lipoatrophy, among patients taking either stavudine or zidovudine, was associated with an increased risk of insulin resistance (De Wit et al, 2008). Other studies examining the prevalence and predictors of glucose intolerance and insulin resistance, and their association with antiretroviral therapy also found that obesity, gender, ethnicity, a family history of diabetes, and hepatitis C were all predictors (Mehta et al, 2003. Yoon et al, 2004. Amorosa et al, 2005. Salehian et al, 2005. Tien et al, 2008).

1.6.4.2. Pathogenesis

Evidence generally suggests that nucleoside analogs exert their effects on glucose metabolism indirectly, through changes in body composition and mitochondrial toxicity (De Wit et al, 2008). This may be the case in Africa where predominantly NRTIs are used, and where rates of dysglycemia vary between 8-23% (Table 3).

On the other hand, PIs can inhibit the uptake of glucose into cells by interfering with the GLUT4 glucose transporter (Murata et al, 2000. Hertel et al, 2004). Another proposed mechanism is the effects of PIs on adipokine metabolism. PIs reduce adipocyte differentiation by altering SREBP-1 maturation and PPAR-γ expression resulting in a decreased expression of adipocytokines (Zhang et al, 1999, Lenhard et al, 2000. Caron et al, 2001). Apart from direct PI-induced effects on insulin resistance, discrete and independent insults to glucose metabolism may arise from HAART-induced dyslipidaemia, lipotoxicity, and lipodystrophy (Gutierrez et al, 2012).
1.6.4.3. Management

The diagnosis of diabetes mellitus should be confirmed using formal laboratory testing which would include a fasting blood glucose or an oral glucose tolerance test and more recently a glycosylated haemoglobin (WHO, 2011). Just like in HIV negative patients, diet and exercise are important in the management of insulin resistance and diabetes in overweight patients with HIV infection (Falutz et al, 2007). Similarly, if this is not achieved drug interventions may be required. Metformin has been shown to reduce insulin resistance and cardiovascular risk parameters in HIV-infected patients with lipodystrophy (Hadigan et al, 2000), but the use of metformin in patients on stavudine may increase the risk of lactic acidosis and may worsen lipoatrophy in HIV infected patients with lipoatrophy. Rosiglitazone has also been shown to improve insulin sensitivity and increased adiponectin levels (Hadigan et al, 2004).

1.6.5. Dyslipidaemia

1.6.5.1. Epidemiology

Lipid abnormalities have been noted in HIV-1 infected patients even before the advent of antiretroviral therapy, and this was characterised by decreased levels of total, LDL and HDL cholesterol and elevated triglyceride levels (Constans et al, 1994. Grunfeld et al, 1998). HIV-1 associated dyslipidaemia is associated with increased levels of interferon α which has been correlated with elevated plasma triglycerides (Carpentier et al, 2005). Similarly, tumour necrosis factor (TNF) α has also been shown to be elevated in HIV-1-positive, antiretroviral therapy-naïve patients. Here it has been shown to interfere with free fatty acid metabolism, lipid oxidation and to attenuate insulin-mediated suppression of lipolysis (Johnson et al, 2004. Oh et al, 2007). The prevalence of lipid abnormalities varies depending on the type of antiretroviral therapy used. In patients receiving a PI-containing antiretroviral therapy, the
prevalence rates of dyslipidaemia were high (>70%) and it predominantly included hypertriglyceridaemia followed by hypercholesterolaemia (Dong et al., 1999. Carr et al., 1999. Savès et al., 2002). Although metabolic alterations are more common among patients with lipodystrophy, they are also present in those without body changes; in fact these metabolic abnormalities may precede the body fat changes (Vigouroux et al., 1999. Mulligan et al., 2000). Results from various studies are contradictory when comparing risk factors for the PI-related hyperlipidaemia. Although elevations in serum triglyceride and cholesterol levels have been associated with all the available PIs, hypertriglyceridaemia seems more frequent in patients receiving particular types of PI combination therapy (Carr et al., 1998. Carr et al., 1999). In a review (Calza et al., 2003), different PI-based therapies were associated with different lipid profiles, but hypertriglyceridaemia was noted to be more frequent in patients receiving a ritonavir, ritonavir/saquinavir or ritonavir/lopinavir combination therapy when compared with indinavir, nelfinavir and amprenavir based therapies. Atazanavir is the most lipid stable PI currently available (Cahn PE et al., 2004). In several studies, hypertriglyceridaemia has been found to be associated with male gender, advanced age, a higher body mass index, diagnosis of AIDS, higher mean CD4 count, and elevated cholesterol and triglyceride plasma levels prior to the PI therapy initiation while in other studies, gender, baseline lipid levels, stage of HIV-1 disease and body weight do not appear to be related to the occurrence of hypertriglyceridaemia (Mulligan et al., 2000. Savès et al., 2002. Thiébaut et al., 2003).

Lipid abnormalities also occur in patients on NRTIs such as stavudine and zidovudine. Data from countries in Africa confirm this (Table 3). A study from Senegal, in which the main drugs used were zidovudine and stavudine, showed both triglyceride and HDL-cholesterols were significantly raised in patients when compared to controls but the use of PIs may have
been a confounder (Mercier et al, 2009). In a smaller Ugandan study, where all patients were on stavudine based regimens, a comparison between two NNRTIs, efavirenz and nevirapine was also assessed. After two years of therapy, changes were thought to be related to the use of nevirapine (Buchacz et al, 2008). In a study from Rwanda, where no PIs were used and stavudine was the commonest drug used, high rates of elevated triglycerides and total cholesterol were noted in patients with lipodystrophy (Mutimura et al, 2007). Similarly, a longitudinal study from South Africa showed a significant increase in both triglyceride and cholesterol levels in patients on stavudine followed over a period of two years, but this was noted in patients with and without lipodystrophy (George et al, 2008).

1.6.5.2. Pathogenesis

Concerning the proposed pathogenesis of PI-associated dyslipidaemias which are based partly on speculation in reviews rather than original research - PIs and cellular proteases acting in mitochondrial biogenesis underlie the metabolic alterations because of their homology. PIs bind to cellular retinoic acid binding protein 1 (CRABP1), which has a similar structural homology to the catalytic region of HIV-1 protease enzyme, thus inhibiting the formation of cis-9-retinoic acid from retinoic acid, leading to reduced retinoid X receptor-peroxisome proliferator-activated receptor γ (RXR-PPARγ) activity. This increases apoptosis and decreases proliferation of peripheral adipocytes causing peripheral lipoatrophy, and decreased adiponectin levels. As a result, there is a free fatty acid spill over from apoptotic peripheral adipocytes increasing free fatty acid flux to the liver and skeletal muscle. In the liver, the increased free fatty acid supply and the up-regulation of the triglyceride synthetic pathway through SREBP1 increase hepatic triglyceride and ultimately secretion of

PIs may also bind to low density lipoprotein receptor-related protein 1 (LRP1) interfering with LRP1-lipoprotein lipase complex formation, reducing adipose storage capacity and increasing plasma triglyceride-rich lipoproteins (Oh et al, 2007). It is also possible that PIs may have direct effects on hepatic triglyceride synthesis by up regulating expression of key triglyceride biosynthetic enzymes (Carr et al, 1999).

As discussed in the previous sections, NRTIs can cause mitochondrial dysfunction through inhibition of DNA polymerase-γ, and that disruption of expression of lipid metabolism genes which may be responsible for abnormalities in several cell types such as adipocytes, leading to lipoatrophy, and in skeletal muscle leading to insulin resistance, with secondary dyslipidaemia (Cossarizza et al, 2003. Mallon et al, 2005. Pinti et al, 2006).

1.6.5.3. Management

The diagnosis is made by an overnight fasting lipid profile, similar to that in a HIV negative patient. Exercise, weight reduction and diet are important components in the management of such patients (Lazzaretti et al, 2012. Mutimura et al, 2008. Stein et al, 2008). Generally, when initiating HAART in a patient with cardiovascular risk factors, drugs with a more favourable lipid profile should be given, such as atazanavir.

Other options would include lipid lowering drugs. Most statins are metabolized by the cytochrome P450 3A4 (CYP3A4) enzyme. With regards to NNRTIs, whilst nevirapine is a selective inducer of CYP3A4, efavirenz is both an inducer and inhibitor of CYP3A4 (Dubé et al, 2003). There is no data on the combined use of nevirapine and statins. Efavirenz leads to significant induction of statin metabolism (Gerber et al, 2005) which may lead to decreased
efficacy of statin therapy, but on the other hand, efavirenz metabolism is not affected by statins. This would suggest that an increased dosing of statins may be needed in patients who are taking both agents, but closely monitor for drug toxicity.

All PIs down regulate the activity of CYP3A4, therefore leading to an increased serum concentration of the statin and the potential for adverse reactions such as rhabdomyolysis. Therefore statins such as lovastatin or simvastatin are contraindicated (Dubé et al, 2003), but atorvastatin and pravastatin are other options. Atorvastatin levels are known to significantly increase with the use of PIs – especially with lopinavir/ritonavir, and therefore there is a concern for a rhabdomyolysis.

In contrast to the statins, fibrates are unlikely to have significant drug interactions with antiretroviral medications. Fibrates however have not been shown to reduce cardiovascular morbidity and mortality at the same degree as statin therapy. There is limited data on the use of ezetimibe in HIV positive patients although it was shown to be well tolerated with changes mainly in the LDL cholesterol (Bennett et al, 2007. Coll et al, 2006. Stebbing et al, 2009).

1.6.6. Cardiovascular risk

As summarised in Figure 3, the risk of cardiovascular disease (CVD) is due to a number of factors which include HIV associated inflammation, traditional cardiovascular risk factors and the metabolic side effects from antiretroviral therapy.

1.6.6.1. The role of HIV associated inflammation

Through systemic inflammation, hypercoagulability and platelet activation, HIV infection via its viral proteins causes endothelial dysfunction and CVD (Dau et al, 2008). Independent of cardiovascular risk factors and antiretroviral therapy, HIV-1 replication and a low baseline
CD4 count are associated with an increased risk of myocardial infarctions in HIV-1-infected individuals (Lang et al, 2012). The HIV proteins gp120, Tat, and Nef induce expression of several adhesion molecules and inflammatory cytokines such as intercellular adhesion molecule (ICAM) -1, vascular cell adhesion molecule (VCAM)-1, E-selectin, tumour necrosis factor (TNF)-α, and interleukin (IL)-6 resulting in increased leukocyte adherence to the endothelium (Mu et al, 2007). Increased levels of vWF (von Willebrand factor), a glycoprotein facilitating platelet adhesion, are synthesized in endothelial cells and inflammatory cells, and correlate with circulating levels of inflammatory cytokines (Mu et al, 2007). A hypercoagulable state may also occur depending on plasma viral load. These HIV proteins can also induce endothelial apoptosis and increase endothelial permeability (Mu et al, 2007). Elevated levels of highly sensitivity C-reactive protein (hs-CRP), an inflammatory marker, are associated with an increased risk of cardiovascular events in the general population (Rutter et al, 2004. Pai et al, 2004) as well as in HIV-positive patients (Reingold et al, 2008. Triant et al, 2009).

1.6.6.2. The role of traditional cardiovascular risk factors

The Women's Interagency HIV Study and the Multicenter AIDS Cohort Study evaluated the presence of major risk factors for cardiovascular disease and compared HIV-1 positive with HIV-1 negative controls where the 10-year risk of developing coronary heart disease was estimated using the Framingham risk score. It was noted that HIV-1-infected individuals had lower HDL-cholesterol and elevated triglyceride levels when compared with HIV-1 negative controls and the risk of coronary heart disease was significantly lower in patients who were treatment-naive compared with those patients taking PIs (Kaplan et al, 2007).
CVD is prevalent in many African countries, with studies from various countries demonstrating a rise in traditional risk factors (Dalal et al, 2011). In two studies from South Africa which discuss HIV and coronary artery disease specifically (Becker et al, 2010, 2011), it was noted that HIV infected patients who presented with acute coronary syndromes tended to be younger, were antiretroviral therapy naïve and had less traditional risk factors. They noted a lower atherosclerotic burden in the HIV group on angiography but a larger burden of thrombus, suggesting a prothrombotic state in the pathogenesis of acute coronary syndrome. They also noted that despite these HIV infected patients having fewer traditional risk factors, a thrombophilic state tended to be more common. In another study, lower HDL-cholesterol levels in the HIV infected patients, but significantly higher hs-CRP, ICAM-1 and VCAM-1 levels were noted (Fourie et al, 2011).

The extent of cardiovascular complications in HIV-1-infected patients on antiretroviral therapy in Africa will become more evident in future long term studies.

1.6.6.3. The metabolic side effects of antiretroviral therapy.

Whilst antiretroviral therapy improves endothelial function (Arildsen et al, 2013); the metabolic side effects are a concern. Evidence from the Data collection on Adverse Effects of Anti-HIV Drugs Study (DAD) and other studies have established that exposure to certain antiretroviral drugs is associated with an increase in the rate of CVD events (Currier et al, 2003. Mary-Krause et al, 2003. DAD Study Group et al, 2007). In particular PI use, is significantly associated with cardiac events (DAD Study Group, 2007). This was mainly seen with the cumulative use of indinavir or lopinavir-ritonavir (Worm et al, 2010). Evidence also implicates NRTI use such as abacavir with an increased risk of myocardial infarction. The DAD cohort noted that there was an increased risk of myocardial infarction in those exposed
to abacavir and didanosine within six months of use prior to the myocardial infarction, but this was not seen with other NRTIs, including tenofovir (Worm et al., 2010). The Strategies for Management of Antiretroviral Therapy (SMART) study preliminary results demonstrated a fourfold risk of myocardial infarction and a significant increase in major cardiovascular events with abacavir use compared to other NRTIs. No association was found with didanosine (SMART study group, 2008). However, a meta-analysis of randomized controlled trials did not support this hypothesis (Cruciani et al., 2011).

Despite evidence that antiretroviral therapy may be associated with cardiovascular risk; its discontinuation may result in an even greater risk of disease. The virus itself may increase cardiovascular risk through endothelial dysfunction as discussed above and therefore suppression with antiretroviral therapy attenuates this risk. The SMART study group (2006) demonstrated that interruption of antiretroviral therapy was associated with an increased risk of major cardiovascular events compared to those on the continuous arm (SMART study group, 2006). There were more fatal and non-fatal cardiovascular events in the former compared to the later group suggesting that viral suppression may actually reduce cardiovascular risk in the short term. Patients with HIV-1 infection also appear to have a high rate of silent myocardial ischemia. In HIV-1-infected patients without known coronary artery disease from the SMART study, baseline electrocardiograms (ECGs) were assessed for the presence of asymptomatic Q-waves or ST segment depression. ECG evidence for asymptomatic ischemic heart disease was present in nearly 11 percent of the patients. (SMART study group, 2006).
Figure 3: Summary of the HIV/HART metabolic complications contributing to CVD.
1.6.6.4. **The role of adipokines and inflammatory markers**

Adipokines may also have a role in the aetiology of CVD in HIV-1 patients. Leptin is expressed mainly in adipose tissue, its main function is to increase energy expenditure, and to inhibit both appetite and weight gain (Mallewa *et al*, 2008). The effect of antiretroviral therapy on leptin levels is controversial; where some studies have suggested that low levels of leptin are associated with lipodystrophy others have demonstrated no effect (Dzwonek *et al*, 2007. Calmy *et al*, 2009). One study from South Africa showed that leptin levels were associated with viral loads and contributed to disease progression (Azzoni *et al*, 2010). Some studies have suggested that because leptin has both proinflammatory and proatherogenic properties (Smith *et al*, 2011. Falasca *et al*, 2010), it may play a role in long-term cardiovascular events in HIV infected patients.

Adiponectin is a 30 kDa cytokine synthesized and secreted by adipose tissue, and is a potent insulin sensitizer (Mallewa *et al*, 2008). Its expression is reduced in obesity and type 2 diabetes, showing an inverse relationship with insulin resistance and visceral adiposity (Cui *et al*, 2011). Studies have shown adiponectin levels were suppressed in patients with HIV infection and fat redistribution, but the underlying mechanisms are not clear (Tong *et al*, 2003. Sankalé *et al*, 2006). Patients treated with antiretroviral therapy especially those with lipodystrophy, showed gradual down regulation in adiponectin serum levels (Luo *et al*, 2009), accompanied by accelerated cardiovascular impairment (Bezante *et al*, 2009). Therefore, the suppression of adiponectin levels in HIV-infected patients under antiretroviral therapy may negatively affect numerous metabolic parameters leading to cardiovascular events (Palios *et al*, 2012).

CRP has been shown to be a marker of increased cardiovascular events (Pai *et al*, 2004), and CRP levels have been shown to be elevated in patients with HIV compared with general

1.7. Management issues - implications for South Africa and other African countries

1.7.1. The issue of stavudine versus tenofovir.

Stavudine continues to be used as alternative first-line treatment in developing countries despite the change in the WHO antiretroviral therapy guidelines for treating patients with HIV infection (WHO, 2010). This is because of a lack of alternatives, shortages of tenofovir and abacavir, contraindications to the use of other NRTI drugs and its low cost and availability in fixed dose combinations. There is a dire need to increase the number of people receiving antiretroviral therapy worldwide, particularly in sub-Saharan Africa. According to the WHO, at least 15 million HIV-1 infected people are in need of therapy (WHO, 2012). In fact, shortages of tenofovir and abacavir continue to exist intermittently at health facilities across South Africa (Schowalter et al, 2012).

The standard dosage for stavudine is 40 mg given twice daily, as this regimen received regulatory approval in 1995 because of its efficacy, but data continues to suggest that stavudine related toxicities have been a major problem in South Africa and other African countries (Bolhaar et al, 2007. Boulle et al, 2007. Geddes et al, 2006. Hernandez Perez et al, 2010. Mutimura et al, 2007. van Griensven et al 2010) and is rarely used in North America and Europe because these side effects (Gallant et al, 20004).
In a landmark trial - the Gilead 903 trial, in 2004 - patients were randomized to receive lamivudine plus efavirenz, with either stavudine 30 or 40 mg twice daily, or tenofovir. This trial showed an equivalent efficacy when comparing the two drugs with a similar incidence of drug resistance rates at failure, but the proportion of patients with investigator-defined lipoatrophy was higher in the stavudine arm when compared to the tenofovir arm. However, the overall rate of serious adverse events between the arms over a period of three years was 27% in the tenofovir arm compared to 25% in the stavudine arm (Gallant et al, 2004).

In 2007, the WHO recommended a reduction in the dose of stavudine from 40mg to 30mg twice daily, after a review of dose-ranging studies that showed the same clinical and virological efficacy at the lower dose of 30mg twice daily but fewer complications, it was thought that the risk of peripheral neuropathy at this dose would be lower and patients were less likely to discontinue treatment (WHO, 2007). This advice did not suggest weight-adjusted doses, despite evidence from a review of clinical trials supporting a dose of 20 mg twice daily for body weights below 60kg (Hill, 2010). However, in 2010, the WHO and South African National Department of Health guidelines changed to a tenofovir based regimen as first line therapy. While the various toxicities associated with stavudine have been well documented, especially with the higher doses, it is less clear whether a lower dose of stavudine would have less toxic side effects.

The optimal dose for stavudine has been continually evaluated since its initial approval in 1995. A meta-analysis of nine clinical trials examining the dose of stavudine demonstrated that a dose of 30mg twice daily has equivalent efficacy to the 40mg standard-dose prescribed to adults weighing >60 kg, with some evidence of fewer side-effects, including neuropathy and to a lesser degree lipoatrophy (Hill et al, 2007). Interestingly, in this same meta-analysis (Hill et al, 2007), weight adjusted dosing and the mean body weight at baseline in three of
these studies (Ruxrungtham et al, 2000. Siangphoe et al, 2004. Ribera et al, 2005) was close to 60 kg, suggesting that 50% of the patients in these trials were receiving either stavudine 30mg twice daily in the control arm, or 20 mg twice daily in the low dose arm. The data in these studies suggested that there was strong evidence that lower doses of stavudine reduced the risk of peripheral neuropathy, and that there was some evidence for a correlation between stavudine dosing and the risk of lipid changes, lactic acidosis, and lipodystrophy (Hill et al, 2010).

Two studies where the “switch strategy” was used, patients were already on stavudine 40mg twice daily and this might have influenced the end results. In the first randomized controlled trial in the USA, the stavudine dose was cut by half, i.e. 40mg to 20mg twice daily in patients ≥ 60 kg or 30mg to 15mg twice daily in patients <60 kg, and this resulted in increased fat mtDNA and decreased lactate suggesting improvement in mitochondrial indices while preserving virologic suppression. The results suggested that for those patients who were more than 90% adherent, switching to low-dose stavudine improved mitochondrial indices while maintaining virologic suppression. (McComsey et al, 2008). Similarly, in the second study from Spain, patients on stavudine 40mg twice daily were either randomised to switch to 30mg twice daily, switch to tenofovir or remain on the same dose of stavudine. Although no differences were noted in mitochondrial indices, there were significant lipid improvements amongst those switched to tenofovir (Milinkovic et al, 2007).

The evidence from these stavudine dose ranging trials supports the use of stavudine at lower doses including doses as low as 20mg twice daily. The question remains on whether to use a lower dose of the cheaper stavudine-based therapy to treat more patients despite the risk of complications or treat fewer patients with better-tolerated but more expensive drugs with fewer side effects such as tenofovir. There have been no such studies in Africa which has the
biggest burden of HIV disease and limited resources. The only way to reliably answer this question would be to undertake a study comparing low dose stavudine with high dose stavudine and with tenofovir. Such a study is important because it is unlikely that stavudine will ever be completely phased out of use in countries with limited financial resources (Menezes et al, 2012).

1.7.2. Diagnostic challenges in resource-limited settings

The monitoring of side effects of HAART is difficult to achieve in resource-limited countries. The lack of laboratory and radiology services provides several challenges to the accurate diagnosis of lipodystrophy and the metabolic complications of HAART such as dyslipidaemia and diabetes. The availability and cost of routine measurements of cholesterol, glucose and glycosylated haemoglobin may be an issue in resource-limited areas. Not all medical facilities have access to sophisticated imaging such as dual-energy radiographic absorptiometry (DEXA) scans, computerised tomography (CT) imaging technology and magnetic resonance imaging (MRI) for assessment of lipodystrophy. An alternative consensus definition for the diagnosis of lipodystrophy is required for resource-limited environments. An objective case definition of lipodystrophy has been developed (Carr et al, 2003), but it requires access to DEXA and CT imaging technology, which are not available in most resource-limited environments. A wide variation in lipoatrophy rates were noted in most studies from Africa (Boullé et al, 2007. van Griensven et al, 2010). It is possible that only cases that were severe enough to warrant a regimen change were noted. Similarly, a wide variability in the rates of peripheral neuropathies was observed in most studies. Therefore grading protocols for the diagnosis of the severity of peripheral
neuropathies are required as nerve conduction testing equipment, electromyography or the expertise and facilities for nerve biopsies are not available in most African centres.

1.7.3. Other treatment considerations

The cost of medications is a major issue for most resource limited countries. In 2002, six million people worldwide were eligible for antiretroviral therapy (using a CD4+ cell count threshold of 200/mm$^3$), but less than 5% of them received treatment. In 2011, in addition to the 8 million people on antiretroviral therapy, nearly 3 million people with CD4+ counts of less than 200/mm$^3$ were still in need therapy (WHO, 2012). The cost of antiretroviral therapy remains a major barrier to the scale-up of treatment. In 2010, approximately 56% of HAART regimens within resource-limited countries still contained stavudine because of its cheaper costs (WHO, 2010). A study from South Africa demonstrated that the price of tenofovir would need to decrease from USD$ 17 to USD$ 6.17 per month to have the same overall cost as stavudine based therapy (Rosen et al, 2008).

Drug–drug interactions are also an important concern, especially drug interactions between antiretroviral drugs and those drugs used for their treating metabolic consequences.

There are no contraindications or drug interactions related to the concurrent use of antiretroviral therapy and the antidiabetic drugs, although there may be an increased risk of lactic acidosis with the use of metformin and NRTIs, an increased risk of liver dysfunction with the use of PIs and thiazolidinediones has been described (Dau et al, 2008). Other agents such as the sulfonylureas and insulin can be used.

With regard to therapy of HAART associated dyslipidaemia, the use of lipid lowering agents may be limited especially because of the drug interactions. As discussed previously, simvastatin and atorvastatin, the commonly available cheaper options, are contraindicated
(National Antiretroviral Treatment Guidelines, 2004), whilst the availability of pravastatin which is generally safer, is limited.

The management of other risk factors such as hypertension include the use of calcium channel blockers, which are available but interactions with certain NNRTIs and PIIs is a possibility, but other classes of antihypertensives are safe (Dau et al, 2008).

The need for plastic surgery with traditional cosmetic techniques such as facial fillers, liposuction, abdominolipectomy, dorsal hump lipectomy and breast reduction has increased dramatically (Wolfert et al, 1999. Strauch et al, 2004. Warren et al, 2008). However, there is a lack of this surgical facility in resource-limited settings. In a recent study from South Africa, nearly 6% of patients considered stopping antiretroviral therapy due the development of lipodystrophy, with 47% patients saying that they would consider surgery to correct unwanted physical changes (Zinn et al, 2013).

1.8. Aims of the thesis

Despite the major improvement in the survival of HIV-1 infected individuals since the South African Government introduced highly active anti-retroviral therapy in the public sector in 2004, there is an increasing recognition that adverse events remain an important source of morbidity and even mortality, especially with the effects of stavudine-related toxicities. Knowledge of the adverse effects of different therapeutic regimes and an understanding of the pathogenesis that modulate the risk of toxicity, are important in the management of HIV/AIDS patients as clinical practice is increasingly moving towards the use of regimes that combine high levels of tolerability and efficacy. In addition, HIV-1-infected individuals on HAART develop a pattern similar to that of the metabolic syndrome, ultimately putting them at risk for impending cardiovascular disease. Most studies on the metabolic effects of
HAART have been done in the West, but there is little data on patients from African countries including South Africa.

With this in mind, the first aim of this PhD was to make use of a large clinic-based survey of HIV-1-positive patients, newly initiated onto HAART over a period of three years (2004-2008), to compare and describe the effect of stavudine-based therapy on acute and chronic toxicities after initiation of HAART. This was the largest study of the side effects of stavudine therapy conducted to date in a South African HIV-1-positive population and formed the basis for the second aim of this PhD which was a clinical trial.

Whilst the various toxicities associated with stavudine have been well documented, what is less clear is whether the dose of stavudine plays a role in the development of these toxicities. Recent studies have suggested that reduced doses of stavudine (i.e. 20 or 30 mg twice daily) diminish its toxicity while maintaining efficacy. It is therefore possible that, because stavudine is cheaper than tenofovir, and its side effects can be minimized with dose reduction, it would allow more patients to be initiated on HAART in countries where the switch away from stavudine may not yet have occurred. In 2009, this open-label randomized controlled clinical trial was undertaken to assess whether there were any early differences (4 weeks) in the effect of tenofovir when compared with low-dose and standard dose stavudine on adipocyte-specific mtDNA copy number, gene expression and other metabolic parameters. In addition, data on anthropometry and body fat distribution, serological markers of inflammation, and lipid and glucose metabolism measured over 48 weeks of follow-up were also assessed. This was to be the first clinical trial to analyse early and long term effects of stavudine and tenofovir in a South African HIV-1-infected population and as a consequence indentify any possible associated change in cardiovascular risk factors associated with the use of these drugs in the South African setting.
1.9. References


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CHAPTER 2

2. A LONGITUDINAL STUDY OF STAVUDINE-ASSOCIATED TOXICITIES IN A LARGE COHORT OF SOUTH AFRICAN HIV INFECTED SUBJECTS.

2.1. Abstract

Background:
There has been a major improvement in the survival of HIV-1 infected individuals since the South African Government introduced highly active anti-retroviral therapy (HAART) in the public sector in 2004. This has brought new challenges which include the effects of stavudine-related toxicities.

Methods:
Prospective analysis of a cohort of 9040 HIV-infected adults who were initiated on HAART at the Themba Lethu Clinic (TLC) in Johannesburg between April 1, 2004 to December 31, 2007, and followed up until June 30, 2008.

Results:
Amongst the 9040 study subjects, 8497(94%) were on stavudine based therapy and 5962 (66%) were women. The median baseline CD4 count was 81 cells/mm$^3$ (IQR 29-149).

Median follow up on HAART was 19 months (IQR: 9.1-31.6). The proportion of HAART-related side effects for stavudine compared to non-stavudine containing regimens were, respectively: peripheral neuropathy, 17.1% vs. 11.2% (p< 0.001); symptomatic hyperlactataemia, 5.7% vs. 2.2% (p< 0.0005); lactic acidosis, 2.5 vs. 1.3% (p=0.072); lipoatrophy, 7.3% vs. 4.6% (p<0.05). Among those on stavudine-based regimens, incidence rates for peripheral neuropathy were 12.1 cases/100 person-years (95%CI 7.0-19.5), symptomatic hyperlactataemia 3.6 cases/100 person-years (95%CI 1.2-7.5), lactic acidosis
1.6 cases/100 person-years (95%CI 0.4-5.2) and lipoatrophy 4.6 cases/100 person-years (95%CI 2.1-9.6). Females experienced more toxicity when compared to males in terms of symptomatic hyperlactataemia (p< 0.0001), lactic acidosis (p< 0.0001), lipoatrophy (p< 0.0001) and hypertension (p< 0.05).

Conclusions:
We demonstrate significant morbidity associated with stavudine. These data support the latest WHO guidelines, and provide additional evidence for other resource limited HAART rollout programs considering the implementation of non-stavudine based regimens as first line therapy.

2.2. Introduction
By the end of 2008, an estimated 33.4 million people worldwide were living with human immunodeficiency virus (HIV) infection (UNAIDS, 2009). Sub-Saharan Africa continues to bear a disproportionate share of the global burden of HIV with 67% of the HIV infections worldwide, of which 68% of them were amongst adults. This region also accounted for 72% of the world’s AIDS related deaths (UNAIDS, 2009). South Africa’s 2009 HIV prevalence rate in the adult population (aged 15-49 years) was estimated to be 17.8% (Department of Health, 2010).

There has been an increase in the provision of highly active antiretroviral therapy (HAART), with up to 44% of adults and children estimated to be receiving therapy with a profound reduction in mortality (Department of Health, 2010) and, with adherence to HAART it is possible to transform HIV from a fatal infection to a chronic and manageable illness (Hogg et al, 1997. Palella et al, 1998. Wit et al, 1999).
However, some of the anti-retroviral agents used in HAART regimens have severe side effects. Prominent amongst these drugs is stavudine, the use of which is associated with lactic acidosis/symptomatic hyperlactataemia, lipoatrophy, and peripheral neuropathy (Boulle et al, 2007). Other side effects of stavudine use include dyslipidaemia and insulin resistance, and it is an independent risk factor for the development of new onset diabetes mellitus (De Wit et al, 2008). Despite these serious side effects up to 60% of HIV-positive patients in low and middle income countries are receiving stavudine (Beck et al, 2006. Renaud-Théry et al, 2007).

The aim of this study was to make use of a large (N=9040) clinic-based survey of HIV-positive patients newly initiated onto HAART over a period of three years, to compare and describe the effect of stavudine based therapy on acute and chronic toxicities after initiation of HAART. We present baseline data gathered before the initiation of HAART and data collected for three years with each patient having a minimum of six months of follow-up. This is the largest study of the side effects of stavudine therapy conducted to date in a South African HIV-positive population.

2.3. Methods

2.3.1. Study population

The study population included HIV-1 infected individuals attending the Themba Lethu Clinic, a public sector HAART rollout facility based at the Helen Joseph Hospital, a teaching hospital attached to the University of the Witwatersrand, Johannesburg, South Africa. This clinic provides free antiretroviral therapy and other specialized services, and is one of the largest HAART rollout clinics in Africa. The program is funded by the South African
National and Gauteng Departments of Health, with support from Right to Care funded by USAID and PEPFAR.

2.3.2. Treatment

HAART was initiated in accordance with the 2004 South African National Antiretroviral Treatment Guidelines, which include initiation criteria of a CD4 count $\leq 200$ cells/mm$^3$ or WHO stage 4 AIDS defining illness irrespective of CD4 count (National Department of Health, 2004). The first line therapy consisted of stavudine, lamuvidine and efavirenz or nevirapine; however kaletra (ritonavir / lopinavir) was used as part of the first line therapy regimen if there were contra-indications to other first line drugs (National Department of Health, 2004). Until October 2007, stavudine was dosed according to patients’ body weight: 30mg for those <60kg and 40mg for those $\geq$60kg. From October 2007, a universal 30mg dose was introduced and 40mg tablets of stavudine were withdrawn from the clinic. Single drug substitutions were permitted depending on the underlying clinical presentation of the patient.

2.3.3. Clinical and laboratory measurements

Patients who met the criteria for initiation of HAART received adherence counseling and screening for opportunistic infections prior to initiation of therapy. A history and physical examination was performed at every visit. All patients had a baseline chest X-ray. Laboratory monitoring was performed according to the clinic protocol, these included a CD4 count, a viral load, a full blood count (FBC), a creatinine and an alanine aminotransferase (ALT). Other serum biochemical tests were carried out as clinically indicated. Lipid levels and glucose were checked in any patient who presented with features suggestive of the lipodystrophy syndrome and diabetes. The results of these tests were not available for
analysis as the majority of patients were seen and diagnosed at other clinics where they would present for acute medical problems, and where their HAART regimen was modified. However, their diagnoses and new HAART regimens were captured for this study.

2.3.4. Diagnosis and definitions

Body mass index (BMI) was defined as body weight divided by the height, squared (kg/m²). Patients who presented with symptoms of numbness or dysesthesia after initiation of HAART were defined as having peripheral neuropathy due to HAART, once other causes were excluded. Symptomatic hyperlactataemia was defined as the presence of suggestive symptoms with an uncuffed venous lactate level > 5 mmol/L with no evidence of a metabolic acidosis; and lactic acidosis was defined as an uncuffed lactate > 5 mmol/l and arterial pH < 7.35 or a total venous CO₂ < 20mmol/l, with other causes such as sepsis, renal failure, diabetic ketoacidosis and dehydration excluded. As a result, milder cases of hyperlactataemia were excluded by this definition. Pancreatitis was defined as the presence of abdominal symptoms with a serum amylase >125 U/L and lipase >60 U/L. The definition of lipoatrophy was based on the development of peripheral fat wasting (face, arms, buttocks or thighs) and/or central abdominal fat accumulation, and may include enlarged breasts. This was usually reported by the patient and confirmed by the doctor or, initially diagnosed by the doctor with patient confirmation. Hypertension was defined by the presence of three separate readings of systolic blood pressure > 140 mmHg and a diastolic blood pressure >90 mmHg. Diabetes was defined as the presence of symptoms with a fasting glucose of >= 7 mmol/L or a random blood glucose of > 11.1mmol/L. Dyslipidaemias were defined by the presence of abnormal lipid levels which included an elevated total cholesterol of > 5mmol/L, an elevated
triglyceride level of > 1.7mmol/L, and elevated LDL level of >3mmol/L. Only cases that were severe enough to warrant a regimen change were noted with certain toxicities.

2.3.5. **Data collection and statistical analysis**

We analyzed prospectively collected longitudinal cohort data from patients attending the clinic. Clinical data from patient records were captured onto an electronic database via a medical management software system, Therapy Edge-HIV™ (Associated Biological Systems, South Africa). Data was analyzed using the SAS® 9.1 statistical software package (SAS Institute, Inc., North Carolina, USA). Baseline characteristics of the study sample were summarized using simple proportions and medians with interquartile ranges. Differences in proportions of the toxicities were compared by initiating regimen with Chi-squared tests. Study subjects were followed from HAART initiation to the earliest of 1) death; 2) loss to follow up; 3) development of toxicity or 4) censor date 31 December 2008. Incidence rates with 95% confidence intervals for toxicity were calculated and compared by initiating regimen type (stavudine-based versus other). Crude and adjusted estimates of the effect of stavudine use on development of incident toxicity were estimated using Cox proportional hazard models. Models were controlled for confounding by baseline body mass index, CD4 count, age, and gender. Though the majority of subjects initiated HAART prior to the introduction of the universal 30mg stavudine dose, models were also adjusted for time period in which HAART was initiated (prior to or post October 2007). Kaplan Meier curves were used to estimate crude time to diagnosis of therapy-related complications stratified by initiated regimen. Subjects with existing toxicity at initiation of HAART were excluded from these analyses, including a peripheral neuropathy, of which the exact number is unavailable.
Use of the data for the study was approved by the Human Research Ethics Committee (Medical) of the University of the Witwatersrand.

2.4. Results

Between 1 April 2004 and 1 July 2008, a total of 15,928 HIV-infected adults enrolled in care at the Themba Lethu Clinic. Of these, 5104 (32%) had early stage HIV infection and did not qualify for HAART, whilst the remaining 10,824 (68%) subjects were initiated on HAART. The study sample included the 9040 patients initiated on treatment between 1 April 2004 and 31 December 2007. The cohort profile is summarized in Figure 1.

2.4.1. Baseline characteristics of the study population

The baseline characteristics of the study cohort are summarized in Table 1. Two thirds of the study group was female. The majority of patients were in the age group 25-44 years. Even though 46.9% of the total study population were defined as being at only WHO Stage 1 for AIDS, they were already on HAART, indicating that their CD4 count rates were below 200 cells/mm$^3$ and this was confirmed by the median baseline CD4 count of 81 cells/mm$^3$ (IQR: 29-149). All patients in this study cohort were HAART naive at baseline. Ninety four percent of patients were on stavudine based regimens at baseline, with 79% of these initiated on stavudine, lamivudine and efavirenz as per the 2004 South African National guidelines (National Department of Health, 2004), while a smaller number were initiated on zidovudine (3.4%), this was because of contraindications to use of stavudine – particularly peripheral neuropathy. Less than one percent were initiated on a tenofovir-based regimen because of either anaemia or a peripheral neuropathy. Patients with a pre-existing peripheral neuropathy were not included in the data analysis.
2.4.2. Retention in care

The median time to follow up for this cohort on HAART was 19 months (IQR: 9.1-31.6). At the end of the study period, a total of 6415 patients (71%) were still alive and in care, 469 were confirmed deceased and a further 2156 were considered lost to follow up. Patients were considered lost to follow up if they missed their last scheduled appointment by more than 90 days or at least 180 days had lapsed since their last visit. It can be assumed that some of the patients who are lost to follow up may have died and so in total, there were 2,625 (16.5%) patients in this cohort who were either lost to follow up or dead by the end of the study period (see Figure 1).

2.4.3. Response to HAART

The patients had a median baseline CD4 count of 81 cells/ mm³ (IQR 29-149). There was a significant (p<0.0001) increase in the CD4 count after initiation with a median of 205 cells/mm³ (IQR 132-293) by 6 months on treatment. After six months of therapy, there was no significant change in BMI (means ± SD; 22.4 ± 5.0 and 23.7 ± 5.5 respectively).

2.5. Acute and chronic toxicities

Amongst the 9040 patients on therapy, 2488 patients (27.5%) had one or more incident toxicities recorded after treatment initiation and this lead to a drug switch. In terms of acute HAART-related toxicities, the proportion of patients diagnosed with peripheral neuropathy was significantly higher in the group receiving stavudine-based therapy (17.1% vs. 11.2%; p<0.001) compared to those on non-stavudine based therapy (Table 2).
Figure 1. Profile of the Study Cohort

15928 HIV-infected patients

5104 patients had early stage HIV infection and did not require therapy
638 patients were treatment experienced or on second line therapy

10186 HIV-infected patients on HAART

1146 patients initiated HAART outside the study period

9040 HIV-infected patients initiated HAART at TLC during the study period

6415 Survived
2156 Lost to follow up
469 Died
Table 1. Cohort characteristics at baseline

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Baseline value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td>(N= 9040)</td>
</tr>
<tr>
<td>Female</td>
<td>5962 (66)</td>
</tr>
<tr>
<td>Male</td>
<td>3078 (34)</td>
</tr>
<tr>
<td>Age (years)</td>
<td>(N=9040)</td>
</tr>
<tr>
<td>&lt;25 years</td>
<td>551 (6.1)</td>
</tr>
<tr>
<td>25-34 years</td>
<td>3822 (42.3)</td>
</tr>
<tr>
<td>35-44 years</td>
<td>3216 (35.5)</td>
</tr>
<tr>
<td>45-54 years</td>
<td>1173 (13.0)</td>
</tr>
<tr>
<td>&gt;55 years</td>
<td>277 (3.1)</td>
</tr>
<tr>
<td>WHO AIDS classification</td>
<td>(N=8714)</td>
</tr>
<tr>
<td>Stage 1</td>
<td>4086 (46.9)</td>
</tr>
<tr>
<td>Stage 2</td>
<td>1310 (15)</td>
</tr>
<tr>
<td>Stage 3</td>
<td>2507 (28.8)</td>
</tr>
<tr>
<td>Stage 4</td>
<td>811 (9.3)</td>
</tr>
<tr>
<td>HAART</td>
<td>(N = 9040)</td>
</tr>
<tr>
<td>d4T/3TC/EFV</td>
<td>7138 (79)</td>
</tr>
<tr>
<td>d4T/3TC/NVP</td>
<td>690 (7.6)</td>
</tr>
<tr>
<td>d4T/3TC/Kaletra</td>
<td>669 (7.4)</td>
</tr>
<tr>
<td>AZT containing regimen</td>
<td>308 (3.4)</td>
</tr>
<tr>
<td>TDF containing regimen</td>
<td>59 (0.7)</td>
</tr>
<tr>
<td>other</td>
<td>176 (1.9)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>22.4 ±5.0 (N = 7010)</td>
</tr>
<tr>
<td>CD4 count (cells/mm³)</td>
<td>81 (29-149) (N = 7605)</td>
</tr>
<tr>
<td>Follow up time on HAART (months)</td>
<td>19.0 (9.1-31.6) (N= 9040)</td>
</tr>
</tbody>
</table>

Data is expressed as N (%) except for BMI (mean±SD), CD4 count (median, IQR) and follow up time on HAART (median, IQR). WHO, World Health Organization, d4T, stavudine; 3TC, lamivudine; EFV, efavirenz; NVP, nevirapine; kaletra, ritonavir/lopinavir; AZT, zidovudine; TDF, tenofovir.

Peripheral neuropathy was reported equally in both genders (approximately 17%), with no difference in time to development between the drug groups.

There was also a significantly higher proportion of patients on stavudine based therapy presenting with symptomatic hyperlactataemia when compared to those on non-stavudine based therapy (5.75 vs. 2.2%, p<0.0005) (Table 2), with females more frequently affected than males (7.1% vs. 2.5 % ;p<0.0001). Although the rate of development of lactic acidosis was the same for both drug groups, it was experienced more frequently in females than males.
(3.3% vs. 0.8%; p< 0.0001) and this was consistent with other studies (Boulle et al, 2007, Bolhaar et al, 2007, Sanne et al, 2009). The median time to onset was the same for both drug groups. Pancreatitis was equally rare in both treatment groups (see Table 2); both genders were equally affected, with the time to onset being the same in both drug groups. When compared to those on non-stavudine based regimens, those receiving stavudine had higher incidence rates of several toxicities including peripheral neuropathy [12.1/100 person-years (95%CI 7.0-19.5) vs. 7.9/100 person-years (95%CI 6.0-10.1)], symptomatic hyperlactataemia[3.6/100 person-years (95%CI 1.2-7.5) vs.1.4 /100 person-years (95%CI 0.7-2.5)], and lactic acidosis [1.6/100 person-years (95%CI 0.4-5.2) vs. 0.8/100 person-years (95%CI 0.3-1.7)] (Table 3).

In terms of the chronic HAART-related metabolic complications, 7.3% presented with lipoatrophy on stavudine based therapy, compared to 4.6% (p<0.05) patients on non-stavudine based therapy (Table 2). The median time to development of lipoatrophy was similar across the 2 treatment groups. Lipoatrophy was more predominantly seen in female than male patients (10.0% vs. 1.6%; p < 0.0001). Incidence rates for lipoatrophy were slightly higher in the stavudine based therapy group [3.0/100 person-years (95%CI 1.9-4.4) versus 4.6/100 person-years (95%CI 2.1-9.6)] compared to those on other regimens (Table 3). Only two percent of patients presented with hypertension, while only 0.3% presented with diabetes and 1.3% with dyslipidaemia on stavudine based therapy. Similar proportions developing these conditions were observed in both groups (Table 2). However, hypertension developed more quickly (p<0.05) in patients taking stavudine than in those not receiving this drug (Table 2). Hypertension was less common in females than males (1.8% vs. 2.4%; p<0.05) but females and males were equally affected in terms of diabetes and dyslipidaemia.
Table 2. Frequency of and time (months) to HAART associated toxic complications by initiating regimen

<table>
<thead>
<tr>
<th>Condition</th>
<th>Stavudine (N= 8497)</th>
<th>Other drugs (N=543)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Peripheral neuropathy</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>1454 (17.1)*</td>
<td>61 (11.2)*</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>6.7 (3.8-111.8)</td>
<td>5.1 (3.1-11.7)</td>
</tr>
<tr>
<td><strong>Symptomatic hyperlactataemia</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>487 (5.7)*</td>
<td>12 (2.2)*</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>14.5 (10.6-20.9)</td>
<td>11.2 (8.7-19.4)</td>
</tr>
<tr>
<td><strong>Lactic Acidosis</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>214 (2.5)</td>
<td>7 (1.3)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>10.8 (9.0-13.5)</td>
<td>11.2 (9.0-13.5)</td>
</tr>
<tr>
<td><strong>Pancreatitis</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>14 (0.2)</td>
<td>1 (0.2)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>10.4 (4.0-13.0)</td>
<td>8.3 (8.3-8.3)</td>
</tr>
<tr>
<td><strong>Lipoatrophy</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>616 (7.3)**</td>
<td>25 (4.6) **</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>17.0 (11.4-23.1)</td>
<td>15.0 (10.5-24.8)</td>
</tr>
<tr>
<td><strong>Hypertension</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>167 (2.0)</td>
<td>17 (3.1)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>9.7 (4.6-18.4)**</td>
<td>16.3 (9.3-30.5)**</td>
</tr>
<tr>
<td><strong>Diabetes</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>28 (0.3)</td>
<td>1 (0.2)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>22.4 (12.5-28.1)</td>
<td>9.0 (9.0-9.0)</td>
</tr>
<tr>
<td><strong>Dyslipidaemia</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>109 (1.3)</td>
<td>9 (1.7)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>24.6 (17.4-34.0)</td>
<td>19.1 (15.3-26.1)</td>
</tr>
</tbody>
</table>

Data is given as n (%) for prevalence and median (IQR) for time to diagnosis; *p<0.005, **p<0.05 versus group receiving stavudine.

In multivariate analyses adjusted for age, gender, baseline BMI, CD4 counts and the stavudine dose at initiation of HAART, the use of stavudine continued to be associated with increased hazard of developing several toxicities: peripheral neuropathy (HR 2.02; 95% CI 1.35-3.03), symptomatic hyperlactataemia (HR 2.81; 95% CI 1.26-6.31), lactic acidosis (adjusted HR 2.55; 95% CI 0.81-8.00) and diabetes mellitus (adjusted HR 2.07; 95% CI 0.28-15.21) though some of these estimates lacked precision (Table 3).
Figures 2, 3, 4 and 5 present Kaplan Meier curves for the crude estimates of the time to development of HAART-related toxicities by initiating regimen. Those initiated on stavudine based regimens were more likely to develop peripheral neuropathy (log rank p< 0.001), hyperlactataemia (log rank p< 0.001), lactic acidosis (log rank p=0.098) or lipoatrophy (log rank p<0.05) than those on non-stavudine based regimens.

Table 3: Crude and adjusted effects of stavudine use on toxicity initiated on HAART

<table>
<thead>
<tr>
<th>Toxicity</th>
<th>Crude Events</th>
<th>Person Time (years)</th>
<th>Rate/ 100 pys* (95% CI)‡</th>
<th>Crude HR§ (95% CI)‡</th>
<th>Adjusted† HR (95% CI)‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peripheral Neuropathy</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>61</td>
<td>775.9</td>
<td>7.9 (6.0-10.1)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>1454</td>
<td>12055.5</td>
<td>12.1 (7.0-19.5)</td>
<td>1.53 (1.19-1.98)</td>
<td>2.02 (1.35-3.03)</td>
</tr>
<tr>
<td>Symptomatic hyperlactataemia</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>12</td>
<td>852.3</td>
<td>1.4 (0.7-2.5)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>487</td>
<td>13690.9</td>
<td>3.6 (1.2-7.5)</td>
<td>2.70 (1.52-4.79)</td>
<td>2.81 (1.26-6.31)</td>
</tr>
<tr>
<td>Lactic acidosis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>7</td>
<td>848.3</td>
<td>0.8 (0.3-1.7)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>214</td>
<td>13805.9</td>
<td>1.6 (0.4-5.2)</td>
<td>2.09 (0.99-4.44)</td>
<td>2.55 (0.81-8.00)</td>
</tr>
<tr>
<td>Pancreatitis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>1</td>
<td>858.9</td>
<td>0.1 (0.003-0.6)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>14</td>
<td>14126.2</td>
<td>0.1 (0.02-2.6)</td>
<td>0.95 (0.13-7.21)</td>
<td>0.40 (0.05-3.16)</td>
</tr>
<tr>
<td>Lipoatrophy</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>25</td>
<td>834.0</td>
<td>3.0 (1.9-4.4)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>616</td>
<td>13534.3</td>
<td>4.6 (2.1-9.6)</td>
<td>1.67 (1.12-2.50)</td>
<td>1.60 (0.92-2.77)</td>
</tr>
<tr>
<td>Hypertension</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>17</td>
<td>845.0</td>
<td>2.0 (1.2-3.2)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>167</td>
<td>13929.0</td>
<td>1.2 (0.2-4.0)</td>
<td>0.66 (0.40-1.08)</td>
<td>0.83 (0.39-1.78)</td>
</tr>
<tr>
<td>Diabetes Mellitus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>1</td>
<td>859.0</td>
<td>0.1 (0.003-0.6)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>28</td>
<td>14122.7</td>
<td>0.2 (0.02-2.6)</td>
<td>1.90 (0.26-14.04)</td>
<td>2.07 (0.28-15.21)</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Other</td>
<td>9</td>
<td>856.5</td>
<td>1.1 (0.4-2.0)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>d4T-based</td>
<td>109</td>
<td>14097.2</td>
<td>0.8 (0.2-4.0)</td>
<td>0.82 (0.41-1.61)</td>
<td>0.61 (0.27-1.40)</td>
</tr>
</tbody>
</table>

*pys = person years
§ HR = hazard ratio estimated from Cox proportional hazard models
† 95% CI = 95% confidence interval
‡ All models adjusted for age, gender, baseline CD4 count, baseline body mass index and time at which HAART was initiated (either prior to or post October 2007)
2.5.1. Other risk factors for development of toxicity

Age, CD4 counts and WHO staging at baseline did not appear to increase the risk of any adverse events after HAART initiation; however, BMIs at baseline did significantly increase the risk of both symptomatic hyperlactataemia and lactic acidosis. Relative to a baseline BMI of <25 kg/m², an increased risk of symptomatic hyperlactataemia was seen for those with a BMI of 25-30 kg/m² (RR 1.7; 95% CI 1.34-2.14, p< 0.0001) and those with a BMI of >30kg/m² (RR 1.82; 95% CI 1.35-2.44, p< 0.0001). The same was seen with lactic acidosis, where an increased risk was seen for those with a BMI of 25-30 kg/m² (RR 2.52; 95% CI 1.74-3.65, p< 0.0001) and those with a BMI of >30kg/m² (RR 3.10; 95% CI 2.00-4.79, p< 0.0001) when compared to subjects with a BMI of <25kg/m².

Figure 2: Kaplan Meier crude estimates of time to development of peripheral neuropathy by initiating regimen (subjects with peripheral neuropathy at initiation of HAART were not included in the analysis).
Figure 3: Kaplan Meier crude estimates of time to development of hyperlactataemia by initiating regimen.

Figure 4: Kaplan Meier crude estimates of time to development of lactic acidosis by initiating regimen.
2.6. Discussion

This study describes data of three years of follow up of 9040 HIV-infected adults initiated on anti-retroviral treatment at the Themba Lethu Clinic, Johannesburg, South Africa. This high number of patients demonstrates the ability of rolling out a successful HAART programme despite being in a resource limited environment, and this, at a rapidity and scale that compares with reports from other African countries (Boullé et al., 2007. Coetzee et al., 2004 Stringer et al., 2006. Coetzee et al., 2004).

Despite the fact that stavudine is associated with significant complications, a large proportion of low and middle-income countries still use stavudine-based HAART as first line therapy (Beck et al., 2006. Renaud-Théry et al., 2007), mainly because of the cost implications of alternative drugs. In the present study nearly 30% of patients had to switch to non-stavudine based regimens, due to major side effects. These findings concur with those from another
large study in South Africa where 21% of patients switched regimens over a similar time period (Boulle et al, 2007).

This study provides estimates of the pattern of toxicities, the predominant ones being peripheral neuropathy, symptomatic hyperlactataemia, and lipoatrophy. Our incidence rates of peripheral neuropathy (12.1/100 person-years) were much higher compared to other African studies: 5.2/100 person-years in Rwanda (van Griensven et al, 2010) and 2.8/100 person-years in another site in South Africa (Boulle et al, 2007). The variability in these rates could be because there are no grading protocols for the severity of peripheral neuropathies.

Our incidence rates of lactic acidosis were similar to a study from another province in South Africa, 1.6 versus 1.9 /100 person-years (Geddes et al, 2006). The proportion of lactic acidosis was slightly higher compared to another study from Botswana (Wester et al, 2007), 2.5% versus 1% in our cohort. When compared to this same study (Wester et al, 2007), the proportion developing symptomatic hyperlactataemia were much higher in our sample (5.7% versus 2%). These higher rates could be explained by the fact that clinicians were ‘aware’ of the higher proportion of females on stavudine in the clinic and were more ‘sensitized’ to the risk factors and serious side effects associated with stavudine. In addition, we had ready access to laboratory testing for lactic acid determination unlike other resource limited countries. The median time to the development of lactic acidosis was a little later in our study when compared to another study from South Africa (10.8 months versus 7.5 months) (Geddes et al, 2006) .

In terms of the chronic toxicities, incidence rates of lipoatrophy were 4.6 /100 person-years on stavudine based therapy and 3.0/100 person-years on the non-stavudine based therapy. Of note, the potential influence of zidovudine in the non-stavudine group may have affected the ability to determine the differences in rates of lipoatrophy although the numbers were
relatively smaller. One study from Rwanda (van Griensven et al, 2010), where patients were also on stavudine based regimens showed a similar incidence rate of lipoatrophy to that reported in the present study at 4.7/100 person-years, while one other study from South Africa (Boulle et al, 2007) showed a lower incidence rate of 1.4/100 person-years. Another study from Rwanda showed a much higher proportion (34%) of patients developing lipoatrophy (Mutimura et al, 2007). The reason for this wide variation in lipoatrophy rates specifically, is possibly due to different diagnostic criteria used for identifying cases. Thus, in the present study and in the studies showing low but similar rates (Boulle et al, 2007, van Griensven et al, 2010), only cases that were severe enough to warrant a regimen change were noted. In the study showing a much higher proportion, milder cases of lipodystrophy that did not require a regimen change, were also recorded (Mutimura et al, 2007). An objective case definition of lipodystrophy has been developed (Carr et al, 2003); however, it requires access to DEXA and CT imaging technology, which is often not available in resource-limited settings. Therefore, an alternative consensus definition for the diagnosis of lipodystrophy is required for such environments.

Zidovudine may account for the relatively higher rates of mitochondrial toxicities noted in the non-stavudine group, but the degree to which it causes these side effects are not the same as that seen with stavudine. Zidovudine continues to be used as part of the second line therapy by the WHO and the South African HIV therapy guidelines – and is also a commonly used alternative drug because of the lack of other options, especially in the presence of contraindications to tenofovir.

A very small number of patients presented with diabetes (0.3%) and dyslipidaemias (1.3%). However, this could be because lipid or glucose levels were only tested when clinically
suspected due to cost implications, therefore probably underestimating the true prevalence of these metabolic disorders.

The major strength of this study is the large sample size. This cohort is similar to many other resource-limited settings where there is rapid scaling-up of comprehensive HIV care and HAART. It allowed for a relatively long duration of follow up of up to three years and a fairly high retention rate of 70%, with all clinicians working on common protocols for defining the various HAART-associated toxicities. However, despite this, these findings must be considered in the light of potential limitations. It is possible that only severe toxicities that warranted a change in regimen may have been reported and this may have led to an underestimation of the rates of some of the HAART-related toxicities, particularly lipoatrophy. Also, plasma glucose and serum lipid levels were not routinely measured and thus the true rates for glucose intolerance, diabetes and dyslipidaemia were not attained. Lower reported rates of the chronic toxicities could also be related to the rates of death (5%) and loss to follow up (24%), the majority of which occurred within the first six months on treatment as described in a previous report (Sanne et al, 2009).

2.7. Conclusion

These results support the move away from stavudine based regimens towards less toxic combination regimens as advocated by the World Health Organisation (WHO, 2006. WHO, 2010). South Africa has implemented these treatment guidelines, where tenofovir has now replaced stavudine as first line therapy. However in resource-limited countries, where stavudine is still being used because of cost implications, proper pharmacovigilance systems need to be established and alternative consensus definitions and grading protocols are required for identifying various HAART-related toxicities.
2.8. Acknowledgements

Clinical activities at the Themba Lethu Clinic, Helen Joseph Hospital are supported by the South African National and Gauteng Provincial Departments of Health, with additional funding from the United States President’s Emergency Plan for AIDS Relief (PEPFAR) in a grant by USAID to Right to Care and the Helen Joseph Hospital (674-A-00-08-00007-00).

2.9. References


UNAIDS AIDS epidemic update. December 2009:


3. THE EARLY EFFECTS OF STAVUDINE COMPARED TO TENOFOVIR ON ADIPOCYTE GENE EXPRESSION, MITOCHONDRIAL DNA COPY NUMBER AND METABOLIC PARAMETERS IN SOUTH AFRICAN HIV-INFECTED PATIENTS: A RANDOMIZED TRIAL.

3.1. Abstract

Objective:
Stavudine (d4T) is being phased out because of its mitochondrial toxicity and tenofovir (TDF) is recommended as part of first line HAART in South Africa. A prospective, open-label randomized controlled trial comparing standard and low dose d4T with TDF was performed to assess early differences in adipocyte mtDNA copy number, gene expression and metabolic parameters in black South African HIV-infected patients.

Methods:
Sixty patients were randomized 1:1:1 to either standard (30-40 mg) or low (20-30 mg) dose d4T or TDF (300 mg) each combined with lamivudine and efavirenz. Subcutaneous fat biopsies were obtained at weeks 0 and 4. Adipocyte mtDNA copies/cell and gene expression were measured using qPCR. Markers of inflammation, lipid and glucose metabolism were also assessed.

Results:
A 29% and 32% decrease in the mean mtDNA copies/cell was noted in the standard dose ($P<0.05$) and low dose d4T ($P<0.005$) arms respectively when compared to TDF at four weeks. NRF1 and MTCYB gene expression levels were affected by d4T, with a significantly ($P<0.05$) greater fall in expression observed with the standard, but not the low dose compared
to TDF. No significant differences were observed in markers of inflammation, lipid and glucose metabolism.

**Conclusions:**
These results demonstrate early mitochondrial depletion among black South African patients receiving low and standard doses of d4T, with preservation of gene expression levels, except for *NRF1* and *MTCYB*, when compared to patients on TDF.

### 3.2. Introduction

Despite the benefits of the South African Government’s highly active antiretroviral therapy (HAART) roll out programme which commenced in 2004, adverse events remain an important concern for patients on HAART in this country (Boulle *et al.*, 2007, Sanne *et al.*, 2009, Menezes *et al.*, 2011). Understanding the adverse effects of different therapeutic regimens, as well as the underlying molecular and biochemical factors that modulate risk, are critical to improving the management of HIV/AIDS. This is particularly important now that clinical practice is moving towards regimens that combine high levels of both tolerability and efficacy.

Until 2010, South Africa’s treatment guidelines for first-line public-sector HAART recommended stavudine (d4T) with lamivudine (3TC) and either efavirenz (EFV) or nevirapine (NVP) (National Department of Health, 2004). This d4T-based regimen, which is cheap and easy to administer in the short term, is associated with significant morbidity, particularly hyperlactataemia syndromes with long term risks of lipoatrophy, and peripheral neuropathy (Boulle *et al.*, 2007. Sanne *et al.*, 2009. Menezes *et al.*, 2011). The main pathogenic mechanism thought to contribute to the metabolic changes and organ toxicities is mitochondrial toxicity, via the inhibition of mitochondrial DNA (mtDNA) polymerase γ
(POLG) and alterations in messenger RNA (mRNA) gene expression (Mallon et al, 2005. Kim et al, 2008. Sievers et al, 2009. Pace et al, 2003). As a result of the side effects of d4T, South Africa (SA) included tenofovir (TDF) in its recommended first-line public-sector HAART in 2010 (National Department of Health South Africa, 2010), despite its cost implications. A study from South Africa demonstrated that the price of TDF would need to fall from USD$ 17 to USD$ 6.17 per month to have the same overall cost as d4T based therapy (Rosen et al, 2008). TDF has been shown to have a more favourable effect on lipid and mtDNA profiles compared to d4T, with no difference in virologic response (Milinkovic et al, 2007).

While the various toxicities associated with d4T have been well documented, what is less clear is whether the dose of d4T plays a role in the development of these toxicities. The standard dosage for d4T is 40 mg given twice daily as this regimen received regulatory approval because of its efficacy. However, recent studies have suggested that reduced doses of d4T (i.e. 20 or 30 mg twice daily) diminishes its toxicity while maintaining efficacy (Milinkovic et al, 2007. Hill et al, 2007. Gallant et al, 2004).

It is therefore possible that because d4T is cheaper than TDF and its side effects can be minimized via reducing its dose, this would allow more patients to be initiated on HAART in countries where the switch away from d4T may have not yet occurred. Notwithstanding the recent changes in country-level HAART guidelines recommending the use of non-d4T based therapy (British HIV Association guidelines, 2012. Panel on Antiretroviral Guidelines for Adults and Adolescents, 2012), several resource-limited countries have yet to phase out d4T. According to the World Health Organization, in 2010 approximately 56% of HAART regimens within such countries still contained d4T (WHO, 2010).
Therefore, we have conducted an open-label, randomized controlled trial to identify early molecular and biochemical changes and clinical outcomes for patients on either standard or low dose d4T or TDF. We present here the results of four weeks of follow-up which set out to assess if there were any differences in the effect of TDF when compared to low dose and standard dose d4T on adipocyte specific mtDNA copy number, gene expression and other metabolic parameters. To our knowledge this is the first study to analyze the mitochondrial effects of d4T and TDF in a South African HIV-infected population.

3.3. Methods

3.3.1. Study population and inclusion criteria

We enrolled HIV-1 infected, treatment naive individuals aged 18 years or older with a CD4+ cell count below 200 cells/mm³ (consistent with the SA guidelines at the time (National Department of Health, 2004)). Patients were enrolled at the Themba Lethu HIV Clinic at the Helen Joseph Hospital, a public-sector hospital in Johannesburg. Eligible subjects had an absolute neutrophil count (ANC) ≥750/mm, hemoglobin ≥7.0 g/dL and platelet count ≥50,000/mm³. Other inclusion criteria included an alanine and aspartate aminotransferase and an alkaline phosphatase ≤2.5 X upper limit of normal (ULN) and a total bilirubin ≤2.5 X ULN. Patients needed to have a creatinine clearance of ≥60 mL/min using the Cockcroft-Gault formula. In addition, participants of reproductive age had to have a negative serum or urine pregnancy test within 45 days prior to study entry. Those with reproductive potential had to be willing to use at least one reliable form of contraception. Patients were excluded if they were breastfeeding or pregnant and if they had received any antiretroviral drugs (including occupational or sexual post exposure prophylaxis) other than single-dose nevirapine (NVP) within the past six months.
The study was approved by the Human Research Ethics Committee (Medical) of the University of the Witwatersrand and the SA Medicines Control Council (MCC).

3.3.2. Randomization and masking

Patients were randomly assigned 1:1:1 to receive 3TC and EFV in combination with a) a standard dose of d4T (30 mg if weight <60 kg or 40 mg if weight >60 kg) according to the then SA guidelines (National Department of Health, 2004); b) a low-dose of d4T (20 mg if weight <60 kg or 30 mg if weight >60 kg); or c) TDF (300 mg). The definition of the standard and low dose of d4T is similar to several other reported studies (Hill et al, 2007).

Coded identity numbers with the arm allocated were sealed in sequentially numbered envelopes. At enrolment, a member of the data collection team unsealed the next envelope to assign the drug arm allocated to the patient. Those analyzing patient samples were masked to the assignment.

3.3.3. Subcutaneous fat biopsies for the determination of mitochondrial DNA copy number and gene expression

At weeks 0 and 4, subcutaneous fat biopsies from the supra-iliac region were performed under local anesthetic on fasted subjects. Week 4 was chosen to avoid the confounding effects of diet and exercise arising with time. The adipose tissue biopsies were snap frozen in liquid nitrogen prior to storage at -70°C to avoid risk of mtDNA depletion.

Total DNA was extracted at the same time from adipose tissue collected at weeks 0 and 4 for each patient using the QIAmp DNA Mini Kit (QIAGEN, Inc., Hilden, Germany) and a modified protocol (Nolan et al, 2003). Mitochondrial and nuclear gene copy numbers were determined using a quantitative polymerase chain reaction (qPCR) approach and the number
of mitochondria per cell was calculated (Nolan et al., 2003) by comparing the mtDNA content to the nDNA content and by assuming that there are two copies of the nuclear-encoded human growth hormone HGH gene per cell, the number of cells per sample can be calculated and hence the number of mitochondria per cell. Thus, expressed in logarithmic form:

\[
\log_{10}(\text{mtDNA copies/cell}) = \log_{10}(\text{mtDNA copies/ml}) - \log_{10}(\text{nDNA copies/ml}) - \log_{10}(0.5)
\]

Total RNA was extracted using the RNeasy lipid tissue mini kit (QIAGEN), was quantified using a NanoDrop ND-1000 Spectrophotometer (NanoDrop Technologies Inc., Wilmington, Del.) and the integrity assessed using the 2100 Bioanalyser and RNA 6000 picochips (Agilent Technologies, Waldbronn, Germany). A 200 ng aliquot of total RNA was converted into cDNA using the Transcripter high fidelity cDNA synthesis kit with random hexamers (Roche Diagnostics Ltd., Mannheim, Germany). Quantitative real time PCR was performed, in duplicate for each sample, using the SensiMix SYBR Low-ROX kit (Bioline Ltd, Luckenwalde, Germany) on a 7500 Real-Time PCR system (Applied Biosystems, Foster City, Calif.). Each 25 µl reaction mixture contained 1x SensiMix reaction mix, 0.25 µM of each primer and 6.25 ng cDNA. Thermocycling conditions for all primer pairs comprised an initial denaturation step of 95°C for 10 min; 45 amplification cycles of 95°C for 15 sec, 60°C for 30 sec and 72°C for 35 sec and a melt curve step of 95°C for 15 sec, 60°C for 20 sec ramping at 1% to 95°C for 20 sec. Genes assayed included the nuclear genes PPAR-γ coactivator 1α (PGC-1) which is important for mitochondrial biogenesis, nuclear respiratory factor-1 (NRF-1) which is a major respiratory gene transcription regulator and mitochondrial transcription factor-A (TFAM) which binds to and regulates mtDNA. Other genes measured included those involved in mitochondrial energy metabolism - cytochrome c oxidase subunit III (COX 3), cytochrome c oxidase subunit IV (COX 4) and mitochondrial cytochrome B
Two nuclear genes involved in lipid metabolism, leptin (LEP) and lipoprotein lipase (LPL) were also assayed. Primer sequences for these genes were as previously published (Mallon et al., 2005). Gene expression was normalized against three reference genes: β2 microglobulin (B2M) (5’-TGCTGTCTCCATGTTTGATGTATCT-3’ and 5’-TCTCTGCTCCCCACCTCTAAGT-3’), 60S ribosomal protein L 13A (RPL13A) (5’-CCTGGAGGAGAGGAAGGAGA-3’ and 5’-TTGAGGACCTCTGTGTATTTGTCAAA-3’) and hypoxanthine-guanine phosphoribosyltransferase (HPRT) (5’-TGACACTGGCAAAACAATGCA-3’ and 5’-GGTCTTTTCACCAGCAAGCT-3’).

Primers for reference genes were designed using Primer Quest (http://eu.idtdna.com/Scitools/Applications/Primerquest). Comparison of gene expression between the week 0 and week 4 samples were calculated using the 2^ΔΔCq method. In brief, ΔCq values for each patient were calculated for both the genes of interest and the reference genes by subtracting the week 4 sample’s Cq value from the week 0 sample’s Cq value. The relative quantities (2^ΔCq) of the genes of interest and the reference genes were calculated. To normalize to multiple reference genes, the geometric mean of the relative quantities of the reference genes was calculated (Hellemans et al., 2007). The normalized relative quantity of each gene of interest was determined by dividing the relative quantity of the gene of interest by this geometric mean. These normalized relative quantities were log transformed to allow for statistical analysis.

Of note, the patients’ samples at baseline and one month were batched so that the RNA from both time points was extracted at the same time, in addition, the cDNA synthesis was done and the PCRs per gene were also done at the same time. Since samples were compared from the same patient run on the same assay, inter-run variation did not have to be accounted for (i.e. no inter-assay variation for 1 month and baseline samples from each patient).
mtDNA extraction, 100 mg tissue for DNA extraction was used. The PCRs for F8R8 and HGH were run on the same day for the same samples. There was sufficient material to run this experiment on all 60 samples (1 month and baseline). For mRNA extraction, 1 ml lysis reagent per 100 mg tissue was used. Sufficient RNA was also obtained to measure gene expression changes on all 60 samples. The housekeeping genes were not measured per se, they were used to normalise the results. If there was a change in any of the procedures or reagents, it would have been corrected by the use of the housekeeping genes so that the results would represent the changes in gene of interest only.

These results were considered to be robust, as there were several levels of quality control that were employed. The determination of mtDNA copy number was based on a well established protocol (Nolan et al, 2003) that has been standardized and extensively critiqued (Cote et al, 2011). Another important strength of this study was the adoption of the minimum information for publication of quantitative real-time PCR experiments (MIQE) guidelines (Bustin et al, 2009) with the normalization of gene expression to multiple reference genes (Hellemans et al, 2007) which specifically addresses the issue of robustness of data.

3.3.4. Blood measurements

After an overnight fast, serum was taken for the measurement of total cholesterol, high density lipoprotein (HDL)- and low density lipoprotein (LDL) - cholesterol, triglycerides and glucose (Roche Integra analyzer 400, Roche Diagnostics), C-peptide and insulin (Immulite1000 analyzer Diagnostics Corp., Los Angeles, CA), and leptin, adiponectin and hs-CRP (Fluorokine®-Multi analytes profiling (MAP) kit, RnD Systems, Inc). A full blood count was performed and urea, creatinine and electrolytes, liver function test, and CD4 counts (Beckman Coulter flow cytometry, Miami, FL) were also measured. Insulin resistance was
assessed using the homeostasis model assessment (HOMA) method (Matthews et al, 1985) calculated as: (fasting serum insulin (mU/L) X fasting plasma glucose (mmol/L))/22.5. A low HOMA-IR value indicates high insulin sensitivity, whereas, a high HOMA-IR value indicates low insulin sensitivity i.e., insulin resistance.

3.3.5. Statistical analysis

The primary endpoint was to determine if there are significant molecular and biochemical differences between all arms of treatment – with no threshold given to determine the presence of such depletion. Sample size calculations were based on the assumption that at least 70% of participants exposed to d4T would show features of mitochondrial depletion or features of lipodystrophy (Nolan et al, 2003), while the percentage of participants exposed to TDF experiencing these outcomes would be 30%. With the power of 80%, α=0.05, and equal sample size per group, we required 28 subjects per group. Recruitment to the entire trial was stopped early after a data safety and monitoring board (DSMB) analysis of the first 60 patients showed that the group given d4T at both standard and low doses produced a greater fall in the mean mtDNA copies/cell than in the arm receiving TDF. Furthermore, the DSMB considered that there were compelling ethical grounds to stop the high dose d4T because the new South African guidelines were shortly to be implemented and TDF had become freely available. Accordingly, all patients on 40mg dose d4T were switched to TDF. Furthermore, all patients were still followed-up until the end of the study period, which was 48 weeks. As a result, because a “per-protocol” analysis was undertaken, the study may be underpowered to detect differences between the three arms, but this did not affect the differences that were observed.
Data were analyzed after creating an anonymized database and calculating differences between week 0 and 4 readings, using STATISTICA® version 10 (StatSoft, Inc.). Outliers, calculated using (lower quartile - 1.5 X interquartile range; upper quartile + 1.5 X interquartile range), were excluded from the analysis as they skewed the results. They were only excluded from the DNA analysis (2 samples) and not the RNA analysis. Comparisons were made both by randomization arm as well as by the dosage of drug given. The mtDNA copy number and gene expression data were analyzed, using an ANOVA test for multiple comparisons. Markers of inflammation, lipid and glucose metabolism analyzed with a Kruskall-Wallis test. Threshold of significance was set at \( P=0.05 \).

3.4. Results

3.4.1. Baseline characteristics of the study population

Ninety-one HIV-infected patients were screened. Sixty patients qualified and consented, and were randomized 1:1:1 to one of three treatment arms between September 2008 and December 2009. The baseline characteristics of the 60 enrolled patients are summarized in Table 1. There was a predominance of females (85%) in this study and 59/60 were black. A total of 78% were WHO stage 1 while 3% of patients were WHO stage 4, despite low CD4 counts. There was no statistically significant difference in the markers of inflammation, lipid and glucose metabolism between the arms at baseline.

3.4.2. Mitochondrial DNA copy numbers and gene expression levels in adipocytes

The effects of HAART on mtDNA copy number and gene expression were assessed in biopsy samples of subcutaneous adipose tissue (figure 1). The mean \( \log_{10} \) mtDNA copies/cell at baseline and week 4 for the standard dose d4T arm (30-40 mg) was \( 2.74 \pm 0.21 \) and \( 2.61 \pm \)
0.31 respectively and for the low dose d4T arm (20-30 mg), it was 2.63 ± 0.23 and 2.46 ± 0.18 respectively. Whilst it was 2.70 ± 0.14 and 2.69 ± 0.19 respectively, for the TDF arm. There was a 29% decrease in the mean mtDNA copies/cell from baseline to four weeks in standard dose d4T arm (30-40 mg) ($P<0.05$), and a 32% decrease in the low dose d4T arm (20-30 mg) ($P<0.005$), when compared to the TDF (300 mg) arm which had only a 4% decrease in the mean mtDNA copies/cell.

With each individual dose of d4T (20 mg, 30 mg and 40 mg), there was also a drop in mean mtDNA copy numbers (22%, 35%, and 31% respectively) versus TDF (300 mg) (4%) at four weeks of HAART (figure 1). The decrease in mtDNA copy number for both the d4T 30 mg and 40 mg doses was significantly higher than TDF 300 mg ($P<0.005$ and $P<0.05$ respectively). The drop in the mtDNA copy number with the d4T 20 mg dose was not significant when compared to TDF ($P=0.40$).

The relative gene expression of $MTCYB$ was significantly lower in the standard dose (30-40 mg) when compared to the TDF (300 mg) arm ($P<0.05$), but not in the low dose (20-30 mg) d4T arm (table 2). There were minimal changes in the expression of $COX3$ and $COX4$ when comparing the difference at four weeks for all three arms.

With respect to the expression of the nuclear genes involved in the regulation of mitochondrial transcription, there were minimal changes in $PGCI$ and $TFAM$ expression at week 4 when compared to baseline for all three arms. There was a statistically significant difference in $NRF1$ expression in the standard dose d4T arm (30-40 mg) when compared to the TDF 300 mg arm ($P<0.05$). This was not observed in the low dose (20-30 mg) d4T arm. Expression of $LPL$ and $LEP$ were not statistically significant at week 4.
When the effects of the individual d4T doses (20, 30 and 40 mg) on gene expression were compared to those with TDF, no significant differences were noted for any of the 8 genes that were studied (data not shown).

Table 1. Patient characteristics at the commencement of HAART

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard dose d4T (30 mg/40 mg) (N=20)</th>
<th>Low dose d4T (20 mg/30 mg) (N=20)</th>
<th>TDF (300mg) (N=20)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Gender</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>16 (80%)</td>
<td>18 (90%)</td>
<td>17 (85%)</td>
</tr>
<tr>
<td>Male</td>
<td>4 (20%)</td>
<td>2 (10%)</td>
<td>3 (15%)</td>
</tr>
<tr>
<td><strong>Age (years)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>31(11)</td>
<td>34(12)</td>
<td>33(10)</td>
</tr>
<tr>
<td><strong>WHO AIDS classification</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stage 1</td>
<td>14 (70%)</td>
<td>17 (85%)</td>
<td>16 (80%)</td>
</tr>
<tr>
<td>Stage 2</td>
<td>3 (15%)</td>
<td>1 (5%)</td>
<td>3 (15%)</td>
</tr>
<tr>
<td>Stage 3</td>
<td>1 (5%)</td>
<td>2 (10%)</td>
<td>1 (5%)</td>
</tr>
<tr>
<td>Stage 4</td>
<td>2 (10%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td><strong>BMI (kg/m²)</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>25 (5)</td>
<td>24 (7)</td>
<td>24 (6)</td>
</tr>
<tr>
<td><strong>CD4 count (cells/mm³)</strong></td>
<td>155 (94)</td>
<td>169 (78)</td>
<td>135 (62)</td>
</tr>
<tr>
<td><strong>Inflammatory markers</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>hs-CRP (pg/L)</td>
<td>0.80 (0.55)</td>
<td>0.66 (0.94)</td>
<td>0.75 (0.61)</td>
</tr>
<tr>
<td>Leptin (pg/ml)</td>
<td>2.70 (15.02)</td>
<td>3.17 (4.33)</td>
<td>5.52 (4.99)</td>
</tr>
<tr>
<td>Adiponectin (pg/ml)</td>
<td>11.78 (6.58)</td>
<td>11.76 (8.43)</td>
<td>13.78 (11.82)</td>
</tr>
<tr>
<td><strong>Markers of lipid and glucose metabolism</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C-peptide (nmol/ml)</td>
<td>0.40 (0.30)</td>
<td>0.50 (0.20)</td>
<td>0.40 (0.25)</td>
</tr>
<tr>
<td>Insulin (mIU/L)</td>
<td>2.85 (4.05)</td>
<td>3.75 (4.65)</td>
<td>3.60 (3.00)</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>3.20 (1.35)</td>
<td>3.40 (1.10)</td>
<td>3.80 (1.10)</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.85 (0.40)</td>
<td>0.80 (0.45)</td>
<td>0.80 (0.40)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.70 (0.35)</td>
<td>0.80 (0.70)</td>
<td>0.80 (0.30)</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>1.90 (1.05)</td>
<td>2.15 (1.00)</td>
<td>2.50 (0.70)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>3.95 (0.75)</td>
<td>4.20 (0.80)</td>
<td>4.20 (0.60)</td>
</tr>
<tr>
<td>HOMA</td>
<td>0.54 (0.96)</td>
<td>0.70 (1.05)</td>
<td>0.73 (0.60)</td>
</tr>
</tbody>
</table>

Data is expressed as median (IQR) except for gender and WHO stage which are expressed as N (%).
Figure 1: Comparison of log change in mtDNA copy number at week 4 by arm and dosage.

A) Comparison of the three study arms, showing a significant difference in the standard (30 mg/40 mg) and low dose d4T (20 mg/30 mg) arms compared to TDF. B) A comparison of various dosages of d4T to TDF shows significant differences in the 30 mg and 40 mg d4T doses compared to TDF. The 20 mg dose was not significantly different ($P=0.4$). The line within the box-and-whisker plot marks the mean; the upper and lower boundaries of the box indicate one standard deviation to either side of the mean and the error bars above and below indicate the minimum and maximum values. There were 8 patients in the 20 mg dose stavudine group, 17 patients in the stavudine 30 mg dose group and 15 patients in the stavudine 40 mg dose group.

3.4.3. Markers of inflammation, lipid and glucose metabolism

Following four weeks of HAART, there were no statistically significant changes in the lipid profile, glucose, insulin, C-peptide and HOMA indices when comparing the three arms and individual doses. No changes were noted in hs-CRP, leptin or adiponectin levels (table 2).

3.5. Discussion

This randomized controlled trial was designed to evaluate the in vivo effects of TDF against those of standard and low dose d4T regimens, with the reference regimen based on the Gilead
903 trial of Gallant et al, 2004. The Gilead 903 trial showed better outcomes and less adverse events attributed to mitochondrial toxicity in patients receiving TDF 300 mg when compared to d4T 40 mg given twice daily (Gallant et al, 2004), but did not report mtDNA and gene expression evaluations. Our study is the first to compare mitochondrial effects of these drugs in a black population and at a time point in HAART therapy before the development of peripheral lipoatrophy.

The major finding in our study is the significant depletion, after only 4 weeks of therapy, of adipocyte mtDNA copy number in black South African HIV infected individuals receiving d4T. No effect of TDF on mtDNA copy number was observed. This study shows that the low dose d4T regimen produced a fall in mtDNA copy number similar to that observed with the standard d4T dose but greater than that seen with TDF. This finding is consistent with the results of other studies performed in non-black populations who had developed peripheral lipoatrophy at the time of sampling and after a longer duration of anti-retroviral therapy (Nolan et al, 2003. Schooley et al, 2002. Shikuma et al, 2001. Mallal et al, 2001. Buffet et al, 2005. Stankov et al, 2010. Boothby et al, 2009). Our results therefore suggest that mitochondrial pathology precedes changes in body fat distribution and occurs very early after the initiation of HAART.

This study also provides evidence of a dose response effect in that higher doses of d4T caused a greater loss of mtDNA after four weeks of HAART. Thus, d4T at a dose of 30 mg and 40 mg produced a greater fall in mtDNA than TDF, whereas the 20 mg dose of d4T was associated with a non-significant decrease in mtDNA, when compared to the TDF arm.

Despite the significant depletion in mtDNA, our study did not demonstrate a significant effect of d4T with regards to the expression of genes associated with mitochondrial energy.
Table 2: Comparison of the three arms with changes in relative gene expression in relation to housekeeping genes, markers of inflammation, lipid and glucose metabolism parameters from baseline at week 4:

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard dose d4T (30 mg/40 mg) (N=20)</th>
<th>Low dose d4T (20 mg/30 mg) (N=20)</th>
<th>TDF (300 mg) (N=20)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Genes involved in mitochondrial energy metabolism</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>COX 4</td>
<td>-0.01±0.34</td>
<td>0.04±0.49</td>
<td>0.07±0.54</td>
</tr>
<tr>
<td>COX 3</td>
<td>-0.11±0.39</td>
<td>0.01±0.37</td>
<td>0.13±0.34</td>
</tr>
<tr>
<td>MTCYB</td>
<td>-0.20±0.43*</td>
<td>-0.11±0.50</td>
<td>0.16±0.42</td>
</tr>
<tr>
<td><strong>Nuclear genes involved in the regulation of mitochondrial transcription</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NRF-1</td>
<td>-0.21±0.32*</td>
<td>0.00±0.51</td>
<td>0.11±0.26</td>
</tr>
<tr>
<td>PGC-1α</td>
<td>-0.23±0.75</td>
<td>-0.03±0.36</td>
<td>-0.02±0.57</td>
</tr>
<tr>
<td>TFAM</td>
<td>-0.05±0.23</td>
<td>0.08±0.26</td>
<td>0.02±0.24</td>
</tr>
<tr>
<td><strong>Genes involved in lipid metabolism</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>LEP</td>
<td>0.09±0.34</td>
<td>0.09±0.40</td>
<td>-0.06±0.36</td>
</tr>
<tr>
<td>LPL</td>
<td>0.06±0.22</td>
<td>0.08±0.29</td>
<td>-0.01±0.20</td>
</tr>
<tr>
<td><strong>Inflammatory markers</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>hs-CRP (pg/L)</td>
<td>0.22 (0.39)</td>
<td>0.15 (0.43)</td>
<td>0.11 (0.33)</td>
</tr>
<tr>
<td>Leptin (pg/ml)</td>
<td>0.18 (2.62)</td>
<td>0.59 (2.20)</td>
<td>-0.13 (2.89)</td>
</tr>
<tr>
<td>Adiponectin (pg/ml)</td>
<td>0.86 (5.29)</td>
<td>2.12 (5.33)</td>
<td>1.53 (6.52)</td>
</tr>
<tr>
<td><strong>Markers of lipid and glucose metabolism</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C-peptide (nmol/ml)</td>
<td>0.00 (0.20)</td>
<td>-0.10 (0.10)</td>
<td>0.00 (0.20)</td>
</tr>
<tr>
<td>Insulin (mU/L)</td>
<td>-0.25 (5.00)</td>
<td>0.05 (4.70)</td>
<td>-0.60 (3.30)</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>0.60 (0.65)</td>
<td>0.60 (1.00)</td>
<td>0.60 (0.60)</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.10 (0.25)</td>
<td>0.20 (0.30)</td>
<td>0.20 (0.30)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.20 (0.40)</td>
<td>0.10 (0.30)</td>
<td>0.20 (0.30)</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>0.35 (0.75)</td>
<td>0.40 (0.40)</td>
<td>0.10 (0.70)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>0.30 (0.70)</td>
<td>0.00 (1.10)</td>
<td>0.20 (1.00)</td>
</tr>
<tr>
<td>HOMA</td>
<td>-0.05 (1.12)</td>
<td>-0.08 (0.88)</td>
<td>-0.17 (0.69)</td>
</tr>
</tbody>
</table>

Data expressed as means ± SD except for markers of inflammation, lipid and glucose metabolism which are expressed as median (IQR); *p<0.05 versus TDF (300 mg).
metabolism and biogenesis, apart from \textit{MTCYB} and \textit{NRF-1}, although a step-wise decrease in some of the target genes – \textit{COX 4}, \textit{COX 3}, \textit{MTCYB} and \textit{NFF-1}, suggested toxicity related to a dose response, these trends did not reach statistical significance possibly as a consequence of the low sample size. However, these findings are in contrast to those of Mallon \textit{et al.} who found significant reductions in gene expression without significant depletion of mtDNA after two weeks of HAART although notably, they compared d4T with zidovudine (AZT) and their study was in HIV negative patients (Mallon \textit{et al}, 2005). Significant changes in the expression levels of \textit{NRF1} and \textit{MTCYB} were observed in the subjects given the standard d4T dose but not in those receiving the low dose therapy. \textit{NRF1} is an important regulator of mitochondrial biogenesis and function. A master transcription factor, \textit{NRF1} binds the regulatory regions of genes encoding subunits of the respiratory complex and various constituents of the mtDNA transcription and replication machinery. In addition, it serves to coordinate nucleo-mitochondrial activities by regulating mitochondrial and cytosolic enzymes (Scarpulla \textit{et al}, 2008). These results suggest that downregulation of \textit{NRF1} is an early event in NRTI toxicity. This change in \textit{NRF1} expression is then cascaded to the factors which are under its control, with effects that are only observed much later in the time course of HAART (Pace \textit{et al}, 2003. Sievers \textit{et al}, 2009. Boothby \textit{et al}, 2009).

Similar to our results, changes in \textit{MTCYB} expression have been observed in other studies (Kim \textit{et al}, 2008. Stankov \textit{et al}, 2010). These changes may be associated with increased oxidative stress and apoptosis related to mitochondrial depletion (Kim \textit{et al}, 2008. Komarov \textit{et al}, 2008). The HIV-1 infection itself can also lead to alterations in gene expression for mitochondrial proteins and may have played a role here. The expression of several nuclear and mtDNA-encoded genes for mitochondrial proteins have been shown to be reduced in adipose tissue from HIV-1-infected patients who were not on antiretroviral therapy (Casula \textit{et al}}
In a recent study, an African mtDNA subhaplogroup L1c was implicated for the first time in susceptibility to NRTI associated toxicity. Studies comparing mitochondrial toxicity in African versus Caucasian subjects are limited. This is because of the complexities associated with mtDNA from African population. As a result, not much is known about functional differences in African mitochondria due to mtDNA variation (Canter et al, 2010).

Our findings must be considered in light of the study’s limitations. Firstly, the study was stopped prematurely because of the introduction of the new South African National Treatment Guidelines which may have reduced the statistical power of the study. Our participant sample size however, was much larger than other studies where only 20 and 15 samples were analysed, respectively (Mallon et al, 2005. Kim et al, 2008), and was still sufficient for us to be able to detect significant effects of HAART on mtDNA copy number and gene expression. Another possible limitation is the short duration of therapy (4 weeks) at the time of sampling. Other studies have shown significant mtDNA and mtRNA depletion in HIV infected patients on a longer duration of anti-retroviral therapy of months to years (Pace et al, 2003. Sievers et al, 2009). In this study, the fourth week was chosen to avoid the potential confounding effects of changes in diet, exercise or bodyweight.

Despite these limitations, our findings are strengthened by the fact that the laboratory technique used to determine mtDNA copy number was based on a well established protocol (Nolan et al, 2003) that has been standardized and extensively critiqued (Cote et al, 2011). Another important strength of this study was the adoption of the minimum information for publication of quantitative real-time PCR experiments (MIQE) guidelines (Bustin et al, 2009) with the normalization of gene expression to multiple reference genes (Hellemans et al, 2007).

In conclusion, this study demonstrates an early association between mitochondrial depletion and d4T therapy in the black South African population and shows that TDF has a minimal effect on mitochondrial numbers. The expression levels of only 2 of 8 adipocyte genes were significantly affected by d4T therapy when compared with TDF, and this only with the standard dose (30 and 40 mg). These results support the new South African HAART guidelines which have evolved since 2004, with TDF 300 mg currently being recommended as first line therapy (National Department of Health South Africa, 2010). However, minimal effects on gene expression were noted with low dose (20-30 mg) d4T. Therefore, further larger studies using this regimen should be initiated, particularly in light of data showing that this dose is effective and safe (Hill et al, 2007. Milinkovic et al, 2007) and furthermore, it is unlikely that d4T will ever be completely phased out of use in countries with limited financial resources.

3.6. Acknowledgements

We are grateful to the patients who participated in this study and to the staff of the Themba Lethu Clinic at Helen Joseph Hospital, and the Clinical HIV Research Unit, including Dr Thapelo Matoe, Mrs Melissa Hero, Dr Francesca Condradie, Dr Mohammed Rassol, Dr Sharlaa Badal –Faesen, Dr Faizel Laher, Mrs Marlene Naidoo, Ms Doreen Schulze and Mrs
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3.7. References


Stankov MV, Lücke T, Das AM, Schmidt RE, Behrens GM. Mitochondrial DNA depletion and respiratory chain activity in primary human subcutaneous adipocytes treated with


CHAPTER 4

4. A RANDOMIZED CLINICAL TRIAL COMPARING METABOLIC PARAMETERS AFTER 48 WEEKS OF STANDARD AND LOW DOSE STAVUDINE, AND TENOFOVIR THERAPY IN HIV-INFECTED SOUTH AFRICAN PATIENTS.

4.1. Abstract

Objective:
Low dose stavudine therapy may have a lower toxicity profile compared to standard dose. A randomized controlled trial comparing these two doses with tenofovir was performed to assess the effects on anthropometry, markers of inflammation, lipid and glucose metabolism in black South African patients.

Methods:
Sixty patients were randomized 1:1:1 to either standard (30-40 mg) or low dose stavudine (20-30 mg) or tenofovir (300 mg), each combined with lamivudine and efavirenz for 48 weeks. Anthropometry, markers of inflammation, lipid and glucose metabolism were assessed using standard techniques.

Results:
In all three treatment arms, there was a significant increase in lipid levels over the study period. At 48 weeks, fasting glucose ($P<0.005$) and HOMA ($P<0.05$) increased significantly in the standard dose stavudine arm as did insulin and C-peptide levels in both the standard and low dose stavudine arms. At week 48, a significant decrease ($P<0.05$) in adiponectin was noted in the standard dose stavudine arm, but an increase ($P<0.005$) in the tenofovir arm. In both the stavudine arms significant increases in anthropometric measures occurred at 24
weeks but these decreased by week 48. Mitochondrial toxicities occurred in both the stavudine arms. Immunological and virological outcomes were similar for all three arms.

**Conclusions:**

This study highlights the occurrence of metabolic abnormalities with both stavudine and tenofovir treatment. Awareness of the potential increased cardiovascular risk should be of concern with the use of both these therapies.

4.2. **Introduction**


Increased risk of myocardial infarction and rates of dyslipidaemia, insulin resistance and diabetes, and alterations in body fat distribution have been described with the use of HAART (Carr *et al.*, 1999. Carr *et al* 2000. Galli *et al.*, 2002. Friis-Møller *et al.*, 2003. Menezes *et al.*, 2011). Changes in leptin and adiponectin levels that are observed in HIV infection may play a major role in the pathogenesis of HIV/HAART-related metabolic syndrome (Lindegaard *et al.*, 2004. Palios *et al.*, 2012). Furthermore, elevated levels of high sensitivity C-reactive protein (hs-CRP), an inflammatory marker, which have been observed in HIV-infected male patients (Reingold *et al.*, 2008), are associated with an increased risk of cardiovascular events in the general population (Rutter *et al.*, 2004. Pai *et al.*, 2004) as well as in HIV-positive patients (Triant *et al.*, 2009).
Previously, the etiology of these metabolic abnormalities has been attributed to the use of protease inhibitors (PIs) (Carr et al, 1999). More recently, nucleoside reverse transcriptase inhibitors (NRTIs) have also been implicated, particularly stavudine (Boulle et al, 2007. Mutimura et al, 2007. van Griensven et al, 2010. Menezes et al, 2011). Stavudine continues to be used as alternative first-line treatment in developing countries despite the change in the WHO antiretroviral therapy guidelines for treating patients with HIV infection (WHO, 2010). Stavudine was used as first line HIV therapy in South Africa until 2010 and replaced with tenofovir (National Department of Health, South Africa, 2004 & 2010), but it continues to be used because of shortages of tenofovir and abacavir that occur intermittently at health facilities across South Africa (Schowalter et al, 2012). Compared to stavudine, tenofovir has fewer side effects and increases lipid levels to a lesser degree, with no difference in virological response (Gallant et al, 2004).

The recommended dose for stavudine is 40 mg twice daily, but studies have shown that lower doses (20 or 30 mg twice daily) diminishes toxicity while maintaining immunological efficacy (Hill et al, 2007. Milinkovic et al, 2007. McComsey et al, 2008). Therefore, in resource limited countries where the switch away from stavudine to the more expensive tenofovir has not yet occurred, low dose stavudine could still be used, allowing more patients to be initiated on to treatment.

To determine whether low dose stavudine therapy was as effective and safe as tenofovir in a resource-poor environment, we performed an open-label, randomized controlled trial. Biochemical changes and clinical outcomes for patients on either low or standard dose stavudine were compared to those patients receiving tenofovir.
We present here data on anthropometry and body fat distribution, serological markers of inflammation, and lipid and glucose metabolism measured over 48 weeks of follow-up in a cohort of black South African HIV-positive patients.

4.3. Methods

4.3.1. Study population

HIV-1 infected individuals aged 18 years or older with a CD4+ cell count below 200 cells/mm$^3$ were enrolled in this study at the Themba Lethu HIV Clinic at the Helen Joseph Hospital, Johannesburg, South Africa. Inclusion criteria were: an absolute neutrophil count (ANC) $\geq 750$/mm$^3$, a hemoglobin $\geq 7.0$ g/dL, a platelet count $\geq 50,000$/mm$^3$, a creatinine clearance of $\geq 60$ mL/min calculated using the Cockcroft-Gault formula (Cockcroft et al, 1976), and an alanine and aspartate aminotransferase, an alkaline phosphatase and a total bilirubin of $\leq 2.5 \times$ upper limit of normal. Patients of reproductive age had to have a negative serum or urine pregnancy test within 45 days prior to study entry, and had to be willing to use at least one reliable form of contraception. Exclusion criteria were: breastfeeding or pregnancy, receiving antiretroviral drugs including occupational or sexual post-exposure prophylaxis, other than single-dose nevirapine within the previous six months.

Patients were seen at screening and at week 0, 4, 24 and 48. Routine evaluations were taken at each time point, whilst markers of inflammation, lipid and glucose metabolism were measured at week 0, 4 and 48. These evaluations included a review of adverse events and concomitant medications, complete or symptom-directed physical examination, height and weight, hematology and chemistry profiles, calculated creatinine clearance, CD4+ cell count, plasma HIV-1 RNA and study drug accountability. Dual-energy radiographic absorptiometry (DEXA) scanning was performed at baseline and week 48.
Approval was received from the Human Research Ethics Committee (Medical) of the University of the Witwatersrand and the SA Medicines Control Council (MCC). This study was compliant with the revised CONSORT statement guidelines (www.consort-statement.org).

4.3.2. Anti-retroviral therapy

As previously described (Menezes et al, 2012), patients were assigned in random 1:1:1 permuted blocks of five to receive lamivudine and efavirenz in combination with a) standard dose stavudine (30 mg if weight < 60 kg or 40 mg if weight > 60 kg) according to the then SA guidelines (National Department of Health, South Africa, 2004); b) low-dose stavudine (20 mg if weight < 60 kg or 30 mg if weight > 60 kg); or c) tenofovir 300 mg. The definition of the standard and low dose of stavudine is the same as that used in several other studies (Hill et al, 2007). Coded identity numbers with the arm allocated were sealed in sequentially numbered envelopes. At enrolment, a member of the data collection team unsealed the next envelope to assign the drug arm allocated to the patient. Staff analyzing patient samples were blinded to the treatment assigned to each patient.

4.3.3. Blood measurements

Fasting serum was taken for the measurement of total cholesterol, high density lipoprotein (HDL)-, and low density lipoprotein (LDL)-cholesterol, triglycerides and glucose (Roche Integra analyzer 400, Roche Diagnostics, Mannheim, Germany), C-peptide and insulin (Immuleite1000 analyzer Diagnostics Corp., Los Angeles, CA, USA), and leptin, adiponectin and hs-CRP (Fluorokine®-Multianalyte profiling (MAP) kit, RnD Systems, Inc., Minneapolis, MN, USA). A full blood count was performed and creatinine, liver function
tests, viral load (Roche Cobas Amplicor, Indianapolis, IN) and CD4+ count (Beckman Coulter flow cytometry, Miami, FL, USA) were measured. Insulin resistance was assessed using the homeostasis model assessment (HOMA) method (Matthews et al, 1985).

4.3.4. **Anthropometry and body composition measurements**

Height and weight were measured using regularly calibrated instruments. Circumferences of the mid arm, chest, waist, hip and mid-thigh were obtained using a Gulick II measuring tape. Skinfolds of the triceps, biceps, subscapular, and suprailiac were measured using a Harpenden skin fold caliper. For all anthropometry measurements, two independent measurements were taken. If the first two measurements were outside of the acceptable range of variance for a particular body area, then a third measurement was taken. The mean of the two closest measures was then used in analysis. Total lean and fat mass were measured with a Hologic whole-body, dual-energy radiographic absorptiometry (DEXA) scanner (Hologic Inc., Waltham, MA, USA) and software, version 12.4. Staff analyzing performing the DEXA scans were blinded to the treatment assigned to each patient but those performing the anthropometry measurements were not.

4.3.5. **Statistical analysis**

Data that could be normalized was log transformed to normality and analyzed using parametric statistical analyses, whilst the remaining variables were analyzed using non-parametric statistical tests. With a power of 80%, $\alpha = 0.05$, and equal sample size per group, we required 28 patients per treatment arm, assuming that 70% of patients exposed to stavudine and 30% exposed to tenofovir would show features of mitochondrial toxicity (Nolan et al, 2003). However, the trial was stopped early after a data safety and monitoring
board (DSMB) analysis of the first 60 patients showed that the group given stavudine at both standard and low doses produced greater mitochondrial depletion than in the arm receiving tenofovir (Menezes et al, 2012). Furthermore, the DSMB considered that there were compelling ethical grounds to stop the high dose stavudine because the new SA guidelines were shortly to be implemented (National Department of Health, South Africa 2010), and tenofovir had become freely available. At that point, the four patients who were still on 40 mg of stavudine were switched to tenofovir. Those patients who switched therapy or who, for any other reason, had to prematurely leave the study were excluded from the final analysis. Therefore, the trial was not adequately powered to demonstrate non-inferiority in terms of efficacy. Data were analyzed using STATISTICA® version 10 (StatSoft, Inc., Tulsa, OK, USA). Results were summarized using proportions and medians with interquartile ranges. Comparisons were made at different time intervals within each arm using the Wilcoxon matched paired test or Students paired t test, whilst comparisons across arms were performed using ANOVA for baseline levels. The week 4 and week 48 data were analysed using ANOVA and ANCOVA, with adjustment for baseline levels. This adjustment had no effect on the outcomes.

4.4. Results

4.4.1. Baseline characteristics of the study population

Figure 1 shows the cohort profile: 60 eligible patients were randomized 1:1:1 to the three treatment arms between September 2008 and December 2009. There was a predominance of females (85%) in this study and 59 of the 60 patients were black (Table 1). All patients were clinically asymptomatic with 78% of the patients being WHO stage 1.
4.4.2. **Lipid metabolism**

There were significant increases in both total and HDL-cholesterol levels from baseline to weeks 4 and 48 for all three arms (Table 2). Triglyceride levels increased significantly at week 4 in the standard stavudine and tenofovir arms only. The LDL-cholesterol increased significantly in the low dose stavudine arm at weeks 4 and 48, but increased only at week 4 for the standard dose stavudine arm and at week 48 for the tenofovir arm (Table 2). The percentage of patients with hypercholesterolemia, hypertriglyceridemia, elevated LDL and reduced HDL in each arm, as defined by NCEP-ATP III guidelines (2002), were calculated at week 48 and no significant differences were found.

4.4.3. **Glucose metabolism**

A decrease in C-peptide levels was noted at week 4 and a significant increase by week 48 for the low dose stavudine arm, with a significant increase at week 48 also noted in the standard stavudine arm.

When comparing the fasting insulin levels in all patients, an increase was noted at week 48 in both stavudine arms but not in the tenofovir arm. Significant differences were noted when comparing the change in insulin levels at week 48 from baseline between the standard dose stavudine and tenofovir arm (Table 3).

When comparing the HOMA scores, a significant increase was noted at week 48 for the standard stavudine arm only.

4.4.4. **Inflammatory markers**

There was a significant increase in the levels of hs-CRP at week 4 in the standard dose stavudine arm and in the leptin levels at week 4 for the low dose stavudine arm, and at week
Figure 1: Profile of the Trial Cohort

Enrollment
- Assessed for eligibility (N=91)
  - Excluded (N=31)
    - Not meeting inclusion criteria (n=30)
    - Withdrew (n=1)
  - Randomized (N=60)

Allocation
- Allocated to Standard dose stavudine (30/40mg), lamivudine, efavirenz. (N=20)
- Allocated to Low dose stavudine (20/30 mg), lamivudine, efavirenz. (N=20)
- Allocated to Tenofovir (300mg) lamivudine, efavirenz. (N=20)

Follow up
- Discontinued therapy (N=12)
  - Lost to follow-up (n=1)
  - Withdrew (n=1)
  - Pregnancy (n=1)
  - Death (n=1)
  - Advice (DSMB) (n=4)
  - Serious adverse events (n=4)
- Discontinued therapy (N=1)
  - Serious adverse events (n=1)
- Discontinued therapy (N=1)
  - Serious adverse events (n=1)

Analysis
- Analyzed (N=8)
  - Excluded from analysis (n=12)
    (reasons above)
- Analyzed (N=19)
  - Excluded from analysis (n=1)
    (reasons above)
- Analyzed (N=19)
  - Excluded from analysis (n=1)
    (reasons above)
Table 1. Patient characteristics at the commencement of HAART

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard dose stavudine (30 mg/40 mg) (N=20)</th>
<th>Low dose stavudine (20 mg/30 mg) (N=20)</th>
<th>Tenofovir (300 mg) (N=20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>African race</td>
<td>20 (100%)</td>
<td>19 (95%)</td>
<td>19 (95%)</td>
</tr>
<tr>
<td>Gender, Female</td>
<td>16 (80%)</td>
<td>18 (90%)</td>
<td>17 (85%)</td>
</tr>
<tr>
<td>Age (years)</td>
<td>31.0 (11.0)</td>
<td>34.0 (12.0)</td>
<td>33.0 (10.0)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>25 (5)</td>
<td>24 (7)</td>
<td>24 (6)</td>
</tr>
<tr>
<td>Hemoglobin (g/d)</td>
<td>12.3 (2.7)</td>
<td>12.3 (2.2)</td>
<td>12.0 (2.0)</td>
</tr>
<tr>
<td>GGT (IU/L)</td>
<td>24 (24)</td>
<td>30 (11)</td>
<td>22 (11)</td>
</tr>
<tr>
<td>ALT (IU/L)</td>
<td>21 (21)</td>
<td>20 (15)</td>
<td>18 (12)</td>
</tr>
<tr>
<td>Creatinine clearance (ml/min)</td>
<td>108 (17)</td>
<td>108 (25)</td>
<td>99 (40)</td>
</tr>
</tbody>
</table>

Data are expressed as median (IQR) except for race and gender which are expressed as N (%). BMI: Body Mass Index, GGT: gamma-glutamyltransferase, ALT: alanine aminotransferase.

48 for the tenofovir arm (Table 2). A significant decrease in the adiponectin levels was noted in the standard dose stavudine arm at week 48, and an increase at weeks 4 and 48 in the tenofovir arm. The week 48 adiponectin level was significantly higher in the tenofovir than the standard dose stavudine arm, and this was similar when comparing the change in adiponectin levels over the course of the study between the standard dose stavudine and tenofovir arms (Table 3).

4.4.5. Immunological and virological efficacy

No differences were noted at week 24 in CD4+ cell counts and viral loads. There was a significant increase in the CD4+ cell counts in all three arms at week 48 when compared to baseline (Table 2).
Table 2: Comparison of the markers of inflammation, glucose and lipid metabolism at different time intervals:

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard dose stavudine (30 mg/40 mg)</th>
<th>Low dose stavudine (20 mg/30 mg)</th>
<th>Tenofovir (300 mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Time (week)</td>
<td>0</td>
<td>4</td>
<td>48</td>
</tr>
<tr>
<td>N</td>
<td>20/20</td>
<td>20/20</td>
<td>8/20</td>
</tr>
<tr>
<td>Total Cholesterol (mmol/L)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>3.20</td>
<td>4.10***</td>
<td>4.35*</td>
</tr>
<tr>
<td></td>
<td>(1.35)</td>
<td>(1.35)</td>
<td>(1.20)</td>
</tr>
<tr>
<td>HDL-Cholesterol (mmol/L)</td>
<td>0.85</td>
<td>1.00*</td>
<td>1.35*</td>
</tr>
<tr>
<td></td>
<td>(0.40)</td>
<td>(0.50)</td>
<td>(0.55)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.70</td>
<td>0.90**</td>
<td>0.95</td>
</tr>
<tr>
<td></td>
<td>(0.35)</td>
<td>(0.45)</td>
<td>(0.20)</td>
</tr>
<tr>
<td>LDL-Cholesterol (mmol/L)</td>
<td>1.90+</td>
<td>2.60***</td>
<td>2.35</td>
</tr>
<tr>
<td></td>
<td>(1.05)</td>
<td>(1.15)</td>
<td>(1.20)</td>
</tr>
<tr>
<td>C-peptide (nmol/ml)</td>
<td>0.40</td>
<td>0.50</td>
<td>0.50*</td>
</tr>
<tr>
<td></td>
<td>(0.30)</td>
<td>(0.20)</td>
<td>(0.20)</td>
</tr>
<tr>
<td>Insulin (mU/L)</td>
<td>2.85</td>
<td>3.65</td>
<td>3.95*</td>
</tr>
<tr>
<td></td>
<td>(4.05)</td>
<td>(3.00)</td>
<td>(2.60)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>3.95</td>
<td>4.40</td>
<td>4.40*</td>
</tr>
<tr>
<td></td>
<td>(0.75)</td>
<td>(0.40)</td>
<td>(0.70)</td>
</tr>
<tr>
<td>HOMA</td>
<td>0.54</td>
<td>0.69</td>
<td>0.83*</td>
</tr>
<tr>
<td></td>
<td>(0.96)</td>
<td>(0.81)</td>
<td>(0.59)</td>
</tr>
<tr>
<td>hs-CRP (pg/L)</td>
<td>0.80</td>
<td>0.84*</td>
<td>0.79</td>
</tr>
<tr>
<td></td>
<td>(0.55)</td>
<td>(0.75)</td>
<td>(0.35)</td>
</tr>
<tr>
<td>Leptin (pg/ml)</td>
<td>2.69+</td>
<td>4.09</td>
<td>1.84</td>
</tr>
<tr>
<td></td>
<td>(15.02)</td>
<td>(10.08)</td>
<td>(4.43)</td>
</tr>
<tr>
<td>Adiponectin (pg/ml)</td>
<td>11.78</td>
<td>13.53*</td>
<td>7.21*+</td>
</tr>
<tr>
<td></td>
<td>(6.58)</td>
<td>(4.08)</td>
<td>(3.86)</td>
</tr>
<tr>
<td>CD4 count (cells/mm³)</td>
<td>155 (94)</td>
<td>ND</td>
<td>285 *</td>
</tr>
<tr>
<td></td>
<td>(88650)</td>
<td></td>
<td>(52)</td>
</tr>
<tr>
<td>Viral load, (copies/ml)</td>
<td>150000</td>
<td>ND</td>
<td>102*</td>
</tr>
<tr>
<td></td>
<td>(88650)</td>
<td></td>
<td>(2)</td>
</tr>
<tr>
<td>Viral load, &lt;400 copies/ml, N (%)</td>
<td>-</td>
<td>-</td>
<td>7/8(88%)</td>
</tr>
<tr>
<td>Viral load, &lt;50 copies/ml, N (%)</td>
<td>-</td>
<td>-</td>
<td>7/8(88%)</td>
</tr>
</tbody>
</table>

Data is expressed as median (IQR); *p<0.05; **p<0.005; *** p<0.0005 versus week 0; + p<0.05 versus same time point in tenofovir arm; ND-Not done.
Table 3: Changes from baseline to week 48 in markers of glucose and lipid metabolism, inflammatory markers, and anthropometric measurements in each of the 3 study arms:

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard dose stavudine (30 mg/40 mg) (N=20)</th>
<th>Low dose stavudine (20 mg/30 mg) (N=20)</th>
<th>Tenofovir (300 mg) (N=20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>8/20</td>
<td>19/20</td>
<td>18/20</td>
</tr>
<tr>
<td>C-peptide (nmol/ml)</td>
<td>0.1 (0.3)</td>
<td>0.1 (0.3)</td>
<td>0.1 (0.3)</td>
</tr>
<tr>
<td>Insulin (mU/L)</td>
<td>2.3 (4.4)*</td>
<td>1.4 (5.3)</td>
<td>0.8 (5.1)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>0.6 (1.3)</td>
<td>0.3 (0.7)</td>
<td>0.5 (0.8)</td>
</tr>
<tr>
<td>HOMA</td>
<td>0.5 (1.0)</td>
<td>0.3 (1.2)</td>
<td>0.2 (1.2)</td>
</tr>
<tr>
<td>Total Cholesterol (mmol/L)</td>
<td>0.7 (1.7)</td>
<td>1.4 (2.5)</td>
<td>0.7 (1.6)</td>
</tr>
<tr>
<td>HDL-Cholesterol (mmol/L)</td>
<td>0.3 (0.8)</td>
<td>0.4 (0.8)</td>
<td>0.5 (0.9)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.2 (0.6)</td>
<td>0.1 (0.6)</td>
<td>0.1 (0.3)</td>
</tr>
<tr>
<td>LDL-Cholesterol (mmol/L)</td>
<td>0.2 (0.9)</td>
<td>0.7 (1.5)</td>
<td>0.2 (0.8)</td>
</tr>
<tr>
<td>hs-CRP (pg/L)</td>
<td>0.1 (0.4)</td>
<td>0.1 (0.9)</td>
<td>0.1 (0.5)</td>
</tr>
<tr>
<td>Leptin (pg/ml)</td>
<td>0.8 (4.7)</td>
<td>1.4 (11.2)</td>
<td>1.9 (13)</td>
</tr>
<tr>
<td>Adiponectin (pg/ml)</td>
<td>-0.6 (6.8)*</td>
<td>0.1 (12.4)</td>
<td>3.6 (3.5)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>0.5 (1.8)</td>
<td>1.3 (2.6)</td>
<td>0.5 (4.0)</td>
</tr>
<tr>
<td>Mid-arm (cm)</td>
<td>1.5 (2.5)</td>
<td>0.0 (3.0)</td>
<td>0.5 (5.1)</td>
</tr>
<tr>
<td>Waist (cm)</td>
<td>2.0 (7.0)</td>
<td>4.0 (9.0)</td>
<td>2.8 (8.6)</td>
</tr>
<tr>
<td>Hip (cm)</td>
<td>3.0 (11)</td>
<td>1.0 (6.8)</td>
<td>1.4 (10.8)</td>
</tr>
<tr>
<td>Mid-thigh (cm)</td>
<td>2.0 (6.0)</td>
<td>2.0 (5.3)</td>
<td>2.0 (6.0)</td>
</tr>
<tr>
<td>Triceps (mm)</td>
<td>4.0 (17.6)</td>
<td>0.1 (11.4)</td>
<td>6.0 (24.3)</td>
</tr>
<tr>
<td>Subscapular (mm)</td>
<td>3.0 (17.6)</td>
<td>3.6 (6.9)</td>
<td>-0.9 (16.4)</td>
</tr>
<tr>
<td>Suprailiac (mm)</td>
<td>3.6 (18.6)</td>
<td>5.8 (11.5)</td>
<td>2.1 (7.1)</td>
</tr>
<tr>
<td>Trunk Fat (g)</td>
<td>1565.2 (3939.9)</td>
<td>1467.1 (4172)</td>
<td>470.2 (2935.9)</td>
</tr>
<tr>
<td>Trunk % Fat</td>
<td>4.10 (11.8)</td>
<td>3.20 (7.1)</td>
<td>1.20 (7.3)</td>
</tr>
<tr>
<td>Tot Limb Fat (g)</td>
<td>0.0 (5700.9)</td>
<td>-68.4 (2754.6)</td>
<td>0.0 (3126.5)</td>
</tr>
<tr>
<td>Tot Limb % Fat</td>
<td>-1.5 (7.9)</td>
<td>-0.9 (3.9)</td>
<td>0.0 (3.6)</td>
</tr>
</tbody>
</table>

Data expressed as median (IQR); p value by Kruskal Wallis; *p<0.05 versus Tenofovir.

Similarly, there was a significant decrease in viral loads (≤ 400 copies/ml) noted in all three arms at week 48 when compared to baseline but number of patients who had virological suppression at week 48 did not differ significantly between treatment arms (Table 2).

An ITT analysis was performed for CD4 counts and viral loads at week 48 and no significant differences were noted.
4.4.6. **Anthropometric measurements and body composition**

There was a significant increase in the BMI of patients in the standard dose stavudine arm by weeks 4 and 24 and in the low dose stavudine arm by weeks 24 and 48; although no significant changes were observed for patients in the tenofovir arm (Table 4). A significant increase was noted in the waist measurements at weeks 24 and 48 in both the standard and low dose stavudine arms but only at week 48 in the tenofovir arm. A significant increase in hip measurements was noted by week 24 in both stavudine arms but this was not observed with tenofovir. A similar trend was noted in the mid thigh measurements, but with an increase at week 48 for the tenofovir arm.

With regards to the skinfold measurements, a similar increasing trend was noted at week 24 for the two stavudine arms with a tendancy for measures to fall by week 48 to levels close to, or below those at baseline. For the tenofovir arm, there was an increase in skinfold measurements across the study period but changes were not statistically significant. (Table 4) Exactly 47% of the patients in the standard stavudine dose arm, 40 % in the low dose stavudine arm, and 55% in the tenofovir arm \( (P=0.637) \) developed a BMI \( \geq 25\text{kg/m}^2 \) at week 48.

4.4.7. **Safety and therapy switching**

There were 14 adverse events (AEs) in the standard stavudine dose arm, 17 AEs in the low dose stavudine arm and 12 AEs in the tenofovir arm. There was one case of renal dysfunction, in the tenofovir arm. There was one death on the standard stavudine dose arm due to a high grade B cell lymphoproliferative disorder. One patient was lost to follow up, one patient withdrew from the study and one patient fell pregnant and was withdrawn from the study.
With regards to mitochondrial AEs, there were three patients in the standard stavudine dose arm who developed a lactic acidosis while one patient developed a hyperlactatemia. In the low dose stavudine arm, three patients developed a peripheral neuropathy, of which one of these patients also had an associated hyperlactatemia and clinical features of lipoatrophy warranting switch in therapy.

Within the standard dose stavudine arm, 12 patients had to switch therapy for various reasons. At week 0, these subjects differed from those who did not change therapy in terms of higher measures of body fat, hsCRP and leptin levels. Notably, at baseline 66.7% of patients who switched therapy were overweight/obese compared to 12.5% of patients who did not switch therapy ($P<0.05$). A significant difference was noted when comparing the median (IQR) suprailiac skinfold thickness (43.3 mm (25.3) vs. 16 mm (12.9); $P<0.005$), mean truncal % fat ($32.98 \pm 10.23 \text{ vs. } 21.44 \pm 9.93; P<0.05$), hs-CRP (0.86 pg/L (0.53) vs. 0.54 pg/L (0.64); $P<0.05$), and leptin levels (8.74 pg/ml (26.57) vs. 1.37 pg/ml (1.62); $P<0.005$) for those who switched therapy in standard dose stavudine arm to those who did not.

4.5. Discussion

Despite the move away from stavudine to towards less toxic regimens, more than 1.55 million people were still receiving stavudine therapy by the end of 2012 (WHO, 2010). It is known that even though stavudine is cheaper and effective as initial therapy, the standard dose (30 or 40 mg) is associated with long term complications. However, studies have demonstrated excellent antiviral efficacy with a lower dose of 20 mg (Hill et al, 2007. Milinkovic et al, 2007. McComsey et al, 2008). Generic tenofovir is used as a non-stavudine first line therapy but is more expensive (Rosen et al, 2008). This study was a prospective, open-label randomized controlled trial designed to evaluate the in vivo effects of tenofovir
Table 4: Comparison of the anthropometric measurements and body fat composition at different time intervals:

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard dose stavudine (30 mg/40 mg)</th>
<th>Low dose stavudine (20 mg/30 mg)</th>
<th>Tenofovir (300 mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 weeks 4 weeks 24 weeks 48 weeks</td>
<td>0 weeks 4 weeks 24 weeks 48 weeks</td>
<td>0 weeks 4 weeks 24 weeks 48 weeks</td>
</tr>
<tr>
<td>N</td>
<td>20/20 20/20 19/20 8/20</td>
<td>20/20 20/20 20/20 19/20</td>
<td>20/20 20/20 20/20 19/20</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.82 (4.92) 24.87* (4.62) 24.84** (5.65) 24.69 (4.64)</td>
<td>23.45 (6.50) 23.92 (6.20) 25.48*** (5.50) 23.79* (5.61)</td>
<td>24.25 (5.68) 24.08 (4.44) 24.88 (5.32) 25.06 (6.36)</td>
</tr>
<tr>
<td></td>
<td>Mid-arm (cm)</td>
<td></td>
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<tr>
<td></td>
<td>28.75 (4.50) 29.00 (3.75) 30.5** (4.50) 31.0* (2.50)</td>
<td>28.75 (6.50) 28.00 (7.00) 30.0* (7.00) 29.00 (5.50)</td>
<td>29.00 (4.75) 29.50 (4.75) 30.00 (4.50) 31.50 (8.00)</td>
</tr>
<tr>
<td></td>
<td>Chest (cm)</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>81.75 (8.00) 81.50 (6.50) 83.00 (11.00) 82.00 (12.00)</td>
<td>79.00 (6.25) 79.00 (11.50) 79.50 (8.20) 80.50* (7.00)</td>
<td>79.75 (7.50) 80.75 (10.50) 81.00 (8.75) 81.00 (10.00)</td>
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<tr>
<td></td>
<td>Waist (cm)</td>
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<td></td>
<td>82.00 (8.00) 81.00 (9.00) 83.00* (11.00) 83.00* (14.50)</td>
<td>75.75 (12.50) 81.00 (13.00) 80.50** (14.75) 82.00** (13.00)</td>
<td>78.75 (13.50) 78.00 (12.50) 83.75 81.50* (12.25) (15.00)</td>
</tr>
<tr>
<td></td>
<td>Hip (cm)</td>
<td></td>
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<tr>
<td></td>
<td>100.50 (13.00) 102.50 (12.00) 105.00* (10.00) 102.00 (11.50)</td>
<td>99.50 (16.25) 99.00 (21.00) 101.50** (14.00) 98.00 (9.00)</td>
<td>100.00 (14.50) 101.00 (16.00) 103.50 (12.00) 109.00 (16.00)</td>
</tr>
<tr>
<td></td>
<td>Mid-thigh (cm)</td>
<td></td>
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<tr>
<td></td>
<td>57.00 (8.75) 58.00 (6.75) 59.50** (7.00) 57.00 (7.00)</td>
<td>55.25 (8.75) 54.00 (13.00) 56.00* (8.50) 56.50 (5.00)</td>
<td>59.50 (10.00) 60.00 (13.50) 61.25 (9.50) 63.00* (12.00)</td>
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<tr>
<td></td>
<td>Triceps skinfold (mm)</td>
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<tr>
<td></td>
<td>43.70 (37.80) 49.80 (22.50) 63.00* (33.20) 43.40 (33.20)</td>
<td>43.50 (29.70) 51.00 (30.40) 50.70*** (27.10) 39.80 (20.40)</td>
<td>46.50 (29.90) 46.10* (33.30) 52.50 (27.00) 54.00 (50.50)</td>
</tr>
<tr>
<td></td>
<td>Biceps skinfold (mm)</td>
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<tr>
<td></td>
<td>16.00 (10.10) 16.80 (8.20) 18.60 (14.00) 12.40 (9.00)</td>
<td>18.00 (13.20) 19.40 (15.00) 18.00 (13.40) 13.2* (10.80)</td>
<td>15.10 (11.40) 16.50 (9.40) 20.10 (10.90) 18.00 (10.80)</td>
</tr>
<tr>
<td></td>
<td>Subscapular skinfold (mm)</td>
<td></td>
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<tr>
<td></td>
<td>32.80 (23.90) 30.50 (22.00) 36.00 (25.40) 34.70 (17.70)</td>
<td>25.80 (16.30) 29.60 (18.00) 31.80*** (18.60) 29.00 (21.00)</td>
<td>30.40 (16.90) 32.90 (18.70) 35.70 (20.00) 36.60 (23.40)</td>
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<tr>
<td></td>
<td>Suprailiac skinfold (mm)</td>
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<tr>
<td></td>
<td>27.00 (28.60) 27.40 (26.90) 27.60* (31.40) 25.30 (14.80)</td>
<td>25.80 (23.10) 26.00 (26.10) 30.30* (19.20) 27.6* (18.00)</td>
<td>28.60 (24.70) 27.60 (25.30) 30.00 (22.50) 33.00 (28.40)</td>
</tr>
<tr>
<td></td>
<td>Trunk fat (g)</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>17687.70 (1903.50) 19759.00 (14212.70)</td>
<td>17538.70 (13110.20) 19490.45 (12170.20)</td>
<td>23000.10 (10254.40) 24924.90 (17053.00)</td>
</tr>
<tr>
<td></td>
<td>Trunk % fat</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td>28.10 (19.50) 30.80 (19.60)</td>
<td>29.40 (18.00) 33.90 (12.80)</td>
<td>34.60 (11.70) 37.10 (18.00)</td>
</tr>
<tr>
<td></td>
<td>Total limb fat</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>16878.60 (12021.10) 21512.15 (13510.75)</td>
<td>16138.95 (16609.90) 17378.30 (12214.90)</td>
<td>21961.00 (10268.60) 24105.20 (17061.50)</td>
</tr>
<tr>
<td></td>
<td>Total limb % fat</td>
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<td></td>
</tr>
<tr>
<td></td>
<td>28.90 (21.00) 37.60 (23.30)</td>
<td>30.40 (15.10) 35.00 (13.70)</td>
<td>35.70 (12.40) 40.45 (17.60)</td>
</tr>
</tbody>
</table>

Data is expressed as median (IQR); * p<0.05; **p<0.005; *** p<0.0005 versus week 0; ND-Not done.
against those of standard and low dose stavudine regimens, with the reference regimen based on the Gilead 903 trial by Gallant et al (Gallant et al, 2004). The Gilead 903 trial showed better outcomes and less adverse events attributed to mitochondrial toxicity in patients receiving tenofovir 300 mg when compared to patients receiving 40 mg stavudine given twice daily (Gallant et al, 2004). To our knowledge, this work presents the first trial to compare the use of these drugs in a black population.

Studies in treatment naïve HIV infected patients have shown that peripheral fat initially increases during the first few months of HAART, but decreases linearly thereafter and truncal fat increases around six months of therapy (Mallon et al, 2003. Magkos et al, 2011). Similarly, in the current study significant increases in the majority of anthropometric measurements were noted in the stavudine arms by six months with a drop towards one year of therapy, whilst such changes were seen in a minority of the anthropometric variables with tenofovir therapy.

This study demonstrated that insulin and C-peptide levels increased significantly in both stavudine arms, but not with tenofovir therapy. However, significant rises in glucose and HOMA levels were only observed with the standard stavudine dose. Previous research showed that the severity of lipoatrophy, among patients taking either stavudine or zidovudine, was associated with an increased risk of insulin resistance (De Wit et al, 2008).

Consistent with other studies (Savès et al, 2002. Gallant et al, 2004. Shlay et al, 2005), we demonstrated a significant increase in fasting lipids after one year of HAART, and this was noted within both stavudine arms and with tenofovir. Lipid abnormalities have been noted in HIV-1 infected patients even before the advent of antiretroviral therapy, with cholesterol levels being lower and triglyceride levels higher in HIV-infected compared to non-infected subjects (Constans et al, 1994). In the present study, the early increase of cholesterol levels
after only 4 weeks of therapy may be explained by the 'return to baseline' effect of commencing antiretroviral therapy, but this cannot explain the rise in triglyceride levels. Furthermore, dyslipidaemia has been widely reported with exposure to NRTIs, particularly with stavudine or zidovudine, and less so with tenofovir (Savè et al., 2002. Mallon et al., 2003. Gallant et al., 2004. Shlay et al., 2005. De Wit et al., 2008. Magkos et al., 2011). The observed effects might also be associated with the use of efavirenz. However, its effects on lipids remain largely conflicting (van Leth et al., 2004. Jones et al., 2005).

Inflammatory markers appear to have a role in the pathogenesis of HIV/HIVART related metabolic syndrome. The effect of HAART on leptin levels is controversial; a few studies have demonstrated that low levels of leptin are associated with lipodystrophy while other studies have demonstrated no effect of HAART on leptin (Dzwonek et al., 2007. Calmy et al., 2009). We noted an increase in leptin levels at week 4 for patients in the low dose stavudine arm and at week 48 for the tenofovir arm. Patients on HAART, especially those with lipodystrophy, show a gradual reduction in serum adiponectin levels, which is thought to be associated with increased cardiovascular risk (Bezante et al., 2009). In our study, there was a significant decrease at week 48 in the standard dose stavudine arm but a significant increase in the tenofovir arm at 4 and 48 weeks. These changes in serum adiponectin levels in the stavudine arm may be significant in terms of the ongoing risk of lipoatrophy as well as metabolic complications. hs-CRP has been shown to be a marker of increased cardiovascular events (Pai et al., 2004). There was a tendency for hs-CRP levels to increase in all 3 treatment arms, but this reached statistical significance only in the standard dose stavudine group at 4 weeks and then fell to baseline levels at 48 weeks.

The AEs associated with mitochondrial toxicities were noted only with the stavudine regimens. However, none of these were seen in individuals treated with the 20 mg dose.
Our findings must be considered in light of the study’s limitations. Our sample size was small as the study had to be stopped prematurely because of the mitochondrial results (Menezes et al., 2012) and due to the introduction of the new South African National HAART guidelines. Secondly, despite this being a randomized controlled trial, it was open labeled, which may have been a confounding factor. The metabolic abnormalities may be more prevalent with increasing duration of therapy and therefore not seen after only 48 weeks of therapy. However, these data are important as our study is the first randomized trial assessing the frequency of NRTI-associated metabolic toxicities related to the two most relevant antiretroviral agents currently used in a routine clinical setting in black African populations.

In conclusion, the current study demonstrates that in South African HIV-positive patients, tenofovir has more favorable effects on anthropometry and adipokine levels whilst both stavudine regimens increase fasting insulin and C-peptide levels, with the higher stavudine dose also causing increased fasting glucose and higher HOMA levels. However, both stavudine and tenofovir cause raised lipid levels. Therefore, awareness of the potential increased cardiovascular risk should be of concern with the use of both these therapies. More AEs were noted in the standard compared to the low dose stavudine arm. Therefore, where options are limited and there is an absolute need for stavudine to be used, low dose stavudine may be a cautious option as it is equally efficient in viral suppression and has lower rates of complications than the standard dose.

4.6. Acknowledgements

We are grateful to the patients who participated in this study and staff of the Themba Lethu Clinic, Clinical HIV Research Unit and Department of Radiology at Helen Joseph Hospital.
We would like to acknowledge CIPLA SA for their donation of stavudine and lamivudine for the duration of this study.

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CHAPTER 5

5. DISCUSSION AND CONCLUSION

5.1. Summary of the results

There is growing concern regarding the rise in the rates of non-communicable diseases, especially cardiovascular disease, that has occurred in South Africa over the last 15 years. This is likely to increase further in the coming years, with HIV-1 infection and its associated complications, compounding this problem (Mayosi et al, 2009).

Whilst HIV-1 infection has been turned into a chronic, manageable disease, the serious acute mitochondrial and long term metabolic side effects caused by HAART are a major concern especially with the use of stavudine. This drug was used as first line HIV therapy in South Africa until 2010 (National Antiretroviral Treatment Guidelines, 2004). Our clinic-based longitudinal study (Menezes et al, 2011) of 9040 HIV-infected adults, demonstrated the concerns associated with stavudine based therapy. Although this study demonstrated a fairly high retention rate of 70%, nearly 30% of the patients had to switch to non-stavudine based regimens, due to major side effects. Those patients receiving stavudine had higher incidence rates of acute toxicities which included peripheral neuropathy [12.1/100 person-years (95% CI 7.0-19.5) vs. 7.9/100 person-years (95% CI 6.0-10.1)], symptomatic hyperlactataemia [3.6/100 person-years (95% CI 1.2-7.5) vs. 1.4 /100 person-years (95% CI 0.7-2.5)], and lactic acidosis [1.6/100 person-years (95% CI 0.4-5.2) vs. 0.8/100 person-years (95% CI 0.3-1.7)] when compared to those on non-stavudine based regimens.

In terms of the metabolic complications, lipoatrophy was seen in 7.3% of patients on stavudine based therapy compared to 4.6% (p<0.05) patients on non-stavudine based therapy with a very small proportion of patients presented with diabetes and dyslipidaemia.
According to the WHO, more than 1.55 million people will still be receiving stavudine therapy by the end of 2012 (WHO, 2010). Because stavudine is cheaper than tenofovir and its side effects can be minimized by a dose reduction, it is therefore possible that more patients could be initiated on antiretroviral therapy in countries where the switch has still not occurred. In addition, zidovudine which is used as part of second line therapy or as an alternative drug in the presence of contraindications to tenofovir, also causes mitochondrial side effects but to a lesser extent than stavudine (van Griensven et al, 2007). Our clinical trial (Menezes et al, 2012) was the first randomized trial that assessed the mitochondrial pathogenesis of NRTI-associated toxicities in an African population when comparing the use of various doses of stavudine and tenofovir therapy.

A major finding of this study is the significant depletion of adipocyte mtDNA in black South African HIV-1 infected individuals on both high and low dose stavudine therapy after only four weeks of therapy. We also showed that tenofovir had a minimal effect on mitochondrial numbers. There was a 29% and 32% decrease in the mean mtDNA copies/cell in the standard-dose and low-dose stavudine arms, respectively, when compared with tenofovir at four weeks. We also noted that despite the significant depletion in mtDNA, the expression levels of only two of eight adipocyte genes (MTCYB and NRF-1) associated with mitochondrial energy metabolism and biogenesis were significantly affected by standard dose (30 and 40 mg) stavudine therapy when compared with tenofovir, with minimal effects on gene expression with low dose (20-30 mg) stavudine.

Data after one year of therapy demonstrated that both stavudine and tenofovir caused raised lipid levels but surprisingly this was also noted as early as four weeks of therapy. Early insulin resistance was noted with both stavudine regimens with data suggesting that both
stavudine regimens increased fasting insulin and C-peptide levels, with the higher stavudine dose also causing an increased fasting glucose and higher HOMA levels. Tenofovir was noted to have a more favourable effect on anthropometry by one year of therapy. Notably, in both the stavudine arms significant increases in anthropometric measures occurred at 24 weeks but these decreased by week 48. This fat redistribution was not objectively confirmed on DEXA scanning in this study possibly because only total truncal and total limb fat was measured.

We noted an increase in leptin levels after four weeks of therapy for patients in the low dose stavudine arm and after 48 weeks of therapy for the tenofovir arm. In our study, there was a significant decrease in serum adiponectin levels at week 48 in the standard dose stavudine arm but a significant increase in the tenofovir arm at 4 and 48 weeks. The hs-CRP serum levels demonstrated a tendency to increase in all three treatment arms, but this reached statistical significance only in the standard dose stavudine group at 4 weeks and then fell to baseline levels at 48 weeks.

This trial demonstrated that there was a significant increase in the CD4+ cell counts in all three arms after one year of therapy when compared to baseline, and similar levels of virological suppression (≤ 400 copies/ml).

The adverse events associated with mitochondrial toxicities were noted only with the stavudine regimens but none were seen in individuals treated with the 20 mg dose.

5.2. Discussion of the results

The first study (Chapter 2) demonstrated the ability of a resource limited clinic to roll out a successful HAART programme at a pace and scale that compared with other African
countries (Boulle et al, 2007. Coetzee et al, 2004. Stringer et al, 2006). This was the largest such study conducted in a South African HIV-1-positive population.

When compared to other studies from Africa (Boulle et al, 2007. van Griensven et al, 2010. Mutimura et al, 2007), there was a wide variation in the rates of lipoatrophy; the reason for this is possibly due to the use of different diagnostic criteria. It would appear that only cases that were presenting with severe enough body dysmorphic changes to warrant a regimen change were noted in our study. The study from Rwanda, showed a significantly higher proportion of lipodystrophy because milder cases of lipodystrophy that did not warrant a regimen change, were also recorded (Mutimura et al, 2007). The low rates of diabetes and dyslipidaemias may be because lipid or glucose levels were only tested when clinically suspected due to cost implications, probably underestimating the true prevalence of dyslipidaemia and diabetes in our clinic. Other reasons could be that the patients may have died (death rates of 5%) or lost to follow up (24%), the majority of which occurred within the first six months on therapy.

In addition to the disparity in the rates of toxicities, this study highlighted other issues associated with the management of HIV-1 infected patients in resource limited environments, such as the importance of close monitoring for adverse events and when to anticipate such problems. There is a need for grading protocols for assessing the severity of peripheral neuropathies, as well as an alternative definition for the diagnosis of lipodystrophy considering the lack of access to routine DEXA and MRI imaging technology. Routine metabolic monitoring such as glucose and lipid profiles must also be instituted.

With regards to the clinical trial, these findings were consistent with studies undertaken in non-African populations who developed peripheral lipoatrophy at the time of sampling and after a longer duration of anti-retroviral therapy (Boothby et al, 2009. Buffet et al, 2005.
Our results suggest that mitochondrial pathology precedes changes in body fat distribution and occurs very early after the initiation of antiretroviral therapy. In addition, \textit{NRF1} is known to be an important regulator of mitochondrial biogenesis and function, where it binds to the regulatory regions of genes encoding subunits of the respiratory complex and various constituents of the mtDNA transcription and replication machinery. It serves to coordinate nucleo-mitochondrial activities by regulating mitochondrial and cytosolic enzymes (Scarpulla \textit{et al}, 2008). These results suggest that downregulation of \textit{NRF1} is an early event in NRTI toxicity. This change in \textit{NRF1} expression triggers a cascade of events under its control, with effects that are only observed much later in the time course of antiretroviral therapy (Boothby \textit{et al}, 2009. Pace \textit{et al}, 2003. Sievers \textit{et al}, 2009).

Similarly, changes in \textit{MTCYB} expression have been observed in other studies (Kim \textit{et al}, 2008. Stankov \textit{et al}, 2010). These changes may be associated with increased oxidative stress and apoptosis related to mitochondrial depletion (Kim \textit{et al}, 2008. Komarov \textit{et al}, 2008).


These findings are strengthened by the fact that the laboratory technique used in this study to determine mtDNA copy number was based on a well-established protocol (Nolan \textit{et al}, 2003) that has been standardized and extensively critiqued (Cote \textit{et al}, 2011), and which followed the minimum information for publication of quantitative real-time PCR experiments (MIQE)
guidelines (Bustin et al, 2009) with the normalization of gene expression to multiple reference genes (Hellemans et al, 2007).

In this era where the benefits of HAART have clearly improved the morbidity and mortality of patients with HIV-1 infection, CVD has emerged as a cause for concern in these patients. In our study, data at 48 weeks of therapy demonstrated that in South African, HIV-1 positive patients both stavudine and tenofovir caused raised lipid levels. While dyslipidaemias have been known to be associated with exposure to NRTIs, this has been mainly associated with stavudine and zidovudine, and less so with tenofovir (Gallant et al, 2004. Gallant et al, 2006). Consistent with other studies (Gallant et al, 2004. Savès et al, 2002. Shlay et al, 2005), including studies from Africa (Buchacz et al, 2010. Mutimura et al, 2007. Sinxadi et al, 2010. Zannou et al, 2009), we demonstrated a significant increase in fasting lipids on HAART, but surprisingly this was noted after only 4 weeks of therapy with both stavudine and tenofovir. The observed effects might also be explained by the use of efavirenz, but its effects on lipids remain largely conflicting (van Leth et al, 2004. Jones et al, 2005). Early insulin resistance was noted with both stavudine regimens, and other studies have shown that cumulative exposure to NRTIs such as stavudine and zidovudine is associated with abnormal glucose tolerance (Brown et al, 2005. De Wit et al, 2008. Tien et al, 2008). Tenofovir was noted to have a more favorable effect on anthropometry that stavudine. Studies in treatment naïve HIV infected patients have shown that peripheral fat initially increases during the first few months of HAART, but decreases linearly thereafter and truncal fat increases around six months of therapy (Makgos et al, 2011. Mallon et al, 2003). In fact, studies demonstrated that lipoatrophy tended to be common after one year of therapy and especially in patients who were on higher doses of stavudine (40mg) (Mutimura et al, 2007. van Griensven et al, 2010).
The effect of antiretroviral therapy on leptin levels is controversial; some studies have suggested that low levels of leptin are associated with lipodystrophy while others have demonstrated no effect (Dzwonek et al, 2007. Calmy et al, 2009). We noted early changes in leptin levels in patients on the low dose stavudine arm and late changes in the tenofovir arm.

Patients on HAART, especially those with lipodystrophy, show a gradual reduction in serum adiponectin levels, which is thought to be associated with increased cardiovascular risk (Bezante et al, 2009). We noted a significant decrease after a year of therapy in the standard dose stavudine arm but a significant increase in the tenofovir arm at 4 and 48 weeks. This is an important finding because in non-HIV infected patients low adiponectin levels have been shown to be associated with an increased risk of diabetes and associated metabolic disorders (Koenig et al, 2006).

Our data would suggest that low dose stavudine is more patient friendly than the standard dose, and is equally efficacious in viral suppression and immunological outcomes, and may therefore still be a treatment option in resource limited settings.

These results support the new South African HAART guidelines which have evolved since 2004, with tenofovir 300 mg currently being recommended as first line therapy. This study also demonstrates that in South African, HIV-positive patients both stavudine and tenofovir cause raised lipid levels, although tenofovir has more favourable effects on anthropometry and adipokine levels. Both stavudine regimens increase fasting insulin and C-peptide levels, with the higher stavudine dose also causing increased fasting glucose and higher HOMA levels. This data also suggests that a potential for increased cardiovascular risk exists with the use of both tenofovir and stavudine, and that cardiovascular risk factor modification strategies would be important in the management of patients receiving these therapies.
5.3. Limitations and directions for future studies

1. Even though our sample size was much larger than other studies, and was still sufficient for us to be able to detect significant effects of HAART on mtDNA copy number and gene expression, the trial had to be stopped prematurely on the advice of the DSMB and this may have reduced the statistical power of the study. The DSMB analysis of the first 60 patients showed that the group given stavudine at both standard and low doses produced greater mitochondrial depletion than in the arm receiving tenofovir. Furthermore, the DSMB considered that there were compelling ethical grounds to stop the high dose stavudine because the new SA guidelines were shortly to be implemented (National Department of Health, South Africa 2010), and tenofovir had become freely available. At that point, the four patients who were still on 40 mg of stavudine were switched to tenofovir. The other patients were left on the 20 mg and 30mg stavudine doses, because it took some time before tenofovir was eventually available. Therefore there were 20 patients in the standard dose stavudine arm at baseline and 4 weeks; and only 8 patients at 48 weeks.

2. Adipose biopsies should be performed after a longer duration of antiretroviral therapy as other studies found significant mtDNA and mtRNA depletion on a longer duration of therapy of months to years.

3. Metabolic abnormalities may be more prevalent with increasing duration of therapy and may not be seen after only 48 weeks of therapy; therefore, further studies using the low dose stavudine (20mg stavudine vs. tenofovir) should be initiated to confirm our findings, particularly in light of data showing that this dose is effective and safe and, furthermore, it is unlikely that stavudine will ever be completely phased out of use in countries with limited financial resources.
4. The impact of HIV/AIDS on CVD needs to be studied further. Whilst a lot is known about the aetiology and complications of the epidemic of CVD in the HIV negative population, there is a paucity of data to describe the emergence and impact of CVD in HIV-1-infected patients on antiretroviral therapy in Africa. The emergence of lipodystrophy with its associated complications such as insulin resistance/diabetes, hypertension and dyslipidaemia will have major public health implications in the future and will put further strain on the already limited financial resources available within the public health sector not only in South Africa, but also across antiretroviral therapy programmes in sub-Saharan Africa.

5.4. References


Stankov MV, Lücke T, Das AM, Schmidt RE, Behrens GM. Mitochondrial DNA depletion and respiratory chain activity in primary human subcutaneous adipocytes treated with


WHO. Forecasting antiretroviral demand. 2010.

A longitudinal study of stavudine-associated toxicities in a large cohort of South African HIV infected subjects

Colin N Menezes1,2,3*, Mhairi Maskew2,3, Ian Sanne2,3, Nigel J Crowther4 and Frederick J Raaij5

Abstract
Background: There has been major improvement in the survival of HIV-1 infected individuals since the South African Government introduced highly active anti-retroviral therapy (HAART) in the public sector in 2004. This has brought new challenges which include the effects of stavudine-related toxicities.

Methods: Prospective analysis of a cohort of 9040 HIV-infected adults who were initiated on HAART at the Thembelihle Clinic (TLC) in Johannesburg between April 1, 2004 to December 31, 2007, and followed up until June 30, 2008.

Results: Amongst the 9040 study subjects, 8497(94%) were on stavudine based therapy and 5962 (66%) were women. The median baseline CD4 count was 81 cells/mm³ (IQR 29-149). Median follow up on HAART was 19 months (IQR 9.1-31.6). The proportion of HAART-related side effects for stavudine compared to non-stavudine containing regimens were, respectively: peripheral neuropathy 17.1% vs. 11.2% (p < 0.001); symptomatic hyperlactataemia, 5.7% vs. 2.2% (p < 0.0005); lactic acidosis, 2.5% vs. 1.3% (p = 0.072); lipatrophy, 7.3% vs. 4.6% (p < 0.05). Among those on stavudine-based regimens, incidence rates for peripheral neuropathy were 12.1 cases/100 person-years (95%CI 7.0-19.5), symptomatic hyperlactataemia 3.6 cases/100 person-years (95%CI 1.2-7.5), lactic acidosis 1.6 cases/100 person-years (95%CI 0.4-5.2) and lipatrophy 4.6 cases/100 person-years (95%CI 2.1-9.6). Females experienced more toxicity when compared to males in terms of symptomatic hyperlactataemia (p < 0.0001), lactic acidosis (p < 0.0001), lipatrophy (p < 0.0001) and hypertension (p < 0.05).

Conclusions: We demonstrate significant morbidity associated with stavudine. These data support the latest WHO guidelines, and provide additional evidence for other resource limited HAART rollout programs considering the implementation of non-stavudine based regimens as first line therapy.

Background
By the end of 2008, an estimated 33.4 million people worldwide were living with human immunodeficiency virus (HIV) infection [1]. Southern Africa continues to bear a disproportionate share of the global burden of HIV with 67% of the HIV infections worldwide, of which 68% of them were amongst adults. This region also accounted for 72% of the world’s AIDS related deaths [1]. South Africa’s 2009 HIV prevalence rate in the adult population (aged 15-49 years) was estimated to be 17.8% [2].

There has been an increase in the provision of highly active antiretroviral therapy (HAART), with up to 44% of adults and children estimated to be receiving therapy with a profound reduction in mortality [2] and, with adherence to HAART it is possible to transform HIV from a fatal infection to a chronic and manageable illness [3-5].

However, some of the anti-retroviral agents used in HAART regimens have severe side effects. Prominent amongst these drugs is stavudine, the use of which is associated with lactic acidosis/symptomatic hyperlactataemia, lipatrophy, and peripheral neuropathy [6]. Other side effects of stavudine use include dyslipidaemia and insulin resistance, and it is an independent risk factor for the development of new onset diabetes mellitus [7]. Despite these serious side effects up to 60% of HIV positive
patients in low and middle income countries are receiving stavudine [8,9].

The aim of this study was to make use of a large (N = 9040) clinic-based survey of HIV-positive patients newly initiated onto HAART over a period of three years, to compare and describe the effect of stavudine based therapy on acute and chronic toxicities after initiation of HAART. We present baseline data gathered before the initiation of HAART and data collected for three years with each patient having a minimum of six months of follow-up. This is the largest study of the side effects of stavudine therapy conducted to date in a South African HIV-positive population.

Methods

Study population

The study population included HIV-1 infected individuals attending the Thembisa Lethu Clinic, a public sector HAART roll out facility based at the Helen Joseph Hospital, a teaching hospital attached to the University of the Witwatersrand, Johannesburg, South Africa. This clinic provides free antiretroviral therapy and other specialized services, and is one of the largest HAART roll out clinics in Africa. The program is funded by the South African National and Gauteng Departments of Health, with support from Right to Care funded by USAID and PEPFAR.

Treatment

HAART was initiated in accordance with the 2004 South African National Antiretroviral Treatment Guidelines, which include initiation criteria of a CD4 count ≤ 200 cells/mm³ or WHO stage 4 AIDS defining illness irrespective of CD4 count [10]. The first line therapy consisted of stavudine, lamuvidine and efavirenz or nevirapine; however, kelastra (ritonavir/lopinavir) was used as part of the first line therapy regimen if there were contra-indications to other first line drugs [10]. Until October 2007, stavudine was dosed according to patients' body weight: 30 mg for those < 60 kg and 40 mg for those ≥ 60 kg. From October 2007, a universal 30 mg dose was introduced and 40 mg tablets of stavudine were withdrawn from the clinic. Single drug substitutions were permitted depending on the underlying clinical presentation of the patient.

Clinical and laboratory measurements

Patients who met the criteria for initiating of HAART received adherence counseling and screening for opportunistic infections prior to initiation of therapy. A history and physical examination was performed at every visit. All patients had a baseline chest X-ray, Laboratory monitoring was performed according to the clinic protocol. Other serum biochemical tests were carried out as clinically indicated. The results of these tests were not available for analysis as the majority of patients were seen and diagnosed at other clinics where they would present for acute medical problems, and where their HAART regimen was modified. However, their diagnoses and new HAART regimens were captured for this study.

Diagnosis and definitions

Body mass index (BMI) was defined as body weight divided by the height, squared (kg/m²). Patients who presented with symptoms of numbness or dysesthesia after initiation of HAART were defined as having peripheral neuropathy due to HAART, once other causes were excluded. Symptomatic hyperlactataemia was defined as the presence of suggestive symptoms with an uncuffed venous lactate level > 5 mmol/L with no evidence of a metabolic acidosis; and lactic acidosis was defined as an uncuffed lactate > 5 mmol/L and arterial pH < 7.35 or a total venous CO2 < 20 mmol/L, with other causes such as sepsis, renal failure, diabetic ketoacidosis and dehydration excluded. Pancreatitis was defined as the presence of abdominal symptoms with a serum amylase > 125 U/L and lipase > 60 U/L. The definition of lipoatrophy was based on the development of peripheral fat wasting (face, arms, buttocks or thighs) and/or central abdominal fat accumulation, and may include enlarged breasts. This was usually reported by the patient and confirmed by the doctor or, initially diagnosed by the doctor with patient confirmation. Hypertension was defined by the presence of three separate readings of systolic blood pressure > 140 mmHg and a diastolic blood pressure > 90 mmHg. Diabetes was defined as the presence of symptoms with a fasting glucose of > 7 mmol/L or a random blood glucose of 11.1 mmol/L. Dyslipidemias were defined by the presence of abnormal lipid levels which included an elevated total cholesterol of > 5 mmol/L, an elevated triglyceride level of > 1.7 mmol/L, and elevated LDL level of > 3 mmol/L.

Data collection and statistical analysis

We analyzed prospectively collected longitudinal cohort data from patients attending the clinic. Clinical data from patient records were captured onto an electronic database via a medical management software system, Therapy Edge-HIV™ (Associated Biological Systems, South Africa). Data was analyzed using the SAS® 9.1 statistical software package (SAS Institute, Inc., North Carolina, USA). Baseline characteristics of the study sample were summarized using simple proportions and medians with interquartile ranges. Differences in proportions of the toxicities were compared by initiating regimen with Chi-squared tests. Study subjects were followed from HAART initiation to the earliest of 1) death; 2) loss to follow up; 3) development of toxicity or 4) censor date 31 December 2008. Incidence rates with 95% confidence intervals for toxicity were calculated and compared by initiating regimen type ( stavudine-based versus other). Crude and adjusted estimates of
the effect of stavudine use on development of incident toxicity were estimated using Cox proportional hazard models. Models were controlled for confounding by baseline body mass index, CD4 count, age, and gender. Though the majority of subjects initiated HAART prior to the introduction of the universal 30 mg stavudine dose, models were also adjusted for time period in which HAART was initiated (prior to or post October 2007). Kaplan Meier curves were used to estimate crude time to diagnosis of therapy-related complications stratified by initiated regimen. Subjects with existing toxicity at initiation of HAART were excluded from these analyses.

Use of the data for the study was approved by the Human Research Ethics Committee (Medical) of the University of the Witwatersrand.

Results
Between 1 April 2004 and 1 July 2008, a total of 15,928 HIV-infected adults enrolled in care at the Themba Lethu Clinic. Of these, 5194 (32%) had early stage HIV infection and did not qualify for HAART, whilst the remaining 10,824 (68%) subjects were initiated on HAART. The study sample included the 9040 patients initiated on treatment between 1 April 2004 and 31 December 2007. The cohort profile is summarized in Figure 1.

Baseline characteristics of the study population
The baseline characteristics of the study cohort are summarized in Table 1. Two thirds of the study group was female. The majority of patients were in the age group 25–44 years. Even though 46.9% of the total study population were defined as being at only WHO Stage 1 for AIDS, they were already on HAART, indicating that their CD4 count rates were below 200 cells/mm$^3$ and this was confirmed by the median baseline CD4 count of 81 cells/mm$^3$ (IQR: 29-149).

Ninety four percent of patients were on stavudine based regimens at baseline, with 79% of these initiated on stavudine, lamivudine and efavirenz as per the 2004 South African National guidelines [10], while a smaller number were initiated on zidovudine- (3.4%) and tenofovir-based regimens (0.7%).

Retention in care
The median time to follow up for this cohort on HAART was 19 months (IQR: 9.1-31.6). At the end of the study period, a total of 6415 patients (71%) were still alive and in care, 469 were confirmed deceased and a further 2156 were considered lost to follow up. Patients were considered lost to follow up if they missed their last scheduled appointment by more than 90 days or at least 180 days had lapsed since their last visit. It can be assumed that some of the patients who are lost to follow up may have died and so in total, there were 2,625 (16.5%) patients in this cohort who were either lost to follow up or dead by the end of the study period (see Figure 1).

Response to HAART
The patients had a median baseline CD4 count of 81 cells/mm$^3$ (IQR 29-149). There was a significant (p < 0.0001) increase in the CD4 count after initiation with a median of 265 cells/mm$^3$ (IQR 132-293) by 6 months on treatment. After six months of therapy, there was no significant change in BMI (means ± SD; 22.4 ± 5.0 and 23.7 ± 5.5 respectively).

Acute and chronic toxicities
Amongst the 9040 patients on therapy, 2488 patients (27.5%) had one or more incident toxicities recorded after treatment initiation. In terms of acute HAART-related toxicities, the proportion of patients diagnosed with peripheral neuropathy was significantly higher in the group receiving stavudine-based therapy (17.1% vs. 11.2%, p < 0.001) compared to those on non-stavudine based therapy (Table 2). Peripheral neuropathy was reported equally in both genders (approximately 17%), with no difference in time to development between the drug groups. There was also a significantly higher proportion of patients on stavudine based therapy presenting with symptomatic hyperlactataemia when compared to those on non-stavudine based therapy (5.75 vs. 2.2%, p < 0.0005) (Table 2), with females more frequently affected than males (7.1% vs. 2.5%; p < 0.0001). Although the rate of development of lactic acidosis was the same for both drug groups, it was experienced more frequently in females than males (3.3% vs. 0.8%; p < 0.0001). The median time to onset was the same for both drug groups. Pancreatitis was equally rare in both treatment groups (see Table 2); both genders were equally affected, with the time to onset being the same in both drug groups. When compared to those on non-stavudine based regimens, those receiving stavudine had higher incidence rates of several toxicities including peripheral neuropathy [12.1/100 person-years (95%CI 7.0-19.5) vs. 7.9/ 100 person-years (95%CI 6.0-10.1)], symptomatic hyperlactataemia [3.6/100 person-years (95%CI 1.2-7.5) vs. 1.4/ 100 person-years (95%CI 0.7-2.5)], and lactic acidosis [1.6/ 100 person-years (95%CI 0.4-5.2) vs. 0.8/100 person-years (95%CI 0.3-1.7)] (Table 3).

In terms of the chronic HAART-related metabolic complications, 7.3% presented with lipatrophy on stavudine based therapy, compared to 4.6% (p < 0.05) patients on non-stavudine based therapy. The median time to development of lipatrophy was similar across the 2 treatment groups. Lipatrophy was more predominantly seen in female than male patients (10.0% vs. 1.6%; p < 0.0001). Incidence rates for lipatrophy were slightly higher in the stavudine based therapy group [3.0/100 person-years (95% CI 1.9-4.4) versus 4.6/100 person-years (95%CI 2.1-6.9)].
Figure 1 Profile of the Study Cohort.

compared to those on other regimens (Table 3). Only two percent of patients presented with hypertension, while only 0.3% presented with diabetes and 1.3% with dyslipidaemia on stavudine based therapy. Similar proportions developing these conditions were observed in both groups (Table 2). However, hypertension developed more quickly (p < 0.05) in patients taking stavudine than in those not receiving this drug (Table 2). Hypertension was less common in females than males (1.8% vs. 2.4%; p < 0.05) but females and males were equally affected in terms of diabetes and dyslipidaemia.

In multivariate analyses adjusted for age, gender, baseline BMI, CD4 counts and the time period initiating HAART, the use of stavudine continued to be associated
Table 1 Cohort characteristics at baseline

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Baseline value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td>(N = 9040)</td>
</tr>
<tr>
<td>Female</td>
<td>5962 (66)</td>
</tr>
<tr>
<td>Male</td>
<td>3078 (34)</td>
</tr>
<tr>
<td>Age (years)</td>
<td>(N = 9040)</td>
</tr>
<tr>
<td>&lt; 25 years</td>
<td>551 (6.1)</td>
</tr>
<tr>
<td>25-34 years</td>
<td>3622 (40.3)</td>
</tr>
<tr>
<td>35-44 years</td>
<td>3216 (35.5)</td>
</tr>
<tr>
<td>45-54 years</td>
<td>1173 (13.0)</td>
</tr>
<tr>
<td>&gt; 55 years</td>
<td>277 (3.1)</td>
</tr>
<tr>
<td>WHO AIDS classification</td>
<td>(N = 8714)</td>
</tr>
<tr>
<td>Stage 1</td>
<td>4086 (46.9)</td>
</tr>
<tr>
<td>Stage 2</td>
<td>1310 (15)</td>
</tr>
<tr>
<td>Stage 3</td>
<td>2507 (28.8)</td>
</tr>
<tr>
<td>Stage 4</td>
<td>811 (9.3)</td>
</tr>
<tr>
<td>HAART</td>
<td>(N = 9040)</td>
</tr>
<tr>
<td>d4T/3TC/EFV</td>
<td>7138 (79)</td>
</tr>
<tr>
<td>d4T/3TC/NVP</td>
<td>690 (7.6)</td>
</tr>
<tr>
<td>d4T/3TC/Kaletra</td>
<td>669 (7.4)</td>
</tr>
<tr>
<td>AZT containing regimen</td>
<td>308 (3.4)</td>
</tr>
<tr>
<td>TDF containing regimen</td>
<td>59 (0.7)</td>
</tr>
<tr>
<td>other</td>
<td>176 (1.9)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>22.4 ± 5.0 (N = 7010)</td>
</tr>
<tr>
<td>CD4 count (cells/mm³)</td>
<td>81 (29-149) (N = 7605)</td>
</tr>
<tr>
<td>Follow up time on HAART (months)</td>
<td>190 (51-316) (N = 9040)</td>
</tr>
</tbody>
</table>

Data is expressed as N (%) except for BMI (mean ± SD); CD4 count (median, IQR) and follow up time on HAART (median, IQR). WHO, World Health Organization; d4T, stavudine; 3TC, lamivudine; EFV, etravirine; NVP, nevirapine; Kaletra, ritonavir/lopinavir; AZT, zidovudine; TDF, tenofovir.

Table 2 Frequency of and time to HAART associated toxic complications by initiating regimen

<table>
<thead>
<tr>
<th>Condition</th>
<th>Stavudine (N = 8497)</th>
<th>Other drugs (N = 543)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peripheral neuropathy</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>1545 (17.1)*</td>
<td>61 (1.2)*</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>6.7 (3.8-11.8)</td>
<td>5.1 (3.1-11.7)</td>
</tr>
<tr>
<td>Symptomatic hyperlactataemia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>487 (5.7)*</td>
<td>12 (0.2)*</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>145 (10.6-24.9)</td>
<td>116 (8.7-19.4)</td>
</tr>
<tr>
<td>Lactic Acidosis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>214 (2.5)</td>
<td>7 (0.1)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>108 (9.0-13.5)</td>
<td>112 (9.0-13.5)</td>
</tr>
<tr>
<td>Pancreatitis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>14 (0.2)</td>
<td>1 (0.0)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>10 (4.6-13.0)</td>
<td>8.3 (8.3-8.3)</td>
</tr>
<tr>
<td>Lipoatrophy</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>161 (1.9)</td>
<td>25 (0.4)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>170 (11.4-23.1)</td>
<td>150 (10.5-24.8)</td>
</tr>
<tr>
<td>Hypertension</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>167 (2.0)</td>
<td>17 (0.3)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>97 (4.6-18.4)**</td>
<td>163 (9.3-30.5)**</td>
</tr>
<tr>
<td>Diabetes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>28 (0.3)</td>
<td>1 (0.1)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>224 (12.5-28.1)</td>
<td>90 (9.0-9.0)</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>109 (1.3)</td>
<td>9 (0.1)</td>
</tr>
<tr>
<td>Time to diagnosis</td>
<td>246 (17.4-34.0)</td>
<td>191 (15.3-26.1)</td>
</tr>
</tbody>
</table>

Data is given in n (%) for prevalence and median (IQR) for time to diagnosis. *p < 0.01; **p < 0.05 versus group receiving stavudine.

with increased hazard of developing several toxicities: peripheral neuropathy (HR 2.02; 95% CI 1.35-3.03), symptomatic hyperlactataemia (HR 2.81; 95% CI 1.26-6.31), lactic acidosis (adjusted HR 2.55; 95% CI 0.81-8.00) and diabetes mellitus (adjusted HR 2.07; 95% CI 0.28-15.21) though some of these estimates lacked precision (Table 3).

Figures 2, 3, 4 and 5 present Kaplan Meier curves for the crude estimates of the time to development of HAART-related toxicities by initiating regimen. Those initiated on stavudine based regimens were more likely to develop peripheral neuropathy (log rank p < 0.001), hyperlactataemia (log rank p < 0.001), lactic acidosis (log rank p = 0.098) or lipoatrophy (log rank p < 0.05) than those on non-stavudine based regimens.

Other risk factors for development of toxicity

Age: CD4 counts and WHO staging at baseline did not appear to increase the risk of any adverse events after HAART initiation; however, BMIs at baseline did significantly increase the risk of both symptomatic hyperlactataemia and lactic acidosis. Relative to a baseline BMI of < 25 kg/m², an increased risk of symptomatic hyperlactataemia was seen for those with a BMI of 25-30 kg/m² (RR 1.7; 95% CI 1.34-2.14, p < 0.0001) and those with a BMI of > 30 kg/m² (RR 1.82; 95% CI 1.35-2.44, p < 0.0001). The same was seen with lactic acidosis, where an increased risk was seen for those with a BMI of 25-30 kg/m² (RR 2.52; 95% CI 1.74-3.65, p < 0.0001) and those with a BMI of > 30 kg/m² (RR 3.10; 95% CI 2.00-4.79, p < 0.0001) when compared to subjects with a BMI of < 25 kg/m².

Discussion

This study describes data of three years of follow up of 9040 HIV-infected adults initiated on anti-retroviral treatment at the Thembu Lethu Clinic, Johannesburg, South Africa. This high number of patients demonstrates the ability of rolling out a successful HAART programme despite being in a resource limited environment, and this, at a rapidity and scale that compares with reports from other African countries [6,11,12].

Despite the fact that stavudine is associated with significant complications, a large proportion of low and middle-income countries still use stavudine based HAART as first line therapy [8,9], mainly because of the cost implications of alternative drugs. In the present study nearly 30% of patients had to switch to non-stavudine based regimens, due to major side effects. These findings concur with those from another large study in South Africa where 21% of patients switched regimens over a similar time period [6].

This study provides estimates of the pattern of toxicities, the predominant ones being peripheral neuropathy, symptomatic hyperlactataemia, and lipoatrophy. Our incidence
Table 3 Crude and adjusted effects of stavudine use on toxicity initiated on HAART

<table>
<thead>
<tr>
<th>Toxicity</th>
<th>No events</th>
<th>Person Time (years)</th>
<th>Rate/100 pys* (95% CI)**</th>
<th>Crude HR† (95% CI)**</th>
<th>Adjusted† HR (95% CI)**</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peripheral Neuropathy</td>
<td>61</td>
<td>775.9</td>
<td>7.9 (6.0-10.1)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Other</td>
<td>1454</td>
<td>12055.5</td>
<td>12.1 (7.0-19.5)</td>
<td>1.53 (1.19-1.98)</td>
<td>2.02 (1.35-3.03)</td>
</tr>
<tr>
<td>Symptomatic hyperlactataemia</td>
<td>12</td>
<td>852.3</td>
<td>14.0 (7.2-25.1)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Other</td>
<td>487</td>
<td>13690.9</td>
<td>36.1 (12.7-75.1)</td>
<td>2.70 (1.52-4.79)</td>
<td>2.81 (1.26-6.31)</td>
</tr>
<tr>
<td>Lactic acidosis</td>
<td>7</td>
<td>8483</td>
<td>0.8 (0.3-1.7)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Other</td>
<td>214</td>
<td>13805.9</td>
<td>16.4 (5.4-52.5)</td>
<td>2.09 (0.99-4.44)</td>
<td>2.55 (0.87-8.00)</td>
</tr>
<tr>
<td>Pancreatitis</td>
<td>1</td>
<td>858.9</td>
<td>0.1 (0.003-0.6)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Other</td>
<td>14</td>
<td>14126.2</td>
<td>0.1 (0.02-0.26)</td>
<td>0.95 (0.13-7.21)</td>
<td>0.40 (0.05-3.16)</td>
</tr>
<tr>
<td>Lipatrophy</td>
<td>25</td>
<td>854.9</td>
<td>30.1 (1.9-446.1)</td>
<td>1.67 (1.12-2.50)</td>
<td>1.69 (0.92-2.77)</td>
</tr>
<tr>
<td>Other</td>
<td>616</td>
<td>13334.3</td>
<td>46.2 (21.96)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Hypertension</td>
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<td>845.0</td>
<td>20.2 (1.2-32.2)</td>
<td>0.66 (0.40-1.08)</td>
<td>0.83 (0.39-1.78)</td>
</tr>
<tr>
<td>Other</td>
<td>167</td>
<td>13950.0</td>
<td>1.2 (0.24-0.40)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Diabetes Mellitus</td>
<td>1</td>
<td>859.0</td>
<td>0.1 (0.003-0.6)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Other</td>
<td>28</td>
<td>14122.7</td>
<td>0.2 (0.02-0.26)</td>
<td>1.0</td>
<td>2.07 (0.28-15.21)</td>
</tr>
<tr>
<td>Dyslipidaemia</td>
<td>9</td>
<td>856.5</td>
<td>1.1 (0.4-2.2)</td>
<td>1.0</td>
<td>1.0</td>
</tr>
<tr>
<td>Other</td>
<td>109</td>
<td>14097.2</td>
<td>0.8 (0.2-4.0)</td>
<td>0.62 (0.41-1.61)</td>
<td>0.61 (0.27-1.40)</td>
</tr>
</tbody>
</table>

* pys = person years.
† HR = hazard ratio estimated from Cox proportional hazard models.
** 95% CI = 95% confidence interval.
†† All models adjusted for age, gender, baseline CD4 count, baseline body mass index and time at which HAART was initiated (either prior to or post October 2007).

The rates of peripheral neuropathy (12.1/100 person-years) were much higher compared to other African studies: 5.2/100 person-years in Rwanda [13] and 2.8/100 person-years in another site in South Africa [6]. The variability in these rates could be because there are no grading protocols for the severity of peripheral neuropathies. Our incidence rates of lactic acidosis were similar to a study from another province in South Africa, 1.6 versus 1.9/100 person-years [14]. The proportion of lactic acidosis was slightly higher compared to another study from Botswana [15], 2.5% versus 1% in our cohort. When compared to this same study [15], the proportion developing symptomatic hyperlactataemia were much higher in our sample (5.7% versus 2%). The median time to the development of lactic acidosis was a little later in our study when compared to another study from South Africa (10.8 months versus 7.5 months) [14].

In terms of the chronic toxicities, incidence rates of lipatrophy were 4.6/100 person-years on stavudine based therapy and 3.0/100 person-years on the non-stavudine based therapy. One study from Rwanda [13], where patients were also on stavudine based regimens showed a similar incidence rate of lipatrophy to that reported in the present study at 4.7/100 person-years, while one other study from South Africa [6] showed a lower incidence rate of 1.4/100 person-years. Another study from Rwanda showed a much higher proportion (34%) of patients developing lipatrophy [16]. The reason for this wide variation in lipatrophy rates specifically, is possibly due to different diagnostic criteria used for identifying cases. Thus, in the present study and in the studies showing low but similar rates [6,13], only cases that were severe enough to warrant a regimen change were noted. In the study showing a much higher proportion, milder cases of lipatrophy that did not require a regimen change, were also recorded [16]. An objective case definition of lipatrophy has been developed [17]; however, it requires access to DEXA and CT imaging technology, which is often not available in resource-limited settings. Therefore, an alternative consensus definition for the diagnosis of lipatrophy is required for such environments.

A very small number of patients presented with diabetes (0.3%) and dyslipidaemias (1.3%). However, this could be because lipid or glucose levels were only tested when clinically suspected due to cost implications, therefore probably underestimating the true prevalence of dyslipidaemia and diabetes.

The major strength of this study is the large sample size. This cohort is similar to many other resource-limited settings where there is rapid scaling-up of
Figure 2 Kaplan Meier crude estimates of time to development of peripheral neuropathy by initiating regimen (subjects with peripheral neuropathy at initiation of HAART were not included in the analysis).

Figure 3 Kaplan Meier crude estimates of time to development of hyperlactataemia by initiating regimen.
Figure 4 Kaplan Meier crude estimates of time to development of lactic acidosis by initiating regimen.

Figure 5 Kaplan Meier crude estimates of time to development of lipoatrophy by initiating regimen.
comprehensive HIV care and HAART. It allowed for a relatively long duration of follow up of up to three years and a fairly high retention rate of 70%, with all clinicians working on common protocols for defining the various HAART-associated toxicities. However, despite this, these findings must be considered in the light of potential limitations. It is possible that only severe toxicities that warranted a change in regimen may have been reported and this may have led to an underestimation of the rates of some of the HAART-related toxicities, particularly lipoatrophy. Also, plasma glucose and serum lipids levels were not routinely measured and thus the true rates for glucose intolerance, diabetes and dyslipidemia were not attained. Lower reported rates of the chronic toxicities could also be related to the rates of death (5%) and loss to follow up (24%), the majority of which occurred within the first six months on treatment as described in a previous report [18].

Conclusion

These results support the move away from stavudine based regimens towards less toxic combination regimens as advocated by the World Health Organisation [19,20]. South Africa has implemented these treatment guidelines, where tenofovir has now replaced stavudine as first line therapy. However in resource-limited countries, where stavudine is still being used because of cost implications, proper pharmacovigilance systems need to be established and alternative consensus definitions and grading protocols are required for identifying various HAART-related toxicities.

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Authors’ contributions

Study concept and design: CNM, MM, IS. Acquisition and analysis of data: MMA. Interpretation of data: CNM, MMA. Drafting of manuscript: CNM. Critical reviewers for important intellectual content: OIM, WM, NJC, IS, FJR. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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The early effects of stavudine compared with tenofovir on adipocyte gene expression, mitochondrial DNA copy number and metabolic parameters in South African HIV-infected patients: a randomized trial

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Objectives
Stavudine is being phased out because of its mitochondrial toxicity and tenofovir (TDF) is recommended as part of first-line highly active antiretroviral therapy (HAART) in South Africa. A prospective, open-label, randomized controlled trial comparing standard- and low-dose stavudine with TDF was performed to assess early differences in adipocyte mtDNA copy number, gene expression and metabolic parameters in Black South African HIV-infected patients.

Methods
Sixty patients were randomized 1:1:1 to either standard-dose (30–40 mg) or low-dose (20–30 mg) stavudine or TDF (300 mg) each combined with lamivudine and efavirenz. Subcutaneous fat biopsies were obtained at weeks 0 and 4. Adipocyte mtDNA copies/cell and gene expression were measured using quantitative polymerase chain reaction (qPCR). Markers of inflammation and lipid and glucose metabolism were also assessed.

Results
A 29% and 32% decrease in the mean mtDNA copies/cell was noted in the standard-dose (P < 0.005) and low-dose stavudine (P < 0.005) arms, respectively, when compared with TDF at 4 weeks. Nuclear respiratory factor-1 (NRF1) and mitochondrial cytochrome B (MTCTB) gene expression levels were affected by stavudine, with a significantly (P < 0.05) greater fall in expression observed with the standard, but not the low dose compared with TDF. No significant differences were observed in markers of inflammation and lipid and glucose metabolism.

Conclusions
These results demonstrate early mitochondrial depletion among Black South African patients receiving low and standard doses of stavudine, with preservation of gene expression levels, except for NRF1 and MTCTB, when compared with patients on TDF.

Keywords: gene expression, insulin resistance, lipids, mitochondria, stavudine, tenofovir

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Introduction

Despite the benefits of the South African Government’s highly active antiretroviral therapy (HAART) roll-out programme which commenced in 2004, adverse events remain an important concern for patients on HAART in this country [1–3]. Understanding the adverse effects of different therapeutic regimens, as well as the underlying molecular and biochemical factors that modulate risk, is critical to improving the management of HIV/AIDS. This is particularly important now that clinical practice is moving towards regimens that combine high levels of both tolerability and efficacy.

Until 2010, South Africa’s treatment guidelines for first-line public-sector HAART recommended stavudine with lamivudine (3TC) and either efavirenz (EFV) or nevirapine (NVP) [4]. This stavudine-based regimen, which is cheap and easy to administer in the short term, is associated with significant morbidity, particularly hyperlactataemia syndromes with long-term risks of lipoatrophy, and peripheral neuropathy [1–3]. The main pathogenic mechanism thought to contribute to the metabolic changes and organ toxicities is mitochondrial toxicity, via the inhibition of mitochondrial DNA (mtDNA) polymerase γ (POLG) and alterations in messenger RNA (mRNA) gene expression [5–8]. As a result of the side effects of stavudine, South Africa included tenofovir (TDF) in its recommended first-line public-sector HAART in 2010 [9], despite its cost implications. A study from South Africa demonstrated that the price of TDF would need to fall from US$17 to US$6.17 per month to have the same overall cost as stavudine-based therapy [10]. TDF has been shown to have a more favourable effect on lipid and mtDNA profiles compared with stavudine, with no difference in virological response [11].

While the various toxicities associated with stavudine have been well documented, what is less clear is whether the dose of stavudine plays a role in the development of these toxicities. The standard dosage for stavudine is 40 mg given twice daily, as this regimen received regulatory approval because of its efficacy. However, recent studies have suggested that reduced doses of stavudine (i.e. 20 or 30 mg twice daily) diminish its toxicity while maintaining efficacy [11–13].

It is therefore possible that, because stavudine is cheaper than TDF and its side effects can be minimized via reducing its dose, this would allow more patients to be initiated on HAART in countries where the switch away from stavudine may not have yet occurred. Notwithstanding the recent changes in country-level HAART guidelines recommending the use of non-stavudine-based therapy [14,15], several resource-limited countries have yet to phase out stavudine. According to the World Health Organization, in 2010 approximately 56% of HAART regimens within such countries still contained stavudine [16].

Therefore, we have conducted an open-label, randomized controlled trial to identify early molecular and biochemical changes and clinical outcomes for patients on either standard- or low-dose stavudine or TDF. We present here the results of 4 weeks of follow-up through which we set out to assess whether there were any differences in the effect of TDF when compared with low-dose and standard-dose stavudine on adipocyte-specific mitochondrial DNA copy number, gene expression and other metabolic parameters. To our knowledge this is the first study to analyse the mitochondrial effects of stavudine and TDF in a South African HIV-infected population.

Methods

Study population and inclusion criteria

We enrolled HIV-1-infected, treatment-naive individuals aged 18 years or older with a CD4 cell count below 200 cells/μL (consistent with the South African guidelines at the time [4]). Patients were enrolled at the Tembisa Lebu HIV Clinic at the Helen Joseph Hospital, a public-sector hospital in Johannesburg. Eligible subjects had an absolute neutrophil count (ANC) ≥ 750 cells/μL, haemoglobin ≥ 7.0 g/dL and a platelet count ≥ 50,000 platelets/μL. Other inclusion criteria included an alanine and aspartate aminotransferase measurement and an alkaline phosphatase measurement ≤ 2.5 × the upper limit of normal (ULN) and a total bilirubin measurement ≤ 2.5 × ULN. Patients needed to have a creatinine clearance of ≥ 60 mL/min using the Cockcroft-Gault formula. In addition, female participants of reproductive age had to have a negative serum or urine pregnancy test within 45 days prior to study entry. Those with reproductive potential had to be willing to use at least one reliable form of contraception.

Patients were excluded if they were breastfeeding or pregnant and if they had received any antiretroviral drugs (including occupational or sexual post exposure prophylaxis) other than single-dose nevirapine (NVP) within the past 6 months.

The study was approved by the Human Research Ethics Committee (Medical) of the University of the Witwatersrand and the South African Medicines Control Council (MCC).

Randomization and masking

Patients were randomly assigned 1:1:1 to receive 3TC and EFV in combination with (i) a standard dose of stavudine (30 mg if weight < 60 kg or 40 mg if weight ≥ 60 kg) according to the then South African guidelines [4]; (ii) a
low dose of stavudine [20 mg if weight < 60 kg or 30 mg if weight > 60 kg]; or (iii) TDF (300 mg). The definitions of the standard and low doses of stavudine are similar to those used in several other studies [12]. Coded identity numbers with the arm allocated were sealed in sequentially numbered envelopes. At enrolment, a member of the data collection team unsealed the next envelope to assign the drug arm allocated to the patient. Those analysing patient samples were masked to the assignment.

Subcutaneous fat biopsies for the determination of mitochondrial DNA copy number and gene expression

At weeks 0 and 4, subcutaneous fat biopsies from the supra-iliac region were performed under local anaesthetic on fasted subjects. Week 4 was chosen to avoid the confounding effects of diet and exercise arising with time. The adipose tissue biopsies were snap-frozen in liquid nitrogen prior to storage at -70°C to avoid risk of mtDNA depletion.

Total DNA was extracted at the same time from adipose tissue collected at weeks 0 and 4 for each patient using the QIAamp DNA Mini Kit (Qiagen, Inc., Hilden, Germany) and a modified protocol [17]. Mitochondrial and nuclear gene copy numbers were determined using a quantitative polymerase chain reaction (qPCR) approach and the number of mitochondria per cell calculated as previously described [17]. Total RNA was extracted using the RNeasy Lipid Tissue Mini Kit (Qiagen) and quantified using a NanoDrop ND-1000 Spectrophotometer (NanoDrop Technologies Inc., Wilmington, Delaware) and the integrity was assessed using the 2100 Bioanalyzer and RNA 6000 picochips (Agilent Technologies, Waldbronn, Germany). A 200 ng aliquot of total RNA was converted into cDNA using the Transcripter high-fidelity cDNA synthesis kit with random hexamers (Roche Diagnostics Ltd, Mannheim, Germany). Quantitative real-time PCR was performed, in duplicate for each sample, using the SensiMix SYBR Low-ROX kit (Bioline Ltd, Luckenwalde, Germany) on a 7500 Real-Time PCR system (Applied Biosystems, Foster City, CAI. Each 25 µl reaction mixture contained 1 x SensiMix reaction mix, 0.25 µM of each primer and 6.25 ng of cDNA. Thermocycling conditions for all primer pairs comprised an initial denaturation step of 95°C for 10 min; 45 amplification cycles of 95°C for 15 s, 60°C for 30 s and 72°C for 30 s; and a melt curve step of 95°C for 15 s and 60°C for 20 s ramping at 1°C to 95°C for 20 s. Genes assayed included the nuclear gene peroxisome proliferator-activated receptor (PPAR)-y coactivator 1α (PGC-1α), which is important for mitochondrial biogenesis, nuclear respiratory factor-1 (NRF1), which is a major respiratory gene transcription regulator, and mitochondrial transcription factor-A (TFAM), which binds to and regulates mtDNA. Other genes measured included those involved in mitochondrial energy metabolism: cytochrome c oxidase subunit III (COX3), cytochrome c oxidase subunit IV (COX4) and mitochondrial cytochrome B (MTCYB). Two nuclear genes involved in lipid metabolism, leptin (LEP) and lipoprotein lipase (LPL), were also assayed. Primer sequences for these genes were as previously published [5]. Gene expression was normalized against three reference genes: β2 microglobulin (B2M) (5′-TGCTGTCCATGTATGTATCT-3′ and 5′-TC TCGCTCCCCCTCTCTAGT-3′), 60S ribosomal protein L13A (RPL13A) (5′-CTTGAGGAGAGAAGGAAAGAGA-3′ and 5′-TTGAGGACCTGTGTATTTGTCAA-3′) and hypoxanthine-guanine phosphoribosyltransferase (HPRT) (5′-TGACACTGGGAAACACATGCA-3′ and 5′-GGTCTTT TCACCAGAAAGCT-3′). Primers for reference genes were designed using Primer Quest (http://eu.idtdna.com/ Scitools/Applications/Primerquest). Comparison of gene expression between the week 0 and week 4 samples was performed using the 2-∆∆CT method. In brief, change in quantification cycle (∆Cq) values for each patient were calculated for both the genes of interest and the reference genes by subtracting the Cq value of the week 4 sample from the Cq value of the week 0 sample. The relative quantities (2^-∆∆Cq) of the genes of interest and the reference genes were calculated. To normalize to multiple reference genes, the geometric mean of the relative quantities of the reference genes was calculated [18]. The normalized relative quantity of each gene of interest was determined by dividing the relative quantity of the gene of interest by this geometric mean. These normalized relative quantities were log-transformed to allow for statistical analysis.

Blood measurements

After an overnight fast, serum was taken for the measurement of total cholesterol, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL) cholesterol, triglycerides and glucose (Roche Integra Analyzer 400; Roche Diagnostics), C-peptide and insulin (Immulite1000 Analyzer; Diagnostics Corp., Los Angeles, CA), and leptin, adiponectin and high sensitivity C-reactive protein (hs-CRP) [Fluorokine® Multianalyte profiling (MAP) Kit; RnD Systems, Inc., Minneapolis, MN]. A full blood count and a liver function test were performed, urea, creatinine and electrolytes were measured, and CD4 counts were determined (XL-MCL flow cytometer, Beckman Coulter, Miami, FL). Insulin resistance was assessed using the homeostasis model assessment (HOMA) method [19], and was calculated as: [fasting serum insulin (mU/L) x fasting plasma glucose (mmol/L)]/22.5.
Statistical analysis

Sample size calculations were based on the assumption that at least 70% of participants exposed to stavudine would show features of mitochondrial depletion or features of lipodystrophy [17], while the percentage of participants exposed to TDF experiencing these outcomes would be 30%. With the power of 80%, $\alpha = 0.05$, and equal sample size per group, we required 28 subjects per group. Recruitment to the entire trial was stopped early after a data safety and monitoring board (DSMB) analysis of the first 60 patients demonstrated that the group given stavudine at both standard and low doses showed a greater fall in the mean mtDNA copies/cell than those in the arm receiving TDF. Furthermore, the DSMB considered that there were compelling ethical grounds to stop the high-dose (40 mg) stavudine because the new South African guidelines were shortly to be implemented and TDF had become freely available. Accordingly, all patients on the 40 mg dose of stavudine were switched to TDF. Furthermore, all patients were still followed up until the end of the study period, which was 48 weeks.

As a result, the study may be underpowered to detect differences between the three arms, but this did not affect the differences that were observed.

Data were analysed after creating an anonymized database and calculating differences between week 0 and 4 readings, using STATISTICA® version 10 (StatSoft, Inc., Tulsa, OK). Outliers, calculated using (lower quartile – 1.5 x interquartile range; upper quartile + 1.5 x interquartile range), were excluded from the analysis. Comparisons were made both by randomization arm and by the dosage of drug given. The mitochondrial DNA copy number and gene expression data were analysed using an analysis of variance (ANOVA) for multiple comparisons. Markers of inflammation and lipid and glucose metabolism analysed with a Kruskall–Wallis test. The threshold of significance was set at $P = 0.05$.

Results

Baseline characteristics of the study population

Ninety-one HIV-infected patients were screened. Sixty patients qualified for inclusion and consented to participate in the study, and were randomized 1:1:1 to one of three treatment arms between September 2008 and December 2009. The baseline characteristics of the 60 enrolled patients are summarized in Table 1. There was a predominance of female patients (85%) in this study and 59 of 60

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard-dose stavudine (30 mg/40 mg) (n=20)</th>
<th>Low-dose stavudine (20 mg/30 mg) (n=20)</th>
<th>TDF (300 mg) (n=20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>16 (89%)</td>
<td>18 (99%)</td>
<td>17 (89%)</td>
</tr>
<tr>
<td>Male</td>
<td>4 (21%)</td>
<td>2 (10%)</td>
<td>3 (15%)</td>
</tr>
<tr>
<td>Age (years)</td>
<td>31 (11)</td>
<td>24 (12)</td>
<td>33 (10)</td>
</tr>
<tr>
<td>WHO AIDS class</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stage 1</td>
<td>14 (70%)</td>
<td>17 (89%)</td>
<td>16 (80%)</td>
</tr>
<tr>
<td>Stage 2</td>
<td>3 (15%)</td>
<td>1 (5%)</td>
<td>3 (15%)</td>
</tr>
<tr>
<td>Stage 3</td>
<td>2 (10%)</td>
<td>2 (10%)</td>
<td>1 (5%)</td>
</tr>
<tr>
<td>Stage 4</td>
<td>2 (10%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>25 (5)</td>
<td>24 (7)</td>
<td>24 (6)</td>
</tr>
<tr>
<td>CD4 count (cells/μl)</td>
<td>155 (84)</td>
<td>169 (78)</td>
<td>135 (62)</td>
</tr>
<tr>
<td>Inflammatory markers</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>hs-CRP (μg/L)</td>
<td>0.00 (0.55)</td>
<td>0.66 (0.94)</td>
<td>0.75 (0.81)</td>
</tr>
<tr>
<td>Leptin (μg/ml)</td>
<td>2.70 (15.02)</td>
<td>3.17 (4.32)</td>
<td>5.32 (8.49)</td>
</tr>
<tr>
<td>Adiponectin (μg/ml)</td>
<td>11.70 (8.58)</td>
<td>11.76 (8.43)</td>
<td>13.70 (11.82)</td>
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<tr>
<td>Markers of lipid and glucose metabolism</td>
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<td></td>
</tr>
<tr>
<td>C-peptide (nmol/ml)</td>
<td>0.40 (0.30)</td>
<td>0.50 (0.20)</td>
<td>0.40 (0.25)</td>
</tr>
<tr>
<td>Insulin (μIU/L)</td>
<td>2.80 (4.65)</td>
<td>3.75 (4.65)</td>
<td>3.90 (3.40)</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>3.30 (3.15)</td>
<td>3.40 (1.10)</td>
<td>3.80 (1.10)</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.65 (0.40)</td>
<td>0.80 (0.45)</td>
<td>0.80 (0.40)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.70 (0.35)</td>
<td>0.80 (0.20)</td>
<td>0.80 (0.30)</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>1.90 (1.05)</td>
<td>2.15 (1.00)</td>
<td>2.50 (0.70)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>3.95 (7.75)</td>
<td>4.20 (8.00)</td>
<td>4.20 (8.00)</td>
</tr>
<tr>
<td>HOMA</td>
<td>0.54 (0.96)</td>
<td>0.70 (1.05)</td>
<td>0.73 (0.80)</td>
</tr>
</tbody>
</table>

Data are expressed as median (interquartile range) except for gender and World Health Organization (WHO) stage, which are expressed as n (%). BMI, body mass index; HDL, high-density lipoprotein; HOMA, homeostasis model assessment; hs-CRP, high sensitivity C-reactive protein; LDL, low-density lipoprotein; TDF, tenofovir.

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were Black. A total of 78% were at WHO stage 1 while 3% of patients were at WHO stage 4, despite low CD4 counts. There was no statistically significant difference in the markers of inflammation and lipid and glucose metabolism between the arms at baseline.

Mitochondrial DNA copy numbers and gene expression levels in adipocytes

The effects of HAART on mtDNA copy number and gene expression were assessed in biopsy samples of subcutaneous adipose tissue (Fig. 1). There was a 29% decrease in the mean mtDNA copies/cell from baseline to 4 weeks in the standard-dose stavudine arm (30–40 mg) ($P < 0.05$), and a 32% decrease in the low-dose stavudine arm (20–30 mg) ($P < 0.005$), when compared with the TDF (300 mg) arm, which had only a 4% decrease in the mean mtDNA copies/cell.

For each individual dose of stavudine (20, 30 and 40 mg), there was also a drop in mean mtDNA copy number (22, 35 and 31%, respectively) vs. TDF (300 mg) (4%) at 4 weeks of HAART (Fig. 1). The decrease in mtDNA copy number for both the stavudine 30 and 40 mg doses was significantly higher than for TDF 300 mg ($P < 0.005$ and $P < 0.05$, respectively). The drop in mtDNA copy number with the stavudine 20 mg dose was not significant when compared with TDF ($P = 0.40$).

The relative gene expression of MTCYB was significantly lower in the standard-dose (30–40 mg) arm when compared with the TDF (300 mg) arm ($P < 0.05$), but not in the low-dose (20–30 mg) stavudine arm (Table 2). There were minimal changes in the expression of COX3 and COX4 at 4 weeks when compared to baseline for all three arms.

With respect to the expression of the nuclear genes involved in the regulation of mitochondrial transcription, there were minimal changes in PGC1 and TFAM expression at week 4 when compared with baseline for all three arms. There was a statistically significant difference in NRF1 expression in the standard-dose stavudine arm (30–40 mg) when compared with the TDF 300 mg arm ($P < 0.05$). Such a difference was not observed in the low-dose (20–30 mg) stavudine arm. Changes in the expression of LPL and LEP from baseline were not statistically significant at week 4 in any of the arms.

When the effects of the individual stavudine doses (20, 30 and 40 mg) on gene expression were compared with those of TDF, no significant differences were noted for any of the eight genes that were studied (data not shown).

Markers of inflammation and lipid and glucose metabolism

Following 4 weeks of HAART, there were no statistically significant changes in the lipid profile, glucose, insulin, C-peptide and HOMA indices when comparing the three arms and individual doses. No changes were noted in hs-CRP, leptin or adiponectin levels (Table 2).

Discussion

This randomized controlled trial was designed to evaluate the in vitro effects of TDF against those of standard- and low-dose stavudine regimens, with the reference regimen based on the Gilead 903 trial of Gallant et al. [13]. The Gilead 903 trial showed better outcomes and fewer adverse events attributed to mitochondrial toxicity in patients.
Table 2: Comparison of the three arms in terms of changes in relative gene expression in relation to housekeeping genes, and markers of inflammation and lipid and glucose metabolism parameters from baseline at week 4

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard-dose stavudine (30 mg/40 mg) (n = 20)</th>
<th>Low-dose stavudine (20 mg/30 mg) (n = 20)</th>
<th>TDF (300 mg) (n = 20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Genes involved in mitochondrial energy metabolism</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>COX4</td>
<td>-0.01 ± 0.34</td>
<td>0.04 ± 0.49</td>
<td>0.07 ± 0.54</td>
</tr>
<tr>
<td>COX5</td>
<td>-0.11 ± 0.39</td>
<td>0.01 ± 0.37</td>
<td>0.13 ± 0.34</td>
</tr>
<tr>
<td>MTCYB</td>
<td>-0.20 ± 0.43*</td>
<td>-0.11 ± 0.50</td>
<td>0.16 ± 0.42</td>
</tr>
<tr>
<td>Nuclear genes involved in the regulation of mitochondrial transcription</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NR1F</td>
<td>-0.21 ± 0.32*</td>
<td>0.00 ± 0.51</td>
<td>0.11 ± 0.26</td>
</tr>
<tr>
<td>PGC1α</td>
<td>-0.23 ± 0.75</td>
<td>-0.02 ± 0.36</td>
<td>-0.02 ± 0.57</td>
</tr>
<tr>
<td>IFAM</td>
<td>-0.35 ± 0.23</td>
<td>0.08 ± 0.26</td>
<td>0.02 ± 0.02</td>
</tr>
<tr>
<td>Genes involved in lipid metabolism</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>LEP</td>
<td>0.09 ± 0.34</td>
<td>0.09 ± 0.40</td>
<td>-0.06 ± 0.36</td>
</tr>
<tr>
<td>LPL</td>
<td>0.06 ± 0.22</td>
<td>0.08 ± 0.29</td>
<td>-0.01 ± 0.20</td>
</tr>
<tr>
<td>Inflammatory markers</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>hs-CRP (ng/mL)</td>
<td>0.22 (0.39)</td>
<td>0.15 (0.43)</td>
<td>-0.11 (0.33)</td>
</tr>
<tr>
<td>Leptin (pg/mL)</td>
<td>0.18 (2.62)</td>
<td>0.59 (2.20)</td>
<td>-0.13 (2.89)</td>
</tr>
<tr>
<td>Adiponectin (pg/mL)</td>
<td>0.86 (5.29)</td>
<td>2.12 (5.33)</td>
<td>1.53 (6.52)</td>
</tr>
<tr>
<td>Markers of lipid and glucose metabolism</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C-peptide (mmol/L)</td>
<td>0.00 (0.20)</td>
<td>-0.10 (0.10)</td>
<td>0.00 (0.20)</td>
</tr>
<tr>
<td>Insulin (mmol/L)</td>
<td>-0.25 (0.00)</td>
<td>0.05 (4.70)</td>
<td>-0.60 (3.30)</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>0.60 (0.65)</td>
<td>0.60 (1.00)</td>
<td>0.60 (0.68)</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.10 (0.21)</td>
<td>0.20 (0.10)</td>
<td>0.20 (0.30)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.20 (0.40)</td>
<td>0.10 (0.30)</td>
<td>0.20 (0.30)</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>0.35 (0.75)</td>
<td>0.40 (0.40)</td>
<td>0.10 (0.70)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>0.30 (0.70)</td>
<td>0.00 (1.10)</td>
<td>0.20 (1.00)</td>
</tr>
<tr>
<td>HOMA</td>
<td>-0.05 (1.12)</td>
<td>-0.08 (0.88)</td>
<td>-0.17 (0.69)</td>
</tr>
</tbody>
</table>

Data are expressed as means ± standard deviation except for markers of inflammation and lipid and glucose metabolism, which are expressed as median (interquartile range); *P<0.05 vs. TDF (300 mg).

receiving TDF 300 mg when compared with stavudine 40 mg given twice daily [13], but did not report mtDNA and gene expression evaluations. Our study is the first to compare mitochondrial effects of these drugs in a Black population and at a time-point in HAART therapy before the development of peripheral lipatrophy.

The major finding in our study is the significant depletion, after only 4 weeks of therapy, of adipocyte mtDNA copy number in Black South African HIV-infected individuals receiving stavudine. No effect of TDF on mtDNA copy number was observed. This study shows that the low-dose stavudine regimen produced a fall in mitochondrial DNA copy number similar to that observed with the standard stavudine dose but greater than that seen with TDF. This finding is consistent with the results of other studies performed in non-Black populations who had developed peripheral lipatrophy at the time of sampling and after a longer duration of antiretroviral therapy [17,20–25]. Our results therefore suggest that mitochondrial pathology precedes changes in body fat distribution and occurs very early after the initiation of HAART.

This study also provides evidence of a dose-response effect in that higher doses of stavudine caused a greater loss of mtDNA after 4 weeks of HAART. Thus, stavudine at doses of 30 and 40 mg produced a greater fall in mtDNA than TDF, whereas the 20 mg dose of stavudine was associated with a nonsignificant decrease in mtDNA. When compared with the TDF arm. Despite the significant depletion in mtDNA, our study did not demonstrate a significant effect of stavudine with regard to the expression of genes associated with mitochondrial energy metabolism and biogenesis, apart from MTCYB and NRF1. This is similar to the findings of other studies [6,0,25], where only minimal changes were reported at the gene expression level. However, these findings are in contrast to those of Mallon et al., who found significant reductions in gene expression without significant depletion of mtDNA after 2 weeks of HAART, although, notably, they compared stavudine with zidovudine (ZDV) and their study was in HIV-negative patients [5]. Significant changes in the expression levels of NRF1 and MTCYB were observed in the subjects given the standard stavudine dose but not in those receiving the low-dose therapy. NRF1 is an important regulator of mitochondrial biogenesis and function. A master transcription factor, NRF1 binds the regulatory regions of genes encoding subunits of the respiratory complex and various
constituents of the mtDNA transcription and replication machinery. In addition, it serves to coordinate nucleo-mitochondrial activities by regulating mitochondrial and cytosolic enzymes [26]. These results suggest that down-regulation of *NRF1* is an early event in nucleoside reverse transcriptase inhibitor (NRTI) toxicity. This change in *NRF1* expression is then cascaded to the factors that are under its control, with effects that are only observed much later in the time-course of HAART [7,8,25].

Similar to our results, changes in MTCYB expression have been observed in other studies [6,24]. These changes may be associated with increased oxidative stress and apoptosis related to mitochondrial depletion [6,27].

Our findings must be considered in light of the study’s limitations. Firstly, the study was stopped prematurely because of the introduction of the new South African National Treatment Guidelines, which may have reduced the statistical power of the study. Our participant sample size, however, was much larger than those of other studies [1,5,6], and was still sufficient for us to be able to detect significant effects of HAART on mtDNA copy number and gene expression. Another possible limitation is the short duration of therapy (4 weeks) at the time of sampling. Other studies have shown significant mtDNA and mRNA depletion in HIV-infected patients in a longer duration of antiretroviral therapy of months to years [7,8]. In this study, the 4th week was chosen to avoid the potential confounding effects of changes in diet, exercise or body weight.

Despite these limitations, our findings are strengthened by the fact that the laboratory technique used to determine mtDNA copy number was based on a well-established protocol [17] that has been standardized and extensively critiqued [28]. Another important strength of this study was the adoption of the minimum information for publication of quantitative real-time PCR experiments (MIQE) guidelines [29] with the normalization of gene expression to multiple reference genes [18].

The findings of this study are important as previous trials in Europe, the USA and Australia did not examine the effects of mitochondrial changes in an African population [5–8,11,17,20–25]. Our study is the first randomized trial assessing the mitochondrial pathogenesis of NRTI-associated toxicities in an African population.

In conclusion, this study demonstrates an early association between mitochondrial depletion and stavudine therapy in the Black South African population and shows that TDF has a minimal effect on mitochondrial numbers. The expression levels of only two of eight adipocyte genes were significantly affected by stavudine therapy when compared with TDF, and this only with the standard dose (30 and 40 mg). These results support the new South African HAART guidelines which have evolved since 2004, with TDF 300 mg currently being recommended as first-line therapy [9]. However, minimal effects on gene expression were noted with low-dose (20–30 mg) stavudine. Therefore, further larger studies using this regimen should be initiated, particularly in light of data showing that this dose is effective and safe [11–13] and, furthermore, it is unlikely that stavudine will ever be completely phased out of use in countries with limited financial resources.

Acknowledgements

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Conflicts of interest: No conflicts of interest are declared.

Authors’ contributions: Study concept and design: CNM and IS. Acquisition of data: DVA and CNM. Laboratory analysis: CD, TDP and RD. Analysis of data: CNM, RD, CD, TDP, MM, MFF, PM and NJC. Interpretation of data: CNM, RD, CD, TDP and NJC. Drafting of manuscript: CNM. Critical revisions for important intellectual content: all authors. Read and approved the final manuscript: all authors.

References


A randomized clinical trial comparing metabolic parameters after 48 weeks of standard- and low-dose stavudine therapy and tenofovir disoproxil fumarate therapy in HIV-infected South African patients

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Objectives
Low-dose stavudine therapy may have a lower toxicity profile compared with standard dose. A randomized controlled trial comparing these two doses of stavudine with tenofovir disoproxil fumarate (tenofovir DF) was performed to assess the effects on anthropometry, markers of inflammation, and lipid and glucose metabolism in Black South African patients.

Methods
Sixty patients were randomized 1:1:1 to either standard-dose (30–40 mg) or low-dose (20–30 mg) stavudine or tenofovir DF (300 mg), each combined with lamivudine and efavirenz, for 48 weeks. Anthropometry, markers of inflammation, and lipid and glucose metabolism were assessed using standard techniques.

Results
In all three treatment arms, there was a significant increase in lipid levels over the study period. At 48 weeks, fasting glucose level ($P < 0.005$) and homeostasis model assessment (HOMA) score ($P < 0.05$) increased significantly in the standard-dose stavudine arm, as did insulin and C-peptide levels in both the standard- and low-dose stavudine arms. At week 48, a significant decrease ($P < 0.05$) in adiponectin was noted in the standard-dose stavudine arm, but there was an increase ($P < 0.005$) in the tenofovir DF arm. In both the stavudine arms, significant increases in anthropometric measures occurred at 24 weeks but these decreased by week 48. Mitochondrial toxicities occurred in both the stavudine arms. Immunological and virological outcomes were similar for all three arms.

Conclusions
This study highlights the occurrence of metabolic abnormalities with both stavudine and tenofovir DF treatment. Awareness of the potential increased cardiovascular risk should be of concern with the use of both these therapies.

Keywords: glucose, lipids, randomized trial, stavudine, tenofovir disoproxil fumarate

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Introduction

The success in controlling HIV-1 infections with highly active antiretroviral therapy (HAART) [1,2] has been marred by the emergence of metabolic complications in HIV-positive patients after long-term therapy [3–7].

An increased risk of myocardial infarction, increased rates of dyslipidemia, insulin resistance and diabetes, and alterations in body fat distribution have been described with the use of HAART [3–7]. Changes in leptin and adiponectin levels that are observed in HIV infection may play a major role in the pathogenesis of HIV/HaART-related metabolic syndrome [8,9]. Furthermore, elevated levels of high-sensitivity C-reactive protein (hs-CRP), an inflammatory marker, which have been observed in HIV-infected male patients [10], are associated with an increased risk of cardiovascular events in the general population [11,12] and in HIV-positive patients [13].

Previously, the aetiology of these metabolic abnormalities has been attributed to the use of protease inhibitors (PIs) [4]. More recently, nucleoside reverse transcriptase inhibitors (NRTIs) have been implicated, particularly stavudine [3,14–16]. Stavudine continues to be used as an alternative first-line treatment in developing countries despite the change in the World Health Organization (WHO) antiretroviral therapy guidelines for treating patients with HIV infection [17]. Stavudine was used as first-line HIV therapy in South Africa until 2010, when it was replaced with tenofovir disoproxil fumarate (tenofovir DF) [18,19], but it continues to be used because of shortages of tenofovir DF and abacavir that occur intermittently at health facilities across South Africa [20].

In the Gilead 903 trial [21], no major differences were noted between stavudine and tenofovir DF with regard to serious adverse events (AEs). Other studies have shown modest rates of nephrotoxicity with tenofovir DF [22–25], with one study finding significant nephrotoxicity at a yearly incidence of 1% [26].

The recommended dose for stavudine is 40 mg twice daily, but studies have shown that lower doses (20 or 30 mg twice daily) diminish toxicity while maintaining immunological efficacy [27–29]. Therefore, in resource-limited countries where the switch away from stavudine to the more expensive tenofovir DF has not yet occurred, low-dose stavudine could still be used, allowing more patients to be initiated on treatment.

To determine whether low-dose stavudine therapy was as effective and safe as tenofovir DF in a resource-poor environment, we performed an open-label, randomized controlled trial. Biochemical changes and clinical outcomes for patients on either low- or standard-dose stavudine were compared with those for patients receiving tenofovir DF.

The effects of these two drugs on adipocyte gene expression, mitochondrial DNA copy number and metabolic parameters after only 4 weeks of therapy in a cohort of Black South African HIV-positive patients were discussed in an earlier paper [30]. We present here follow-up data on anthropometry, serological markers of inflammation, and lipid and glucose metabolism measured after 48 weeks of therapy.

Methods

Study population

HIV-1-infected individuals aged 18 years or older with a CD4 cell count < 200 cells/μL were enrolled in this study at the Thembisilehla HIV Clinic at the Helen Joseph Hospital, Johannesburg, South Africa. Inclusion criteria were: an absolute neutrophil count (ANC) ≥ 750 cells/μL, a haemoglobin concentration ≥ 7.0 g/dL, a platelet count ≥ 50 000 cells/μL, a creatinine clearance of ≥ 60 mL/min calculated using the Cockcroft–Gault formula [31], and alanine and aspartate aminotransferase, alkaline phosphatase and total bilirubin concentrations of ≤ 2.5 times the upper limit of normal. Patients of reproductive age had to have a negative smear or urine pregnancy test within 45 days prior to study entry, and had to be willing to use at least one reliable form of contraception. Exclusion criteria were: breastfeeding or pregnancy, and receiving antiretroviral drugs including occupational or sexual post-exposure prophylaxis, other than single-dose nevirapine, within the previous 6 months.

Patients were seen at screening and at weeks 0, 4, 24 and 48. Routine evaluations were performed at each time-point, while markers of inflammation, and lipid and glucose metabolism were measured at weeks 0, 4 and 48. These evaluations included a review of AEs and concomitant medications, a complete or symptom-directed physical examination, height and weight measurements, and evaluations of haematology and chemistry profiles, calculated creatinine clearance, CD4 cell count, plasma HIV-1 RNA and study drug accountability. Dual-energy radiographic absorptiometry (DEXA) scanning was performed at baseline and week 48.

Approval was received from the Human Research Ethics Committee (Medical) of the University of the Witwatersrand and the SA Medicines Control Council (MCC). This study was compliant with the revised CONSORT statement guidelines (http://www.consort-statement.org).
Antiretroviral therapy

As previously described [31], patients were assigned in random 1:1:1 permuted blocks of five to receive lamivudine and efavirenz in combination with (a) standard-dose stavudine (30 mg if weight < 60 kg or 40 mg if weight > 60 kg) according to the then South African guidelines [18]; (b) low-dose stavudine (20 mg if weight < 60 kg or 30 mg if weight > 60 kg); or (c) tenofovir DF 300 mg. The definitions of the standard and low doses of stavudine are the same as those used in other studies [29]. Staff analysing patient samples were blinded to the treatment assigned to each patient.

Blood measurements

Fasting serum was taken for the measurement of total cholesterol, high-density lipoprotein (HDL) cholesterol, low-density lipoprotein (LDL) cholesterol, triglycerides and glucose (Roche Integra Analyzer 400; Roche Diagnostics, Mannheim, Germany), C-peptide and insulin (Immulite1000 Analyzer; Diagnostics Corp., Los Angeles, CA), and leptin, adiponectin and hs-CRP (Fluorokine®-Multianalyte profiling (MAP) kit; R&D Systems, Inc., Minneapolis, MN). A full blood count was performed and creatinine, liver function, viral load (Cobas Amplicor Analyzer, Roche Diagnostics, Indianapolis, IN) and CD4 count (XL-MCL flow cytometer, Beckman Coulter, Miami, FL) were measured. Insulin resistance was assessed using the homeostasis model assessment (HOMA) method [32].

Anthropometry and body composition measurements

Height and weight were measured using regularly calibrated instruments. Circumferences of the mid-arm, chest, waist, hip and mid-thigh were obtained using a Gulick II measuring tape (FitnessMart®, Gays Mills, WI). Skinfolds of the triceps, biceps, subscapular and suprailliac were measured using a Harpenden skin fold calliper (British Indicators, West Sussex, UK). For all anthropometry measurements, two independent measurements were taken. If the first two measurements were outside of the acceptable range of variance for a particular body area, then a third measurement was taken. The mean of the two closest measures was then used in analysis. Total lean and fat masses were measured with a Hologic whole-body, dual-energy radiographic absorptiometry (DEXA) scanner (Hologic Inc., Waltham, MA) and software, version 12.4.

Statistical analysis

Data that could be normalized were log-transformed to normality and analysed using parametric statistical analyses, while the remaining variables were analysed using nonparametric statistical tests. With a power of 80%, α = 0.05, and equal sample size per group, we required 28 patients per treatment arm, assuming that 70% of patients exposed to stavudine and 30% exposed to tenofovir DF would show features of mitochondrial toxicity [33]. However, the trial was stopped early after a data safety and monitoring board (DSMB) analysis of the first 60 patients in March 2010 showed that the group given stavudine at both standard and low doses had greater mitochondrial depletion than those in the arm receiving tenofovir [30]. Furthermore, the DSMB considered that there were compelling ethical grounds to stop treatment with high-dose stavudine because the new South African guidelines were shortly to be implemented [19], and tenofovir DF had become freely available. At that point, the four patients who were still on 40 mg of stavudine were switched to tenofovir. Those patients who switched therapy or who, for any other reason, had to prematurely leave the study were excluded from the final analysis.

Data were analysed using STATISTICA® version 10 (StatSoft, Inc., Tulsa, OK). Results were summarized using proportions and medians with interquartile ranges (IQRs). Comparisons were made at different time intervals within each arm using the Wilcoxon matched pairs test or Student’s paired t-test, while comparisons across arms were performed using analysis of variance (ANOVA) for baseline levels or analysis of covariance (ANCOVA), with adjustment for baseline levels.

Results

Baseline characteristics of the study population

Figure 1 shows the cohort profile: 60 eligible patients were randomized 1:1:1 to the three treatment arms between September 2008 and December 2009.

There was a predominance of female patients (85%) in this study and 58 of the 60 patients were Black (Table 1). All patients were clinically asymptomatic, with 78% of the patients being WHO stage 1.

Mitochondrial toxicity

As previously described [30], the tenofovir DF arm had a 4% decrease in the mean mtDNA copies/cell from baseline to 4 weeks, while there was a 29% decrease in the standard-dose stavudine arm (P < 0.05), and a 32% decrease in the low-dose stavudine arm (P < 0.005).

Relative gene expression was also affected, with mitochondrial cytochrome B (MTCyB) and nuclear respiratory factor-1 (NRF1) expression being significantly lower in the standard-dose stavudine arm compared with the tenofovir DF arm (P < 0.05).
Fig. 1 Profile of the trial cohort. DSMB, data safety and monitoring board.

Table 1 Patient characteristics at the commencement of highly active antiretroviral therapy (HAART)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard-dose stavudine (30/40 mg) (n = 20)</th>
<th>Low-dose stavudine (20/30 mg) (n = 20)</th>
<th>Tenofovir DF (300 mg) (n = 20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>African race</td>
<td>20 (100)</td>
<td>19 (95)</td>
<td>19 (95)</td>
</tr>
<tr>
<td>Female gender</td>
<td>16 (80)</td>
<td>18 (90)</td>
<td>17 (85)</td>
</tr>
<tr>
<td>Age (years)</td>
<td>31 (11)</td>
<td>34 (13)</td>
<td>33 (13)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>64 (17)</td>
<td>62 (17)</td>
<td>66 (19)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>25 (5)</td>
<td>24 (7)</td>
<td>24 (6)</td>
</tr>
<tr>
<td>Haemoglobin (g/dL)</td>
<td>12 (3)</td>
<td>12 (2)</td>
<td>12 (2)</td>
</tr>
<tr>
<td>GGT (IU/L)</td>
<td>24 (24)</td>
<td>30 (11)</td>
<td>22 (11)</td>
</tr>
<tr>
<td>ALT (IU/L)</td>
<td>21 (21)</td>
<td>20 (15)</td>
<td>18 (12)</td>
</tr>
<tr>
<td>Creatinine clearance (mL/min)</td>
<td>108 (17)</td>
<td>108 (25)</td>
<td>99 (40)</td>
</tr>
</tbody>
</table>

Data are expressed as median (interquartile range), except for race and gender which are expressed as n (%). Patients in the standard-dose stavudine arm were given 30 mg if weight < 60 kg or 40 mg if weight > 60 kg; patients in the low-dose stavudine arm were given 20 mg if weight < 60 kg or 30 mg if weight > 60 kg. BMI, body mass index; GGT, gamma-glutamyltransferase; ALT, alanine aminotransferase.
Table 2 Comparison of the markers of inflammation, glucose and lipid metabolism at different time intervals

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard-dose stavudine (30/40 mg)</th>
<th>Low-dose stavudine (20/30 mg)</th>
<th>Tenofovir DF (300 mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 4 48</td>
<td>0 4 48</td>
<td>0 4 48</td>
</tr>
<tr>
<td>n</td>
<td>20/20</td>
<td>20/20</td>
<td>20/20</td>
</tr>
<tr>
<td></td>
<td>0 4 48</td>
<td>0 4 48</td>
<td>0 4 48</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>3.20 4.10** 4.33*</td>
<td>3.40 4.10** 4.50***</td>
<td>3.80 4.20** 4.60***</td>
</tr>
<tr>
<td></td>
<td>(1.35) (1.35) (1.20)</td>
<td>(1.10) (1.70) (1.50)</td>
<td>(1.10) (1.10) (0.7)</td>
</tr>
<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.85 1.00* 1.31*</td>
<td>0.80 1.00*** 1.25***</td>
<td>0.80 1.10*** 1.30***</td>
</tr>
<tr>
<td></td>
<td>(0.40) (0.50) (0.55)</td>
<td>(0.45) (0.50) (0.60)</td>
<td>(0.40) (0.60) (0.40)</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>0.70 0.90** 0.95</td>
<td>0.80 0.80 1.10</td>
<td>0.80 0.90* 0.80</td>
</tr>
<tr>
<td></td>
<td>(0.35) (0.45) (0.20)</td>
<td>(0.20) (0.60) (0.30)</td>
<td>(0.30) (0.40) (0.20)</td>
</tr>
<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>1.90+ 2.00*** 2.35</td>
<td>2.15 2.40** 2.30***</td>
<td>2.50 2.80 2.85*</td>
</tr>
<tr>
<td></td>
<td>(1.00) (1.15) (1.20)</td>
<td>(1.00) (1.20) (1.20)</td>
<td>(0.70) (1.25) (0.70)</td>
</tr>
<tr>
<td>C-peptide (nmol/mL)</td>
<td>0.40 0.50 0.50*</td>
<td>0.50 0.40* 0.60*</td>
<td>0.40 0.45 0.50</td>
</tr>
<tr>
<td></td>
<td>(0.30) (0.20) (0.20)</td>
<td>(0.20) (0.25) (0.30)</td>
<td>(0.25) (0.25) (0.10)</td>
</tr>
<tr>
<td>Insulin (mU/L)</td>
<td>2.85 3.65 3.91*</td>
<td>3.75 3.10 6.25*</td>
<td>3.60 3.00 4.80</td>
</tr>
<tr>
<td></td>
<td>(4.05) (3.00) (2.60)</td>
<td>(4.65) (2.80) (5.50)</td>
<td>(3.00) (2.65) (3.90)</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>3.95 4.40 4.46*</td>
<td>4.20 4.30 4.40</td>
<td>4.20 4.35 4.60</td>
</tr>
<tr>
<td></td>
<td>(0.75) (0.40) (0.70)</td>
<td>(0.80) (0.70) (0.70)</td>
<td>(0.60) (0.70) (0.40)</td>
</tr>
<tr>
<td>HOMA</td>
<td>0.54 0.69 0.83*</td>
<td>0.7 0.51 1.25</td>
<td>0.73 0.59 0.91</td>
</tr>
<tr>
<td></td>
<td>(0.96) (0.81) (0.58)</td>
<td>(1.05) (0.58) (1.30)</td>
<td>(0.60) (0.56) (0.88)</td>
</tr>
<tr>
<td>hs-CRP (pg/mL)</td>
<td>0.80 0.84* 0.79</td>
<td>0.86 0.86 0.94</td>
<td>0.75 0.83 0.93</td>
</tr>
<tr>
<td></td>
<td>(0.55) (0.75) (0.35)</td>
<td>(0.94) (0.82) (0.53)</td>
<td>(0.61) (0.73) (0.56)</td>
</tr>
<tr>
<td>Leptin (pg/mL)</td>
<td>2.69+ 4.09 1.84</td>
<td>3.17 4.03 3.77</td>
<td>5.52 4.79 11.82*</td>
</tr>
<tr>
<td></td>
<td>(15.02) (10.08) (4.43)</td>
<td>(4.33) (6.07) (7.25)</td>
<td>(8.49) (12.92) (27.45)</td>
</tr>
<tr>
<td>Adiponectin (pg/mL)</td>
<td>11.78 13.53 7.21**</td>
<td>11.75 15.90 12.18</td>
<td>13.78 16.09 17.42**</td>
</tr>
<tr>
<td></td>
<td>(6.58) (4.08) (3.86)</td>
<td>(8.43) (11.14) (14.73)</td>
<td>(11.02) (12.47) (10.75)</td>
</tr>
<tr>
<td>CD4 count (cells/μL)</td>
<td>155 ND 285**</td>
<td>169 ND 286**</td>
<td>135 ND 293***</td>
</tr>
<tr>
<td></td>
<td>(94) (52)</td>
<td>(162)</td>
<td>(62)</td>
</tr>
<tr>
<td>Viral load (copies/mL)</td>
<td>10000 ND 12**</td>
<td>42500 ND 12**</td>
<td>18000 ND 12**</td>
</tr>
<tr>
<td></td>
<td>(10000) (50)</td>
<td>(70000)</td>
<td>(11)</td>
</tr>
<tr>
<td>Viral load &lt; 400 copies/mL [n (%)]</td>
<td>- - 7/8 (88)</td>
<td>- - 18/18 (100)</td>
<td>- - 18/18 (95)</td>
</tr>
<tr>
<td>Viral load &lt; 50 copies/mL [n (%)]</td>
<td>- - 7/8 (88)</td>
<td>- - 18/18 (100)</td>
<td>- - 16/18 (94)</td>
</tr>
</tbody>
</table>

HDL, high-density lipoprotein; HOMA, homeostasis model assessment; hs-CRP, high-sensitivity C-reactive protein; LDL, low-density lipoprotein; ND, not done.

Data are expressed as median (interquartile range), unless otherwise stated. *P < 0.05; **P < 0.005; *** P < 0.0005 vs. week 0; n P < 0.05 vs. same time-point in the tenofovir arm.

Lipid metabolism

There were significant increases in both total and HDL cholesterol levels from baseline to weeks 4 and 48 for all three arms (Table 2). Triglyceride levels increased significantly at week 4 in the standard-dose stavudine and tenofovir DF arms only. LDL cholesterol increased significantly in the low-dose stavudine arm at weeks 4 and 48, but increased only at week 4 in the standard-dose stavudine arm and at week 48 in the tenofovir arm (Table 2). The percentages of patients with hypercholesterolaemia, hypertriglyceridaemia, elevated LDL cholesterol and reduced HDL cholesterol in each arm, as defined by NCEP-ATP (National Cholesterol Education Program – Adult Treatment Panel) III guidelines [34], were calculated at week 48 and no significant differences were found.

Glucose metabolism

A decrease in C-peptide levels was noted at week 4 and a significant increase by week 48 in the low-dose stavudine arm, with a significant increase at week 48 also noted in the standard-dose stavudine arm (Table 2).

When fasting insulin levels were compared in all patients, an increase was noted at week 48 in both stavudine arms but not in the tenofovir DF arm. The change in insulin level from baseline to week 48 was significantly higher in the standard-dose stavudine arm [median (IQR) 2.3 (4.4) mmol/L] than in the tenofovir DF arm [median (IQR) 0.8 (5.1) mmol/L; P < 0.05].

When the HOMA scores were compared, a significant increase was noted at week 48 for the standard-dose stavudine arm only (Table 2).

Inflammatory markers

There was a significant increase in the level of hs-CRP at week 4 in the standard-dose stavudine arm and in lepitin levels at week 4 in the low-dose stavudine arm, and at week 48 in the tenofovir DF arm (Table 2).

A significant decrease in the adiponectin level was noted in the standard-dose stavudine arm at week 48, and an
increase at weeks 4 and 48 in the tenofovir DF arm. The week 48 adiponectin level was significantly higher in the tenofovir DF arm than in the standard-dose stavudine arm, and the change in adiponectin level from baseline to week 48 in the tenofovir DF arm [3.6 (3.5) pg/mL] was significantly greater ($P < 0.05$) than that in the standard-dose stavudine arm [-0.6 (6.8) pg/mL].

Immunological and virological efficacy

No differences were noted among treatment arms at week 24 in CD4 cell count or viral load. There was a significant increase in the CD4 cell counts in all three arms at week 48 when compared with baseline (Table 2). No significant differences were noted when the median CD4 count changes from baseline to week 48 were compared; the median (IQR) change was 111 (54) cells/µL for standard-dose stavudine, 122 (161) cells/µL for low-dose stavudine, and 131 (126) cells/µL for tenofovir DF.

Similarly, a significant decrease in viral load (<50 copies/µL) was noted in all three arms at week 48 when compared with baseline, but the number of patients who had virological suppression at week 48 did not differ significantly among treatment arms (Table 2).

An intent-to-treat (ITT) analysis was performed for CD4 count and viral load at week 48 and no significant differences were noted.

Anthropometric measurements and body composition

There was a significant increase in the body mass index (BMI) of patients in the standard-dose stavudine arm by weeks 4 and 24 and in the low-dose stavudine arm by weeks 24 and 48, although no significant changes were observed for patients in the tenofovir DF arm (Table 3).

A significant increase was noted in waist circumference at weeks 24 and 48 in both the standard- and low-dose stavudine arms but only at week 48 in the tenofovir DF arm. A significant increase in hip circumference was noted by week 24 in both stavudine arms but this was not observed with tenofovir DF. A similar trend was noted for mid-thigh, but with an increase at week 48 in the tenofovir DF arm.

With regard to skinfolds, a similar increasing trend was noted at week 24 for the two stavudine arms, with a tendency for skinfold thicknesses to fall by week 48 to levels close to or below those at baseline. In the tenofovir DF arm, there was an increase in skinfold thicknesses across the study period but changes were not statistically significant (Table 3).

Forty-seven per cent of patients in the standard-dose stavudine arm, 40% in the low-dose stavudine arm, and 55% in the tenofovir DF arm ($P = 0.637$) developed a BMI ≥ 25 kg/m² at week 48.

Safety and therapy switching

There were 14 AEs in the standard-dose stavudine arm, 17 AEs in the low-dose stavudine arm and 12 AEs in the tenofovir DF arm. There were no statistically significant changes in creatinine clearance in any of the three arms. There was one case of renal dysfunction, in the tenofovir DF arm. There was one death in the standard-dose stavudine arm caused by a high-grade B-cell lymphoproliferative disorder. One patient was lost to follow-up, one patient withdrew from the study and one patient became pregnant and was withdrawn from the study.

With regard to mitochondrial AEs, there were three patients in the standard-dose stavudine arm who developed lactic acidosis, while one patient developed hyperlactataemia. In the low-dose stavudine arm, three patients developed peripheral neuropathy, of whom one also had associated hyperlactataemia and clinical features of lipoatrophy warranting a switch of therapy.

Within the standard-dose stavudine arm, 12 patients had to switch therapy for various reasons. At week 0, these patients differed from those who did not change therapy in having higher body fat measurements and hs-CRP and leptin levels. Notably, at baseline 66.7% of patients who switched therapy were overweight/obese compared with 12.5% of patients who did not switch therapy ($P < 0.05$). Significant differences were noted between those who switched and did not switch therapy in the standard-dose stavudine arm in median suprailiac skinfold thickness [median (IQR) 43.3 (25.3)] mm, respectively; $P < 0.005$, mean per cent truncal fat (mean 32.98 ± SD 10.23 rs. 21.44 ± 9.93, respectively; $P < 0.05$), median hs-CRP [median (IQR) 0.86 (0.53) vs. 0.54 (0.64) pg/L, respectively; $P < 0.05$], and median leptin level [median (IQR) 8.74 (26.57) vs. 1.37 (1.62) pg/mL, respectively; $P < 0.005$].

Discussion

Despite the move away from stavudine towards less toxic regimens, more than 1.55 million people were still receiving stavudine therapy by the end of 2012 [17]. It is known that, although stavudine is cheaper than other antiretroviral drugs and is effective as initial therapy, the standard dose (30 or 40 mg) is associated with short- and long-term complications. However, studies have demonstrated excellent antiviral efficacy with a lower dose of 20 mg [27–29]. The Gilead 903 trial showed no major
<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Standard-dose stavudine (30/40 mg)</th>
<th>Low-dose stavudine (20/30 mg)</th>
<th>Tenofovir (300 mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>20/20</td>
<td>20/20</td>
<td>20/20</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.82 (4.62)</td>
<td>24.82 (4.62)</td>
<td>24.82 (4.62)</td>
</tr>
<tr>
<td>Mid-arm circumference (cm)</td>
<td>28.75 (5.50)</td>
<td>28.75 (5.50)</td>
<td>28.75 (5.50)</td>
</tr>
<tr>
<td>Waist circumference (cm)</td>
<td>81.75 (8.00)</td>
<td>81.75 (8.00)</td>
<td>81.75 (8.00)</td>
</tr>
<tr>
<td>Hip circumference (cm)</td>
<td>100.50 (10.00)</td>
<td>100.50 (10.00)</td>
<td>100.50 (10.00)</td>
</tr>
<tr>
<td>Mid-thigh circumference (cm)</td>
<td>57.00 (6.00)</td>
<td>57.00 (6.00)</td>
<td>57.00 (6.00)</td>
</tr>
<tr>
<td>Triceps skinfold thickness (mm)</td>
<td>43.70 (12.00)</td>
<td>43.70 (12.00)</td>
<td>43.70 (12.00)</td>
</tr>
<tr>
<td>Biceps skinfold thickness (mm)</td>
<td>16.00 (6.00)</td>
<td>16.00 (6.00)</td>
<td>16.00 (6.00)</td>
</tr>
<tr>
<td>Subscapular skinfold thickness (mm)</td>
<td>32.80 (12.00)</td>
<td>32.80 (12.00)</td>
<td>32.80 (12.00)</td>
</tr>
<tr>
<td>Suprailiac skinfold thickness (mm)</td>
<td>27.00 (12.00)</td>
<td>27.00 (12.00)</td>
<td>27.00 (12.00)</td>
</tr>
<tr>
<td>Trunk fat (g)</td>
<td>17987.70 (11903.50)</td>
<td>17987.70 (11903.50)</td>
<td>17987.70 (11903.50)</td>
</tr>
<tr>
<td>Trunk % fat</td>
<td>28.10 (19.50)</td>
<td>28.10 (19.50)</td>
<td>28.10 (19.50)</td>
</tr>
<tr>
<td>Total limb fat (g)</td>
<td>18078.10 (12921.10)</td>
<td>18078.10 (12921.10)</td>
<td>18078.10 (12921.10)</td>
</tr>
<tr>
<td>Total limb % fat</td>
<td>28.90 (21.00)</td>
<td>28.90 (21.00)</td>
<td>28.90 (21.00)</td>
</tr>
</tbody>
</table>

BMI, body mass index; ND, not done.
Data are expressed as median (IQR), unless otherwise stated. *P < 0.05; **P < 0.005; ***P < 0.0005 vs. week 0.
differences in the rate of clinical AEs between the stavudine and tenofovir DF arms, apart from lipid changes and investigator-reported lipodystrophy [21]. Generic tenofovir DF is used as non stavudine first-line therapy but is more expensive [35].

This was a prospective, open-label randomized controlled trial designed to evaluate the in vivo effects of tenofovir DF against those of standard- and low-dose stavudine regimens, with the reference regimen based on the Gilead 903 trial [21]. To our knowledge, this work presents the first trial to compare the use of these drugs in a Black South African population.

Studies in treatment-naïve HIV-infected patients have shown that peripheral fat initially increases during the first few months of HAART, but decreases linearly thereafter and truncal fat increases at around 6 months of therapy [36,37]. Similarly, in the current study significant increases in the majority of anthropometric measurements were noted in the stavudine arms by 6 months, with a drop towards 1 year of therapy, while such changes were seen in a minority of the anthropometric variables with tenofovir DF therapy. This study demonstrated that insulin and C-peptide levels increased significantly in both stavudine arms, but not with tenofovir DF therapy. However, significant rises in glucose level and HOMA score were only observed with the standard stavudine dose. Previous research showed that the severity of lipodystrophy in patients taking either stavudine or zidovudine was associated with an increased risk of insulin resistance [38].

Consistent with other studies [21,39,40], we found a significant increase in fasting lipids after 1 year of HAART, and this was noted within both stavudine arms and with tenofovir DF. Lipid abnormalities were noted in HIV-1-infected patients even before the advent of antiretroviral therapy, with cholesterol levels being lower and triglyceride levels higher in HIV-infected compared with uninfected subjects [41]. In the present study, the early increase of cholesterol levels after only 4 weeks of therapy may be explained by the ‘return to baseline’ effect of commencing antiretroviral therapy, but this cannot explain the rise in triglyceride levels. Furthermore, dyslipidaemia has been widely reported with exposure to NRTIs, particularly stavudine or zidovudine, and less so with tenofovir DF [21,36-40,42].

Inflammatory markers appear to have a role in the pathogenesis of HIV/HAART-related metabolic syndrome. The effect of HAART on leptin levels is controversial; a few studies have demonstrated that low levels of leptin are associated with lipodystrophy, while other studies have demonstrated no effect of HAART on leptin [43,44]. We noted an increase in leptin levels at week 4 for patients in the low-dose stavudine arm and at week 48 for the tenofovir DF arm. Patients on HAART, especially those with lipodystrophy, show a gradual reduction in serum adiponectin levels, which is thought to be associated with increased cardiovascular risk [45]. In our study, there was a significant decrease at week 48 in the standard-dose stavudine arm but a significant increase in the tenofovir DF arm at 4 and 48 weeks. These changes in serum adiponectin levels in the stavudine arm may be significant in terms of the ongoing risk of lipodystrophy as well as metabolic complications. hs-CRP has been shown to be a marker of an increased risk of cardiovascular events [12]. There was a tendency for hs-CRP levels to increase in all three treatment arms, but this reached statistical significance only in the standard-dose stavudine group at 4 weeks and then fell to baseline levels at 48 weeks.

The AEs associated with mitochondrial toxicities were noted only with the stavudine regimens.

Our findings must be considered in the light of the study’s limitations. First, our sample size was small, as the study had to be stopped prematurely because of the mitochondrial results [30] and following the introduction of the new South African National HAART guidelines. Secondly, despite this being a randomized controlled trial, it was open-label, which may have been a confounding factor. The metabolic abnormalities may be more prevalent with increasing duration of therapy and therefore not seen after only 48 weeks of therapy. However, these data are important as our study is the first randomized trial assessing the frequency of NRTI-associated metabolic toxicities related to the two most relevant antiretroviral agents currently used in a routine clinical setting in Black African populations.

In conclusion, the current study demonstrates that, in South African HIV-positive patients, tenofovir DF has more favourable effects on anthropometry and adipokine levels while both stavudine regimens increase fasting insulin and C-peptide levels, with the higher stavudine dose also causing increased fasting glucose levels and higher HOMA scores. However, both stavudine and tenofovir DF caused raised lipid levels. Therefore, awareness of the potential increased cardiovascular risk should be of concern with the use of both these therapies. More AEs were noted in the standard-dose compared with the low-dose stavudine arm. Therefore, where options are limited and there is an absolute need for stavudine to be used, low-dose stavudine may be a carefully considered option as it is equally efficient in viral suppression and has lower rates of complications than the standard dose.

Acknowledgements

We are grateful to the patients who participated in this study and the staff of the Themba Lethu Clinic, Clinical HIV

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HIV Medicine (2013)
References


34 Third report of the national cholesterol education program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (Adult Treatment Panel III) final report. *Circulation* 2002; 106: 3143–3241.


6.4. Appendix 4 – Ethics

UNIVERSITY OF THE WITWATERSRAND, JOHANNESBURG
Division of the Deputy Registrar (Research)

HUMAN RESEARCH ETHICS COMMITTEE (MEDICAL)
R14/49 Menezes

CLEARANCE CERTIFICATE

PROJECT
A Cohort Study: Review of the Mitochondrial Toxicities of HIV-Infected Individuals on Highly Anti-Retroviral Therapy - A SA Perspective

INVESTIGATORS
Dr CN Menezes

DEPARTMENT
Infectious Disease Unit

DATE CONSIDERED
07.06.29

DECISION OF THE COMMITTEE*
APPROVED UNCONDITIONALLY

Unless otherwise specified this ethical clearance is valid for 5 years and may be renewed upon application.

DATE 07.07.26

CHAIRPERSON (Professors PE Cleaton-Jones, A Dhai, M Vorster, C Feldman, A Woodiwiss)

*Guidelines for written ‘informed consent’ attached where applicable

cc: Supervisor : Prof FJ Raal

DECLARATION OF INVESTIGATOR(S)
To be completed in duplicate and ONE COPY returned to the Secretary at Room 10005, 10th Floor, Senate House, University.
I/We fully understand the conditions under which I am/we are authorized to carry out the abovementioned research and I/we guarantee to ensure compliance with these conditions. Should any departure to be contemplated from the research procedure as approved I/we undertake to resubmit the protocol to the Committee. I agree to a completion of a yearly progress report.

PLEASE QUOTE THE PROTOCOL NUMBER IN ALL ENQUIRIES

197
Dr CN Menezes,

Clinical HIV Research Unit
Posnet Suite 178
Private Bag x2600
Houghton
2041
Fax: 011 482 2130

Dear Dr Menezes,

PROTOCOL NO: CHRU 01

PROTOCOL TITLE: Molecular, Biochemical and Clinical Differences Between Stavudine and Tenofovir, each Combined with Lamivudine and Efavirenz, in HIV-Infected Patients - A South African Perspective

PRC REFERENCE NUMBER: 070603

***************************************************************

Please be advised that your trial application was:

APPROVED

The Expert Reviewer (s): Prof PA Cooper

Also reviewed by: Ms R Wills: Acting Chairperson Protocol Review Committee
Dr S Khan: Gauteng Department of Health
Dr ML Likibi: Gauteng Department of Health

Yours sincerely

MS RENÉ WILLS
Chairperson: Protocol Review Committee
29 August 2007

cc.
Clinical HIV Research Unit C
Mrs Mariene Nadoo
Tel 011 276 8809 Cell 064 403 2616 Fax 011 482 2130

A wholly owned subsidiary of the University of the Witwatersrand
29 August 2007

Mrs Marlene Naidoo
Regulatory Manager
Wits Clinical HIV Research Unit
Helen Joseph Hospital
Perth Road, Westdene
Fax: 011 482 2130

Dear Mrs Naidoo,

PROTOCOL: CHRU 01 - MOLECULAR, BIOCHEMICAL AND CLINICAL DIFFERENCES BETWEEN STAVUDINE AND TENOFOVIR, EACH COMBINED WITH LAMIVUDINE AND Efavirenz, IN HIV-INFECTED PATIENTS - A SOUTH AFRICAN PERSPECTIVE

ETHICS REFERENCE NO: 070603

RE: FINAL ETHICS APPROVAL

This is to certify that the above-mentioned trial was reviewed by the University of the Witwatersrand, Human Research Ethics Committee (HREC), and the Protocol Review Committee (PRC) on: 29 June 2007.

The University of the Witwatersrand, Human Research Ethics Committee Approval Granted for the above mentioned study is valid for five years. Where required by Sponsor to have approval on a more frequent basis it remains the responsibility of the Sponsor and Investigator to apply for continuing review and approval, or for the duration of the Trial.

1. THIS APPROVAL IS SUBJECT TO THE FOLLOWING PROVISOS:

* A copy of the MCC Approval and/or MCC Notification letter must be submitted to the Ethics Regulatory Office Secretariat before the study commences.

* The study is conducted according to the protocol submitted to the University of the Witwatersrand, Human Research Ethics Committee. Any amendments to the protocol must first be submitted to the Human Research Ethics Committee for approval.

* During the study, the University of the Witwatersrand, Human Research Ethics Committee is informed immediately of:
  - Any Unexpected Serious Adverse Events or Unexpected Adverse Drug Reactions, which, in the Investigator and/or the Sponsor’s opinion are suspected to be related to the study drug. (International and Local Reports).
  - Any data received during the trial which, may cast doubt on the validity of the continuation of the study.

* The University of the Witwatersrand, Human Research Ethics Committee is notified of any decision to discontinue the study and the reason stated.

* The Investigators authorized by this approval participate in this study. Additional Investigators shall be submitted to the University of the Witwatersrand, Human Research Ethics Committee for approval prior to their participation in the study.

* In the event of an authorized Investigator ceasing to participate in the study, the University of the Witwatersrand, Human Research Ethics Committee must be informed and the reason for such cessation given.

2. PRINCIPLES OF INFORMED CONSENT:
3. PROGRESS REPORTS:
* The University of the Witwatersrand, Human Research Ethics Committee requests that the MCC Progress Reports be submitted twice a year either in March and September or six monthly from start of study to the HREC Secretariat Office - 011 274 9281 and a report of the final results, at the conclusion of the study.

4. TRANSPORT AND STORAGE OF BLOOD AND TISSUE SAMPLES IN SOUTH AFRICA:
* If blood specimens are to be stored for future analysis and it is planned that such analysis will be done outside Wits then the blood must be stored at Wits with release of sub-samples ONLY once projects have been approved by the local Research Ethics Committee applicable to where the research will be done, as well as by the Wits Human Research Ethics Committee: (Medical).

5. REIMBURSEMENT TO PATIENTS FOR TRANSPORT:
* The Human Research Ethics Committee: (Medical) does not agree with the R150 reimbursements per visit as stipulated by the Medicines Control Council of SA but that reimbursement should be appropriate according to the situation.

6. GENETIC TESTING
* The Human Research Ethics Committee: Medical will not approve open-ended genetic testing as this does not fit the Human Research Ethics Committee criteria.

7. THE SUPPORTING APPROVAL DOCUMENTS ARE ATTACHED:

7.1 Ethics Approval Form signed by the Chairperson of the HREC - Kindly return the copy of the Approval Form signed by the Principal Investigator/s per fax: 011 274 9281 for our records.

7.2 Protocol Review Committee Approval Signature page signed by the Chairperson of the PRC,

7.3 List of members present at the HREC meeting held as per INDEPENDENT ETHICS COMMITTEE APPROVAL FORM 2003.

8. WE AWAIT YOUR RESPONSES AS REQUESTED:
* MCC Approval and/or Notification before the above study may commence.
* Copy of Approval Form signed by the Principal Investigator.
* Kindly forward the above to the undersigned at fax: 011 274 9281 at your earliest convenience.

The above has been noted for the Ethics Committee information and records.

KINDLY FORWARD TO THE RELEVANT INVESTIGATORS / CRA / SPONSOR / STUDY CO-ORDINATORS - WHERE APPLICABLE

Regards,

[Signature]

PROF PETER CLEATON-JONES

For and on behalf of the Human Research Ethics Committee: (Medical)
Dear Dr Menezes,

AUTHORISATION FOR THE IMPORTATION OF UNREGISTERED MEDICINE IN TERMS OF SECTION 21 OF THE MEDICINES AND RELATED SUBSTANCES CONTROL ACT, 1965 (ACT 101 OF 1965)

PRODUCT: TENOFOVIR DISOPROXIL FUMARATE, LOPINAVIR, ZIDOVUDINE, Efavirenz, Lamivudine, Stavudine.

Your application letter dated 31 Aug 2006 refers

1. RESOLUTION AND APPROVAL
   It was recently resolved by the Medicines Control Council that; the clinical trial application according to the following Protocol be approved :-
   CHRU 01 protocol version 1.0 dated 30-Aug-06
   Molecular, Biochemical and clinical differences between Stavudine and Tenofovir, each combined with Lamivudine and Efavirenz, in HIV-infected patients-A South African Perspective.

1.1 BEFORE COMMENCEMENT OF TRIAL
   Please Note: Copies of written Ethics Committee approval(s) to be submitted to MCC before the study commences

2. AUTHORISATION
   Authorisation is hereby granted for the importation and administration of a sufficient quantity, for the duration of the trial, of the unregistered medicine:
   TENOFOVIR DISOPROXIL FUMARATE, LOPINAVIR, ZIDOVUDINE, EFAVIREN, LAMIVUDINE, STAVUDINE,
   solely for the purpose of a clinical trial to be conducted by:
   Dr CN Menezes
   Dr M John
   Prof FJ Raal
   Dr IM Sarne

   Clinical HIV Research Unit, Helen Joseph Hospital
   Clinical HIV Research Unit, Helen Joseph Hospital
   Clinical HIV Research Unit, Helen Joseph Hospital
   Clinical HIV Research Unit, Helen Joseph Hospital

   Principal

3. PLEASE FORWARD
   It is a requirement that a copy of this letter be forwarded to all the relevant Trialist(r)s, including the approving Ethics Committee(s).
4. THIS AUTHORISATION IS SUBJECT TO THE FOLLOWING PROVISOS:

(a) The Council shall be informed immediately of any toxic effects or death, which may occur during the Clinical Trial and of any data received which, might cast doubt on the validity of the continuation of the Clinical Trial.

(b) The Council shall be notified of any decision to discontinue the Clinical Trial. The reason for such cancellation shall be stated.

(c) The Clinical Trial shall be conducted in accordance with the Protocol submitted to the Council. Any Amendment(s) to the Protocol shall first be submitted to the Council for approval. All Clinical Trials shall be conducted in accordance with ICH GCP Guidelines and the South African Clinical Trials Guidelines.

(d) The medicine shall be administered by or under the direction of the authorised Trialist. In the case where the Trialist permits another Medical Practitioner to administer a medicine, which is exempted from the registration for the purpose of the Trial, the Trialist shall remain responsible for any eventualities arising from such usage.

(e) Where a Trialist who is not authorised in the initial Authorisation, is requested to participate in the Clinical Trial, the Council requests that the relevant MCC Curriculum Vitae Format be completed detailing their Full Names, Address and Qualifications of the proposed Trialist (Practitioner) concerned, and be submitted to the Council for Approval.

(f) In the event of the authorised Trialist ceasing to participate in the Clinical Trial, the Council shall be informed and the reason for such cessation shall be given.

5. PROGRESS REPORTS

The Council must be furnished with signed six-monthly Progress Report from each Trialist including a report of the Final Results.

6. INFORMED CONSENT

It is a Council requirement that in all Clinical Trials the 'Principles of Informed Consent' should be adhered to. This applies to Trial Volunteers, as well as Participants (Patients). (Reference: Section 4.8 of ICH GCP Guidelines and Section 3.5 of SACT Guidelines).

PLEASE NOTE: Dispenser at site: Mrs C Barker. Ethics approval dated 29/08/2007 is noted.

Yours faithfully,

MRS NOLLINTU FUNANI
FOR AND ON BEHALF OF REGISTRAR OF MEDICINE

MCC TRIAL REFERENCE NO. 20061022